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Applicant: Novo Nordisk A/S, Novo Allé
DK-2880 Bagsværd

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Karin Schlichting
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Phytases (*myo*-inositol hexakisphosphate phosphohydrolases; EC 3.1.3.8) are
5 enzymes that hydrolyze phytate (*myo*-inositol hexakisphosphate) to *myo*-inositol and
inorganic phosphate and are known to be valuable feed additives.

A phytase was first described in rice bran in 1907 [Suzuki et al., Bull. Coll. Agr.
Tokio Imp. Univ. 7, 495 (1907)] and phytases from *Aspergillus* species in 1911 [Dox and
Golden, J. Biol. Chem. 10, 183-186 (1911)]. Phytases have also been found in wheat bran,
10 plant seeds, animal intestines and in microorganisms [Howsen and Davis, Enzyme Microb.
Technol. 5, 377-382 (1983), Lambrechts et al., Biotech. Lett. 14, 61-66 (1992), Shieh and
Ware, Appl. Microbiol. 16, 1348-1351 (1968)].

The cloning and expression of the phytase from *Aspergillus niger* (ficcum) has been
described by Van Hartingsveldt et al., in Gene, 127, 87-94 (1993) and in European Patent
15 Application, Publication No. (EP) 420 358 and from *Aspergillus niger* var. *awamori* by
Piddington et al., in Gene 133, 55-62 (1993).

Cloning, expression and purification of phytases with improved properties have been
disclosed in EP 684 313. However, since there is a still ongoing need for further improved
phytases, especially with respect to their thermostability, it is an object of the present
20 invention to provide the following process which is, however, not only applicable to
phytases.

A process for the preparation of a consensus protein, whereby such process is
characterized by the following steps:

- 25 a) at least three, preferably four amino acid sequences of a defined protein family
are aligned by any standard alignment program known in the art;
- b) amino acids at the same position according to such alignment are compared
regarding their evolutionary similarity by any standard program known in the
art, whereas the degree of similarity provided by such a program which defines
the least similarity of the amino acids that is used for the determination of an
30 amino acid of corresponding positions is set to a less stringent number and the
parameters are set in such a way that it is possible for the program to determine

- 5 from only 2 identical amino acids at a corresponding position an amino acid for the consensus protein; however, if among the compared amino acid sequences are sequences that show a much higher degree of similarity to each other than to the residual sequences, these sequences are represented by their consensus sequence determined as defined in the same way as in the present process for the consensus sequence of the consensus protein or a vote weight of 1 divided by the number of such sequences is assigned to every of those sequences;
- 10 c) in case no common amino acid at a defined position can be identified by the program, any of the amino acids of all sequences used for the comparison, preferably the most frequent amino acid of all such sequences is selected or an amino acid is selected on the basis of the consideration given in Example 2;
- d) once the consensus sequence has been defined, such sequence is back-translated into a DNA sequence, preferably using a codon frequency table of the organism in which expression should take place;
- 15 e) the DNA sequence is synthesized by methods known in the art and used either integrated into a suitable expression vector or by itself to transform an appropriate host cell;
- f) the transformed host cell is grown under suitable culture conditions and the consensus protein is isolated from the host cell or its culture medium by
- 20 methods known in the art.

In a preferred embodiment of this process step b) can also be defined as follows:

25 b) amino acids at the same position according to such an alignment are compared regarding their evolutionary similarity by any standard program known in the art, whereas the degree of similarity provided by such program is set at the lowest possible value and the amino acid which is the most similar for at least half of the sequences used for the comparison is selected for the corresponding position in the amino acid sequence of the consensus protein.

In another preferred embodiment the consensus sequence is used in order to improve

30 a specific protein. In this process first a consensus sequence is determined from a number of highly homologous sequences according to steps a), b) and c) as described above. In a second step the amino acid sequence of another protein which is homologous to the consensus sequence is compared with the consensus sequence and in a third step only those amino acid residues are replaced in the amino acid sequence of the other protein

which clearly differ from the consensus sequence of this protein family calculated under moderately stringent conditions whereas at all positions of the alignment where no preferred single amino acid can be determined under moderately stringent conditions the amino acids of the other protein remain unchanged.

5 By using this preferred embodiment the consensus sequence derived from a number of highly homologous sequences is used in order to replace only certain amino acid residues in the protein in such a manner that only those amino acid residues are replaced which clearly and unambiguously differ from the corresponding consensus sequence of this protein family which has been calculated on moderately stringent conditions. At all other
10 positions of the alignment, however, where the method of the present invention is not able to determine clearly a preferred amino acid residue under moderately stringent conditions the amino acid residues of the other protein are maintained unchanged.

A further preferred embodiment is a process wherein at first a consensus sequence is determined from homologous sequences as described above. In a second step the active
15 center of the protein comprising all amino acid residues that are involved in forming the active center is determined in the consensus sequence and in the sequence of a single homologous protein as well. The single homologous protein may have preferred properties like high specific activity or different pH dependency of enzymatic activity. In a third step some or all amino acid residues that are involved in forming the active centre of the
20 homologueous protein are inserted into the backbone of the consensus sequence. The result thereof is a chimeric protein having the active centre derived from a single protein and the backbone of the consensus sequence.

The active centre of the protein can be determined e.g. by using any analysis of the three-dimensional structure of the protein, e.g. by homology modelling on the basis of a
25 known 3D-structure of a known protein. Frequently the single homologueous protein is an enzyme.

It is furthermore an object of the present invention to provide such a process, wherein the program used for the comparison of amino acids at a defined position regarding their evolutionary similarity is the program "PRETTY". It is more specifically an
30 object of the present invention to provide such a process, wherein the defined protein family is the family of phytases, especially wherein the phytases are of fungal origin.

It is furthermore an object of the present invention to provide such processes, wherein the host cell is of eukaryotic, especially fungal, preferably *Aspergillus* or yeast, preferably *Saccharomyces* or *Hansenula* origin.

It is also an object of the present invention to provide a consensus protein obtainable preferably obtained, by such processes and specifically the consensus protein, which has the amino acid *sequences shown in Figures 2, 4 and 6* or a variant thereof. A "variant" refers in the context of the present invention to a consensus protein with amino acid sequence shown in Figure 2, 5, 7, and 8 wherein at one or more positions amino acids have been deleted, added or replaced by one or more other amino acids with the proviso that the resulting sequence provides for a protein whose basic properties like enzymatic activity (type of and specific activity), thermostability, activity in a certain pH-range (pH-stability) have not significantly been changed. "Significantly" means in this context that a man skilled in the art would say that the properties of the variant may still be different but would not be unobvious over the ones of the consensus protein with the amino acid sequence of Figure 2 itself.

A "mutein" refers in the context of the present invention to replacements of the amino acid in the amino acid sequences of the consensus proteins shown in Figure 2 which lead to consensus proteins with further improved properties e.g. activity. Such muteins can be defined and prepared on the basis of the teachings given in European Patent Application number 97810175.6, e. g. Q50L, Q50T, Q50G, Q50L-Y51N, or Q50T-Y51N. "Q50L" means in this context that at position 50 of the amino acid sequence (Figure 2) the amino acid Q has been replaced by amino acid L.

In addition, a food, feed or pharmaceutical composition comprising a consensus protein as defined above is also an object of the present invention.

In this context "at least three preferably four amino acid sequences of such defined protein family" means that three, four, five, six to 12, 20, 50 or even more sequences can be used for the alignment and the comparison to create the amino acid sequence of the consensus protein. "Sequences of a defined protein family" means that such sequences fold into a three dimensional structure, wherein the alpha-helices, the beta-sheets and beta-turns are at the same position so that such structures are, as called by the man skilled in the art, largely superimposable. Furthermore these sequences characterize proteins which show the same type of biological activity, e.g. a defined enzyme class, e.g. the phytases. As known in the art, the three dimensional structure of one of such sequences is sufficient to allow the modelling of the structure of the other sequences of such a family. An example, how this can be effected, is given in the Reference Example of the present case. "Evolutionary similarity" in the context of the present invention refers to a scheme which classifies amino acids regarding their structural similarity which allows that one amino acid can be replaced by another amino acid with a minimal influence on the overall structure, as this is done e.g.

by programs, like "PRETTY", known in the art. The phrase "the degree of similarity provided by such a program...is set to less stringent number" means in the context of the present invention that values for the parameters which determine the degree of similarity in the program used in the practice of the present invention are chosen in a way to allow the
5 program to define a common amino acid for a maximum of positions of the whole amino acid sequence, e. g. in case of the program PRETTY a value of 2 or 3 for the THRESHOLD and a value of 2 for the PLURALITY can be choosen. Furthermore, "a vote weight of one divided by the number of such sequences" means in the context of the present invention that the sequences which define a group of sequences with a higher
10 degree of similarity as the other sequences used for the determination of the consensus sequence only contribute to such determination with a factor which is equal to one divided by a number of all sequences of this group.

As mentioned before should the program not allow to select the most similar amino acid, the most frequent amino acid is selected, should the latter be impossible the man
15 skilled in the art will select an amino acid from all the sequences used for the comparison which is known in the art for its property to improve the thermostability in proteins as discussed e.g. by

Janecek, S. (1993), *Process Biochem.* 28, 435-445 or

Fersht, A. R. & Serrano, L. (1993), *Curr. Opin. Struct. Biol.* 3, 75-83.

20 Alber, T. (1989), *Annu. Rev. Biochem.* 58, 765-798 or

Matthews, B. W. (1987), *Biochemistry* 26, 6885-6888.

Matthews, B. W. (1991), *Curr. Opin. Struct. Biol.* 1, 17-21.

The stability of an enzyme is a critical factor for many industrial applications. Therefore, a lot of attempts, more or less successful, have been made to improve the
25 stability, preferably the thermostability of enzymes by rational (van den Burg *et al.*, 1998) or irrational approaches (Akanuma *et al.*, 1998). The forces influencing the thermostability of a protein are the same as those that are responsible for the proper folding of a peptide strand (hydrophobic interactions, van der Waals interactions, H-bonds, salt bridges, conformational strain (Matthews, 1993). Furthermore, as shown by Matthews *et al.* (1987),
30 the free energy of the unfolded state has also an influence on the stability of a protein. Enhancing of protein stability means to increase the number and strength of favorable interactions and to decrease the number and strength of unfavorable interactions. It has been possible to introduce disulfide linkages (Sauer *et al.*, 1986) to replace glycine with

alanine residues or to increase the proline content in order to reduce the free energy of the unfolded state (Margarit et al, 1992; Matthews, 1987a). Other groups concentrated on the importance of additional H-bonds or salt bridges for the stability of a protein (Blaber et al, 1993) or tried to fill cavities in the protein interior to increase the buried hydrophobic surface area and the van der Waals interactions (Karpusas et al, 19898). Furthermore, the stabilization of secondary structure elements, especially α -helices, for example, by improved helix capping, was also investigated (Munoz & Serrano, 1995).

However, there is no fast and promising strategy to identify amino acid replacements which will increase the stability, preferably the thermal stability of a protein. Commonly, the 3D structure of a protein is required to find locations in the molecule where an amino acid replacement possibly will stabilize the protein's folded state. Alternative ways to circumvent this problem are either to search for a homologous protein in a thermo- or hyperthermophile organism or to detect stability-increasing amino acid replacements by a random mutagenesis approach. This latter possibility succeeds in only 10^3 to 10^4 mutations and is restricted to enzymes for which a fast screening procedure is available (Arase et al, 1993; Risse et al, 1992). For all these approaches, success was variable and unpredictable and, if successful, the thermostability enhancements nearly always were rather small.

Here we present an alternative way to improve the thermostability of a protein. Imanaka et al (1986) were among the first to use the comparisons of homologous proteins to enhance the stability of a protein. They used a comparison of proteases from thermophilic with homologous ones of mesophilic organisms to enhance the stability of a mesophilic protease. Serrano et al (1993) used the comparison of the amino acid sequences of two homologous mesophilic RNases to construct a more thermostable Rnase. They mutated individually all of the residues that differ between the two and combined the mutations that increase the stability in a multiple mutant. Pantoliano et al (1989) and, in particular, Steipe et al (1994) suggested that the most frequent amino acid at every position of an alignment of homologous proteins contribute to the largest amount to the stability of a protein. Steipe et al (1994) proved this for a variable domain of an immunoglobulin, whereas Pantoliano et al (1989) looked for positions in the primary sequence of subtilisin in which the sequence of the enzyme chosen to be improved for higher stability was singularly divergent. Their approach resulted in the replacement M50F which increased the T_m of subtilisin by 1.8 °C.

Steipe et al. (1994) proved on a variable domain of immunoglobulin that it is possible to predict a stabilizing mutation with better than 60% success rate just by using a statistical method which determines the most frequent amino acid residue at a certain position of this domain. It was also suggested that this method would provide useful results

not only for stabilization of variable domains of antibodies but also for domains of other proteins. However, it was never mentioned that this method could be extended to the entire protein. Furthermore, nothing is said about the program which was used to calculate the frequency of amino acid residues at a distinct position or whether scoring matrices were
5 used as in the present case.

It is therefore an object of the present invention to provide a process for the preparation of a consensus protein comprising a process to calculate an amino acid residue for nearly all positions of a so-called consensus protein and to synthesize a complete gene from this sequence that could be expressed in a pro- or eukaryotic expression system.

10 DNA sequences of the present invention can be constructed starting from genomic or cDNA sequences coding for proteins, e.g. phytases known in the art [for sequence information see references mentioned above, e.g. EP 684 313 or sequence data bases, for example like Genbank (Intelligenetics, California, USA), European Bioinformatics Institute (Hinton Hall, Cambridge, GB), NBRF
15 (Georgetown University, Medical Centre, Washington DC, USA) and Vecbase (University of Wisconsin, Biotechnology Centre, Madison, Wisconsin, USA) or disclosed in the figures by methods of in vitro mutagenesis [see e.g. Sambrook et al., Molecular Cloning, Cold Spring Harbor Laboratory Press, New York]. A widely used strategy for "site directed mutagenesis", as originally outlined by Hurchinson and Edgell [J. Virol. 8, 181 (1971)],
20 involves the annealing of a synthetic oligonucleotide carrying the desired nucleotide substitution to a target region of a single-stranded DNA sequence wherein the mutation should be introduced [for review see Smith, Annu. Rev. Genet. 19, 423 (1985) and for improved methods see references 2-6 in Stanssen et al., Nucl. Acid Res., 17, 4441-4454 (1989)]. Another possibility of mutating a given DNA sequence which is also preferred for
25 the practice of the present invention is the mutagenesis by using the polymerase chain reaction (PCR). DNA as starting material can be isolated by methods known in the art and described e.g. in Sambrook et al. (Molecular Cloning) from the respective strains. For strain information see, e.g. EP 684 313 or any depository authority indicated below. *Aspergillus niger* [ATCC 9142], *Myceliophthora thermophila* [ATCC 48102],
30 *Talaromyces thermophilus* [ATCC 20186] and *Aspergillus fumigatus* [ATCC 34625] have been redeposited according to the conditions of the Budapest Treaty at the American Type Culture Cell Collection under the following accession numbers: ATCC 74337, ATCC 74340, ATCC 74338 and ATCC 74339, respectively. It is however, understood that DNA encoding a consensus protein in accordance with the present invention can also be
35 prepared in a synthetic manner as described, e.g. in EP 747 483 or the examples by methods known in the art.

The process of the present invention can preferably be used in order to improve the thermostability of the enzyme phytase. After having constructed different consensus phytase sequences it was possible to decide whether single amino acid replacements had a positive or a negative effect on the protein stability. It is therefore another subject of the present invention to improve the thermostability of a phytase.

In this embodiment single amino acids are changed in the sequence of the phytase by the introduction of at least one mutation selected from the group consisting of

| | |
|-------|-------|
| E58A | F54Y |
| D69K | I73V |
| D197N | K94A |
| T214L | R101A |
| E222T | N153K |
| E267D | V158I |
| R291I | A203G |
| R329H | S205G |
| S364T | V217A |
| A379K | A227V |
| G404A | V234L |
| | P238A |
| | Q277E |
| | A287H |
| | A292Q |
| | V366I |
| | A396S |
| | E415Q |
| | G437A |
| | E451R |

In the above-given mutations the number represents the position in the consensus phytase-1-sequence as given in Figure 2 and the letter before the number represents the amino acid in the phytase which is replaced by the respective amino acid behind the number. The numbers given correspond to the consensus phytase sequence or relate to a
5 corresponding residue calculated by an alignment as shown in Figure 1 when 26 amino acids (signal sequence) are added to the sequences shown in Fig. 1. Those mutations can be introduced into consensus sequences or into sequences of specific enzymes which have been improved by a process of the present invention. The above-mentioned amino acid replacements have a positive effect on the protein stability.

10 Once complete DNA sequences of the present invention have been obtained they can be integrated into vectors by methods known in the art and described e.g. in Sambrook et al. (s.a.) to overexpress the encoded polypeptide in appropriate host systems. However, a man skilled in the art knows that also the DNA sequences themselves can be used to transform the suitable host systems of the invention to get overexpression of the encoded
15 polypeptide. Appropriate host systems are for example fungi, like Aspergilli, e.g. *Aspergillus niger* [ATCC 9142] or *Aspergillus ficuum* [NRRL 3135] or like *Trichoderma*, e.g. *Trichoderma reesei* or yeasts, like *Saccharomyces*, e.g. *Saccharomyces cerevisiae* or *Pichia*, like *Pichia pastoris*, or *Hansenula polymorpha*, e.g. *H. polymorpha* (DSM5215) or plants, as described, e.g. by Pen et al., *Bio/Technology* 11, 811-814 (1994). A man skilled
20 in the art knows that such microorganisms are available from depository authorities, e.g. the American Type Culture Collection (ATCC), the Centraalbureau voor Schimmelcultures (CBS) or the Deutsche Sammlung für Mikroorganismen und Zellkulturen GmbH (DSM) or any other depository authority as listed in the Journal "Industrial Property" [(1991) 1, pages 29-40]. Bacteria which can be used are e.g. *E. coli*,
25 *Bacilli* as, e.g. *Bacillus subtilis* or *Streptomyces*, e.g. *Streptomyces lividans* (see e.g. Anné and Mallaert in *FEMS Microbiol. Letters* 114, 121 (1993). *E. coli*, which could be used are *E. coli* K12 strains e.g. M15 [described as DZ 291 by Villarejo et al. in *J. Bacteriol.* 120, 466-474 (1974)], HB 101 [ATCC No. 33694] or *E. coli* SG13009 [Gottesman et al., *J. Bacteriol.* 148, 265-273 (1981)].

30 Vectors which can be used for expression in fungi are known in the art and described e.g. in EP 420 358, or by Cullen et al. [*Bio/Technology* 5, 369-376 (1987)] or Ward in *Molecular Industrial Mycology, Systems and Applications for Filamentous Fungi*, Marcel Dekker, New York (1991), Upshall et al. [*Bio/Technology* 5, 1301-1304 (1987)] Gwynne et al. [*Bio/Technology* 5, 71-79 (1987)], Punt et al. [*J. Biotechnol.* 17, 19-34 (1991)] and
35 for yeast by Sreekrishna et al. [*J. Basic Microbiol.* 28, 265-278 (1988), *Biochemistry* 28,

4117-4125 (1989)], Hitzemann et al. [Nature 293, 717-722 (1981)] or in EP 183 070, EP 183 071, EP 248 227, EP 263 311. Suitable vectors which can be used for expression in *E. coli* are mentioned, e.g. by Sambrook et al. [s.a.] or by Fiers et al. in Proc'd. 8th Int. Biotechnology Symposium" [Soc. Franc. de Microbiol., Paris (Durand et al., eds.), pp. 680-697 (1988)] or by Bujard et al. in Methods in Enzymology, eds. Wu and Grossmann, Academic Press, Inc. Vol. 155, 416-433 (1987) and Stüber et al. in Immunological Methods, eds. Lefkovits and Pernis, Academic Press, Inc., Vol. IV, 121-152 (1990). Vectors which could be used for expression in *Bacilli* are known in the art and described, e.g. in EP 405 370, Proc'd. Natl. Acad. Sci. USA 81, 439 (1984) by Yansura and Henner, Meth. Enzymol. 185, 199-228 (1990) or EP 207 459. Vectors which can be used for the expression in *H. Polymorpha* are known in the art and described, e.g. in Gellissen et al., Biotechnology 9, 291-295 (1991).

Either such vectors already carry regulatory elements, e.g. promoters, or the DNA sequences of the present invention can be engineered to contain such elements. Suitable promoter elements which can be used are known in the art and are, e.g. for *Trichoderma reesei* the *cbh1*- [Haarki et al., Biotechnology 7, 596-600 (1989)] or the *pki1*-promotor [Schindler et al., Gene 130, 271-275 (1993)], for *Aspergillus oryzae* the *amy*-promotor [Christensen et al., Abstr. 19th Lunten Lectures on Molecular Genetics F23 (1987), Christensen et al., Biotechnology 6, 1419-1422 (1988), Tada et al., Mol. Gen. Genet. 229, 301 (1991)], for *Aspergillus niger* the *glaA*- [Cullen et al., Bio/Technology 5, 369-376 (1987), Gwynne et al., Bio/Technology 5, 713-719 (1987), Ward in Molecular Industrial Mycology, Systems and Applications for Filamentous Fungi, Marcel Dekker, New York, 83-106 (1991)], *alcA*- [Gwynne et al., Bio/Technology 5, 718-719 (1987)], *suc1*- [Boddy et al., Curr. Genet. 24, 60-66 (1993)], *aphA*- [MacRae et al., Gene 71, 339-348 (1988), MacRae et al., Gene 132, 193-198 (1993)], *tpiA*- [McKnight et al., Cell 46, 143-147 (1986), Upshall et al., Bio/Technology 5, 1301-1304 (1987)], *gpdA*- [Punt et al., Gene 69, 49-57 (1988), Punt et al., J. Biotechnol. 17, 19-37 (1991)] and the *pkiA*-promotor [de Graaff et al., Curr. Genet. 22, 21-27 (1992)]. Suitable promoter elements which could be used for expression in yeast are known in the art and are, e.g. the *pho5*-promotor [Vogel et al., Mol. Cell. Biol., 2050-2057 (1989); Rudolf and Hinnen, Proc. Natl. Acad. Sci. 84, 1340-1344 (1987)] or the *gap*-promotor for expression in *Saccharomyces cerevisiae* and for *Pichia pastoris*, e.g. the *aox1*-promotor [Koutz et al., Yeast 5, 167-177 (1989); Sreekrishna et al., J. Basic Microbiol. 28, 265-278 (1988)], or the FMD promoter [Hollenberg et al., EPA No. 0299108] or *MOX*-promotor [Ledeboer et al., Nucleic Acids Res. 13, 3063-3082 (1985)] for *H. polymorpha*.

Accordingly vectors comprising DNA sequences of the present invention, preferably for the expression of said DNA sequences in bacteria or a fungal or a yeast host and such transformed bacteria or fungal or yeast hosts are also an object of the present invention.

It is also an object of the present invention to provide a system which allows for high
5 expression of proteins, preferably phytases like the consensus phytase of the present invention in *Hansenula* characterized therein that the codons of the encoding DNA sequence of such a protein have been selected on the basis of a codon frequency table of the organism used for expression, e.g. yeast as in the present case (see e.g. in Example 3) and optionally the codons for the signal sequence have been selected in a manner as
10 described for the specific case in Example 3. That means that a codon frequency table is prepared on the basis of the codons used in the DNA sequences which encode the amino acid sequences of the defined protein family. Then the codons for the design of the DNA sequence of the signal sequence are selected from a codon frequency table of the host cell used for expression whereby always codons of comparable frequency in both tables are
15 used.

Once such DNA sequences have been expressed in an appropriate host cell in a suitable medium the encoded protein can be isolated either from the medium in the case the protein is secreted into the medium or from the host organism in case such protein is present intracellularly by methods known in the art of protein purification or described in
20 case of a phytase, e.g. in EP 420 358. Accordingly a process for the preparation of a polypeptide of the present invention characterized in that transformed bacteria or a host cell as described above is cultured under suitable culture conditions and the polypeptide is recovered therefrom and a polypeptide when produced by such a process or a polypeptide encoded by a DNA sequence of the present invention are also an object of the present
25 invention.

Once obtained the polypeptides of the present invention can be characterized regarding their properties which make them useful in agriculture any assay known in the art and described e.g. by Simons et al. [Br. J. Nutr. 64, 525-540 (1990)], Schöner et al. [J. Anim. Physiol. a. Anim. Nutr. 66, 248-255 (1991)], Vogt [Arch. Geflügelk. 56, 93-98
30 (1992)], Jongbloed et al. [J. Anim. Sci., 70, 1159-1168 (1992)], Perney et al. [Poultry Sci. 72, 2106-2114 (1993)], Farrell et al., [J. Anim. Physiol. a. Anim. Nutr. 69, 278-283 (1993)], Broz et al., [Br. Poultry Sci. 35, 273-280 (1994)] and Dünghoef et al. [Animal Feed Sci. Technol. 49, 1-10 (1994)] can be used.

In general the polypeptides of the present invention can be used without being
35 limited to a specific field of application, e.g. in case of phytases for the conversion of inositol polyphosphates, like phytate to inositol and inorganic phosphate.

Furthermore the polypeptides of the present invention can be used in a process for the preparation of a pharmaceutical composition or compound food or feeds wherein the components of such a composition are mixed with one or more polypeptides of the present invention. Accordingly compound food or feeds or pharmaceutical compositions comprising one or more polypeptides of the present invention are also an object of the present invention. A man skilled in the art is familiar with their process of preparation. Such pharmaceutical compositions or compound foods or feeds can further comprise additives or components generally used for such purpose and known in the state of the art.

It is furthermore an object of the present invention to provide a process for the reduction of levels of phytate in animal manure characterized in that an animal is fed such a feed composition in an amount effective in converting phytate contained in the feedstuff to inositol and inorganic phosphate.

Before describing the present invention in more detail a short explanation of the Figures enclosed is given below.

Figure 1: Design of the consensus phytase sequence. The letters represent the amino acid residues in the one-letter code. The following sequences were used for the alignment: *phyA* from *Aspergillus terreus* 9A-1 (Mitchell et al, 1997; from amino acid (aa) 27), *phyA* from *A. terreus* cbs1 16.46; (van Loon et al., 1998; from aa 27), *phyA* from *Aspergillus niger* var. *awamori* (Piddington et al, 1993; from aa 27), *phyA* from *A. niger* T213; from aa 27), *phyA* from *A. niger* strain NRRL3135 (van Hartingsveldt et al, 1993; from aa 27), *phyA* from *Aspergillus fumigatus* ATCC 13073 (Pasamontes et al, 1993; from aa 25), *phyA* from *A. fumigatus* ATCC 32722 (van Loon et al, 1998; from aa 27), *phyA* from *A. fumigatus* ATCC 58128 (van Loon et al., 1998; from aa 27), *phyA* from *A. fumigatus* ATCC 26906 (van Loon et al, 1998; from aa 27), *phyA* from *A. fumigatus* ATCC 32239 (van Loon et al, 1998; from aa 30), *phyA* from *Emmericella nidulans* (Pasamontes et al, 1997a; from aa 25), *phyA* from *Talaromyces thermophilus* (Pasamontes et al, 1997a; from aa 24), and *phyA* from *Myceliophthora thermophila* (Mitchell et al, 1997; from aa 19). The alignment was calculated using the program PILEUP. The location of the gaps was refined by hand. Capitalized amino acid residues in the alignment at a given position belong to the amino acid coalition that establish the consensus residue. In bold, beneath the calculated consensus sequence, the amino acid sequence of the finally constructed consensus phytase (Fcp) is shown. The gaps in the calculated consensus sequence were filled by hand according to principals stated in Example 1.

Figure 2: DNA sequence of the consensus phytase-1 gene (*fcp*) and of the primers used for the gene construction. The calculated amino acid sequence (Figure 1) was converted into a DNA sequence using the program BACKTRANSLATE (Devereux *et al.*, 1984) and the codon frequency table of highly expressed yeast genes (GCG program package, 9.0). The signal peptide of the phytase from *A. terreus* cbs.116.46 was fused to the *N*-terminus. The bold bases represent the sequences of the oligonucleotides used to generate the gene. The names of the respective oligonucleotides are alternately noted above or below the sequence. The underlined bases represent the start and stop codon of the gene. The bases written in italics show the two introduced *Eco* RI sites.

Figure 3: Alignment and consensus sequence of five *Basidiomycetes* phytases. The letters represent the amino acid residues in the one-letter code. The amino acid sequences of the phytases from *Paxillus involutus*, phyA1 (aa 21) and phyA2 (aa 21, WO 98/28409), *Trametes pubescens* (aa 24, WO 98/28409), *Agrocybe pediades* (aa 19, WO 98/28409), and *Peniophora lycii* (aa 21, WO 98/28409) starting with the amino acid residues mentioned in parentheses, were used for the alignment and the calculation of the corresponding consensus sequence called "Basidio" (Example 2). The alignment was performed by the program PILEUP. The location of the gaps was refined by hand. The consensus sequence was calculated by the program PRETTY. While a vote weight of 0.5 was assigned to the two *P. involutus* phytases, all other genes were used with a vote weight of 1.0 for the consensus sequence calculation. At positions, where the program was not able to determine a consensus residues, the Basidio sequence contains a dash. Capitalized amino acid residues in the alignment at a given position belong to the amino acid coalition that establish the consensus residue.

Figure 4: Design of consensus phytase-10 amino acid sequence. Adding the phytase sequence of *Thermomyces lanuginosa* (Berka *et al.*, 1998) and the consensus sequence of the phytases from five *Basidiomycetes* to the alignment of Figure 1, an improved consensus sequence was calculated by the program PRETTY. Additionally, the amino acid sequence of *A. niger* T213 was omitted, therefore, using a vote weight of 0.5 for the remaining *A. niger* phytase sequences. For further information see Example 2.

Figure 5: DNA and amino acid sequence of consensus phytase-10. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The sequence of the oligonucleotides which were used to assemble the gene are in bold letters. The label of oligonucleotides and the amino acids, which were changed compared to those for consensus phytase -1, are underlined and their corresponding triplets are highlighted in

small cases. The *fcp10* gene was assembled from the following oligonucleotides: CP-1, CP-2, CP-3.10, CP-4.10, CP-5.10, CP-6, CP-7.10, CP-8.10, CP-9.10, CP-10.10, CP-11.10, CP-12.10, CP-13.10, CP-14.10, CP-15.10, CP-16.10, CP-17.10, CP-18.10, CP-19.10, CP-20.10, CP-21.10, CP-22.10. The newly synthesized oligonucleotides are additionally
5 marked by number 10. The phytase contains the following 32 exchanges: Y54F, **E58A**, D69K, D70G, A94K, N134Q, I158V, S187A, Q188N, **D197N**, S204A, T214L, D220E, L234V, A238P, D246H, T251N, Y259N, **E267D**, E277Q, A283D, **R291I**, A320V, **R329H**, **S364T**, I366V, **A379K**, S396A, **G404A**, Q415E, A437G, A463E. The mutations accentuated in bold letters revealed a stabilizing effect on consensus phytase-1 as tested as
10 single mutation in consensus phytase-1.

Figure 6: Alignment for the design of consensus phytase-11. In contrast to the design of consensus phytase-10, for the design of the amino acid sequence of consensus phytase-11, all *Basidiomycetes* phytases were used as independent sequences using an assigned vote weight of 0.2 for each *Basidiomycetes* sequence. Additionally, the amino acid sequence of
15 *A. niger* T213 was used in that alignment, again.

Figure 7: DNA and amino acid sequence of consensus phytase-1-thermo[8]-Q50T-K91A. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (*).

20 Figure 8: DNA and amino acid sequence of consensus phytase-10-thermo[3]-Q50T-K91A. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (*).

Figure 9: DNA and amino acid sequence of *A. fumigatus* ATCC 13073 phytase a-mutant.
25 The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (*).

Figure 10: DNA and amino acid sequence of consensus phytase-7. The amino acids are written above the corresponding DNA sequence using the one-letter code. The sequence of
30 the oligonucleotides used to assemble the gene are in bold letters. Oligonucleotides and amino acids that were exchanged are underlined and their corresponding triplets are highlighted in small cases. The *fcp7* gene was assembled from the following

oligonucleotides: CP-1, CP-2, CP-3, CP-4.7, CP-5.7, CP-6, CP-7, CP-8.7, CP-9, CP-10.7, CP-11.7, CP-12.7, CP-13.7, CP-14.7, CP-15.7, CP-16, CP-17.7, CP-18.7, CP-19.7, CP-20, CP-21, CP-22. The newly synthesized oligonucleotides are additionally marked by number 7. The phytase contains the following 24 exchanges in comparison to the original
 5 consensus phytase: S89D, S92G, A94K, D164S, P201S, G203A, G205S, H212P, G224A, D226T, E255T, D256E, V258T, P265S, Q292H, G300K, Y305H, A314T, S364G, M365I, A397S, S398A, G404A, and A405S.

Figure 11: Differential scanning calorimetry (DSC) of consensus phytase-1 and consensus phytase-10. The protein samples were concentrated to ca. 50-60 mg/ml and extensively
 10 dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-10 (upper graph) yielded a melting temperature of 85.4 °C, which is 7.3 °C higher than the melting point of consensus phytase-1 (78.1 °C, lower graph).

Figure 12: Differential scanning calorimetry (DSC) of consensus phytase-10-thermo-Q50T
 15 and consensus phytase-10-thermo-Q50T-K91A. The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-10-thermo-Q50T (upper graph) yielded a melting temperature of 88.6 °C, while the melting point of consensus phytase-10-thermo-Q50T-K91A was found at 89.3 °C.

Figure 13: Comparison of the temperature optimum between consensus phytase-1,
 20 consensus phytase-10 and consensus phytase-10-thermo-Q50T. For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 86 °C. The diluted supernatant of transformed *S. cerevisiae* strains was used for the determination. The other components of the supernatant showed
 25 no influence on the determination of the temperature optimum: △, consensus phytase-1; ◇, consensus phytase-10; ■, consensus phytase 10-thermo-Q50T.

Figure 14: pH-dependent activity profile and substrate specificity of consensus phytase-10
 and its variants thermo-Q50T and thermo-Q50T-K91A. The phytase activity was
 determined using the standard assay in appropriate buffers (see Example 9) at different pH-
 30 values. Graph a) shows the pH-dependent activity profile of consensus phytase-10 (□), consensus phytase-10-thermo-Q50T (•), and consensus phytase-10-thermo-Q50T-K91A (△). Graph b) shows the corresponding substrate specificity tested by replacement of phytate by the indicated compounds in the standard assay; open bars, consensus phytase-10

(grey bars, consensus phytase-10-thermo-Q50T; dark bars, consensus phytase-10-thermo-Q50T-K91A). The numbers correspond to the following compounds: 1, phytate; 2, *p*-nitrophenyl phosphate; 3, phenyl phosphate; 4, fructose-1,6-bisphosphate; 5, fructose-6-phosphate; 6, glucose-6-phosphate; 7, ribose-5-phosphate; 8, DL-glycerol-3-phosphate; 9, glycerol-2-phosphate; 10, 3-phosphoglycerate; 11, phosphoenolpyruvate; 12, AMP; 13, ADP; 14, ATP.

Figure 15: pH-dependent activity profile and substrate specificity of consensus phytase-1-thermo[8]-Q50T and of consensus phytase-1-thermo[8]-Q50T-K91A. The phytase activity was determined using the standard assay in appropriate buffers (see Example 9) at different pH-values. Graph a) shows the pH-dependent activity profile of the Q50T- (■) and the Q50T-K91A-variant (•). Graph b) shows the corresponding substrate specificities tested by replacement of phytate by the indicated compounds in the standard assay (open bars, consensus phytase-1-thermo[8]-Q50T; filled bars, consensus phytase-1-thermo[8]-Q50T-K91A.). The substrates are listed in the legend of Figure 14.

Figure 16: Differential scanning calorimetry (DSC) of consensus phytase-1-thermo[8]-Q50T and consensus phytase-1-thermo[8]-Q50T-K91A. The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-1-thermo[8]-Q50T (upper graph) showed a melting temperature of 84.7 °C, while the melting point of consensus phytase-1-thermo[8]-Q50T-K91A was found at 85.7 °C.

Figure 17: Comparison of the temperature optimum between consensus phytase-1, consensus phytase-1-thermo[3] and consensus phytase-1-thermo[8]. For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 86 °C. Purified protein from the supernatant of transformed *S. cerevisiae* strains was used for the determination. O, consensus phytase-1; □, consensus phytase-1-thermo[3]; ▲, consensus phytase 1-thermo[8].

Figure 18: Comparison of the pH-dependent activity profile and substrate specificity of consensus phytase-1, consensus phytase-7, and of the phytase from *A. niger* NRRL 3135. The phytase activity was determined using the standard assay in appropriate buffers (see Example 9) at different pH-values. Graph a) shows the pH-dependent activity profile of consensus phytase-1 (■), the phytase from *A. niger* NRRL 3135 (○), and of consensus phytase-7 (▲). Graph b) shows the corresponding substrate specificity tested by replacement of phytate by the indicated compounds in the standard assay (black bars, *A.*

niger NRRL 3135 phytase; grey bars, consensus phytase-1, dashed bars, consensus phytase-7). The substrates are listed in the legend of Figure 14.

Figure 19: Differential scanning calorimetry (DSC) of the phytase from *A. fumigatus* ATCC 13073 and of its stabilized α -mutant, which contains the following amino acid exchanges F55Y, V100I, F114Y, A243L, S265P, N294D.

The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus *A. fumigatus* 13073 phytase (upper graph) revealed a melting temperature of 62.5 °C, while the melting point of the α -mutant was found at 67.0 °C

Figure 20: Comparison of the temperature optimum of *A. fumigatus* 13073 wild-type, its *A. fumigatus* α -mutant, and a further stabilized α -mutant (E59A-S126N-R329H-S364T-G404A). For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 75 °C. The diluted supernatant of transformed *S. cerevisiae* strains was used for the determination. The other components of the supernatant showed no influence on the determination of the temperature optimum.

○, *A. fumigatus* ATCC 13073 phytase; ▲, *A. fumigatus* ATCC 13073 α -mutant; □, *A. fumigatus* ATCC 13073 alpha-mutant-(E59A-S126N-R329H-S364T-G404A)-Q27T; ■, *A. fumigatus* ATCC 13073 α -mutant-(E59A-S126N-R329H-S364T-G404A)-Q27T-K68A. Q27T and K68A corresponds to consensus phytase-1 Q50T and K91A, respectively.

Figure 21: Amino acid sequence of consensus phytase 12 (consphy12) which contains a number of active site residues transferred from the "basidio" consensus sequence to consensus phytase-10-thermo-Q50T-K91A.

Example 1:

Design of the amino acid sequence of consensus phytase-1

Alignment of the amino acid sequences

The alignment was calculated using the program PILEUP from the Sequence
5 Analysis Package Release 9.0 (Devereux *et al.*, 1984) with the standard parameter (gap
creation penalty 12, gap extension penalty 4). The location of the gaps was refined using a
text editor. Table 1 shows the sequences (see Figure 1) without the signal sequence that
were used for the performance of the alignment starting with the amino acid (aa) as
mentioned in Table 1.

10 **Table 1: Origin and vote weight of the phytase amino acid sequences used for the design of
consensus phytase-1**

- *phyA* from *Aspergillus terreus* 9A-1, aa 27, vote weight 0.5 (Mitchell *et al.*, 1997)
- *phyA* from *Aspergillus terreus* cbs116.46, aa 27, vote weight 0.5 (van Loon *et al.*, 1998)
- *phyA* from *Aspergillus niger* var. *awamori*, aa 27, vote weight 0.33 (Piddington *et al.*,
15 1993)
- *phyA* from *Aspergillus niger* T213, aa 27, vote weight 0.33
- *phyA* from *Aspergillus niger* strain NRRL3135, aa 27, vote weight 0.33 (van
Hartingsveldt *et al.*, 1993)
- *phyA* from *Aspergillus fumigatus* ATCC 13073, aa 26, vote weight 0.2 (Pasamontes *et*
20 *al.*, 1997)
- *phyA* from *Aspergillus fumigatus* ATCC 32722, aa 26, vote weight 0.2 (van Loon *et al.*,
1998)
- *phyA* from *Aspergillus fumigatus* ATCC 58128, aa 26, vote weight 0.2 (van Loon *et al.*,
1998)
- 25 - *phyA* from *Aspergillus fumigatus* ATCC 26906, aa 26, vote weight 0.2 (van Loon *et al.*,
1998)
- *phyA* from *Aspergillus fumigatus* ATCC 32239, aa 30, vote weight 0.2 (van Loon *et al.*,
1998)
- *phyA* from *Emericella nidulans* , aa 25, vote weight 1.0 (Roche Nr. R1288, Pasamontes
30 *et al.*, 1997a)
- *phyA* from *Talaromyces thermophilus* ATCC 20186, aa 24, vote weight 1.0 (Pasamontes
et al., 1997a)
- *phyA* from *Myceliophthora thermophila*, aa 19, vote weight 1.0 (Mitchell *et al.*, 1997)

Calculation of the amino acid sequence of consensus phytase-1

Using the refined alignment as input, the consensus sequence was calculated by the program PRETTY from the Sequence Analysis Package Release 9.0 (Devereux *et al.*,
5 1984). PRETTY prints sequences with their columns aligned and can display a consensus sequence for an alignment. A vote weight that pays regard to the similarity between the amino acid sequences of the phytases aligned was assigned to all sequences. The vote weight was set such as the combined impact of all phytases from one sequence subgroup (same species, but from different strains), e. g. the amino acid sequences of all phytases
10 from *A. fumigatus*, on the election was set one, that means that each sequence contributes with a value of 1 divided by the number of strain sequences (see Table 1). By this means, it was possible to prevent that very similar amino acid sequences, e. g. of the phytases from different *A. fumigatus* strains, dominate the calculated consensus sequence.

The program PRETTY was started with the following parameters: The plurality
15 defining the number of votes below which there is no consensus was set on 2.0. The threshold, which determines the scoring matrix value below which an amino acid residue may not vote for a coalition of residues, was set on 2. PRETTY used the PrettyPep.Cmp consensus scoring matrix for peptides.

Ten positions of the alignment (position 46, 66, 82, 138, 162, 236, 276, 279, 280,
20 308; Figure 1), for which the program was not able to determine a consensus residue, were filled by hand according to the following rules: if a most frequent residue existed, this residue was chosen (138, 236, 280); if a prevalent group of similar or phylogenetically equivalent residues occurred, the most frequent or, if not available, one residues of this group was selected (46, 66, 82, 162, 276, 308). If there was either a prevalent residue nor a
25 prevalent group, one of the occurring residues was chosen according to common assumption on their influence on the protein stability (279). Eight other positions (132, 170, 204, 211, 275, 317, 384, 447; Figure 1) were not filled with the amino acid residue selected by the program but normally with amino acids that occur with the same frequency as the residues that were chosen by the program. In most cases, the slight underrating of
30 the three *A. niger* sequences (sum of the vote weights: 0.99) was eliminated by this corrections.

Conversion of the consensus phytase-1 amino acid sequence to a DNA sequence

The first 26 amino acid residues of *A. terreus* cbs116.46 phytase were used as signal
35 peptide and, therefore, fused to the N-terminus of all consensus phytases. For this stretch,

we used a special method to calculate the corresponding DNA sequence. Purvis et al (1987) proposed that the incorporation of rare codons in a gene has an influence on the folding efficiency of the protein. Therefore, at least the distribution of rare codons in the signal sequence of *A. terreus* cbs1 16.46, which was used for the consensus phytase and
5 which is very important for secretion of the protein, but converted into the *S. cerevisiae* codon usage, was transferred into the new signal sequence generated for expression in *S. cerevisiae*. For the remaining parts of the protein, we used the codon frequency table of highly expressed *S. cerevisiae* genes, obtained from the GCG program package, to translate the calculated amino acid sequence into a DNA sequence.

10 The resulting sequence of the *fcp* gene is shown in Figure 2.

Construction and cloning of the consensus phytase-1 gene

The calculated DNA sequence of consensus phytase-1 (*fcp*) was divided into oligonucleotides of 85 bp, alternately using the sequence of the sense and the anti-sense strand. Every oligonucleotide overlaps 20 bp with its previous and its following
15 oligonucleotide of the opposite strand. The location of all primers, purchased by Microsynth, Balgach (Switzerland) and obtained in a PAGE-purified form, is indicated in Figure 2.

PCR-Reactions

In three PCR reactions, the synthesized oligonucleotides were composed to the entire
20 gene. For the PCR, the High Fidelity Kit from Boehringer Mannheim (Boehringer Mannheim, Mannheim, Germany) and the thermo cycler The ProtokolTM from AMS Biotechnology (Europe) Ltd. (Lugano, Switzerland) was used.

Oligonucleotide CP-1 to CP-10 (Mix 1, Figure 2) were mixed to a concentration of 0.2 pMol/ μ l of each oligonucleotide. A second oligonucleotide mixture (Mix 2) was
25 prepared with CP-9 to CP-22 (0.2 pMol/ μ l of each oligonucleotide). Additionally, four short primers were used in the PCR reactions:

CP-a: *Eco* RI

5'-TATATGAATTCATGGGCGTGTTCGTC-3'

CP-b: 5'-TGAAAAGTTCATTGAAGGTTTC-3'
30

CP-c: 5'-TCTTCGAAAGCAGTACAAGTAC-3'

CP-e:

Eco RI

5'-TATATGAATTCTTAAGCGAAAC-3'

5 PCR reaction *a*: 10 µl Mix 1 (2.0 pmol of each oligonucleotide)
 2 µl nucleotides (10 mM each nucleotide)
 2 µl primer CP-a (10 pmol/µl)
 2 µl primer CP-c (10 pmol/µl)
 10,0 µl PCR buffer
10 0.75 µl polymerase mixture
 73.25 µl H₂O

 PCR reaction *b*: 10 µl Mix 2 (2.0 pmol of each oligonucleotide)
 2 µl nucleotides (10 mM each nucleotide)
 2 µl primer CP-b (10 pmol/µl)
 2 µl primer CP-e (10 pmol/µl)
15 10,0 µl PCR buffer
 0.75 µl polymerase mixture (2.6 U)
 73.25 µl H₂O

Reaction conditions for PCR reaction *a* and *b*:

20 step 1 2 min - 45°C
 step 2 30 sec - 72°C
 step 3 30 sec - 94°C
 step 4 30 sec - 52°C
 step 5 1 min - 72°C

Step 3 to 5 were repeated 40-times.

25 The PCR products (670 and 905 bp) were purified by an agarose gel electrophoresis
(0.9% agarose) and a following gel extraction (QIAEX II Gel Extraction Kit, Qiagen,
Hilden, Germany). The purified DNA fragments were used for the PCR reaction *c*.

30 PCR reaction *c*: 6 µl PCR product of reaction *a* (≈50 ng)
 6 µl PCR product of reaction *b* (≈50 ng)
 2 µl primer CP-a (10 pmol/µl)
 2 µl primer CP-e (10 pmol/µl)
 10,0 µl PCR buffer
 0.75 µl polymerase mixture (2.6 U)
 73.25 µl H₂O

35 Reaction conditions for PCR reaction *c*:

step 1 2 min - 94°C
step 2 30 sec - 94°C
step 3 30 sec - 55°C
step 4 1 min - 72°C

5 Step 2 to 4 were repeated 31-times.

The resulting PCR product (1.4 kb) was purified as mentioned above, digested with *Eco* RI, and ligated in an *Eco* RI-digested and dephosphorylated pBsk(-)-vector (Stratagene, La Jolla, CA, USA). 1 µl of the ligation mixture was used to transform *E. coli* XL-1 competent cells (Stratagene, La Jolla, CA, USA). All standard procedures were
10 carried out as described by Sambrook *et al.* (1987). The DNA sequence of the constructed consensus phytase gene (*fcp*, Figure 2) was controlled by sequencing as known in the art.

Example 2

Design of an improved consensus phytase (consensus phytase-10) amino acid sequence

15 The alignments used for the design of consensus phytase-10 were calculated using the program PILEUP from the Sequence Analysis Package Release 9.0 (Devereux *et al.*, 1984) with the standard parameter (gap creation penalty 12, gap extension penalty 4). The location of the gaps was refined using a text editor.

The following sequences were used for the alignment of the *Basidiomycetes* phytases
20 starting with the amino acid (aa) mentioned in Table 2:

Table 2: Origin and vote weight of five *Basidiomycetes* phytases used for the calculation of the corresponding amino acid consensus sequence (basidio)

- *phyA1* from *Paxillus involutus* NN005693, aa 21, vote weight 0.5 (WO 98/28409)
- *phyA2* from *Paxillus involutus* NN005693, aa 21, vote weight 0.5 (WO 98/28409)
- 25 - *phyA* from *Trametes pubescens* NN9343, aa 24, vote weight 1.0 (WO 98/28409)
- *phyA* from *Agrocybe pediades* NN009289, aa 19, vote weight 1.0 (WO 98/28409)
- *phyA* from *Peniophora lycii* NN006113, aa 21, vote weight 1.0 (WO 98/28409)

The alignment is shown in Figure 3.

In Table 3 the genes, which were used for the performance of the final alignment, are
30 arranged. The first amino acid (aa) of the sequence which is used in the alignment is mentioned behind the organism designation.

Table 3: Origin and vote weight of the phytase sequences used for the design of consensus phytase 10

- *phyA* from *Aspergillus terreus* 9A-1, aa 27, vote weight 0.5 (Mitchell *et al.*, 1997)
- *phyA* from *Aspergillus terreus* cbs116.46, aa 27, vote weight 0.5 (van Loon *et al.*, 1998)
- 5 - *phyA* from *Aspergillus niger* var. *awamori*, aa 27, vote weight 0.5 (Piddington *et al.*, 1993)
- *phyA* from *Aspergillus niger* strain NRRL3135, aa 27, vote weight 0.5 (van Hartingsveldt *et al.*, 1993)
- *phyA* from *Aspergillus fumigatus* ATCC 13073, aa 26, vote weight 0.2 (Pasamontes *et al.*, 1997)
- 10 - *phyA* from *Aspergillus fumigatus* ATCC 32722, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)
- *phyA* from *Aspergillus fumigatus* ATCC 58128, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)
- 15 - *phyA* from *Aspergillus fumigatus* ATCC 26906, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)
- *phyA* from *Aspergillus fumigatus* ATCC 32239, aa 30, vote weight 0.2 (van Loon *et al.*, 1998)
- *phyA* from *Emericella nidulans*, aa 25, vote weight 1.0 (Roche Nr. R1288, Pasamontes *et al.*, 1997a)
- 20 - *phyA* from *Talaromyces thermophilus* ATCC 20186, aa 24, vote weight 1.0 (Pasamontes *et al.*, 1997a)
- *phyA* from *Myceliophthora thermophila*, aa 19, vote weight 1.0 (Mitchell *et al.*, 1997)
- *phyA* from *Thermomyces lanuginosa*, aa 36, vote weight 1.0 (Berka *et al.*, 1998)
- 25 - Consensus sequence of five *Basidiomycetes* phytases, vote weight 1.0 (Basidio, Figure 3)

The corresponding alignment is shown in Figure 4.

Calculation of the amino acid sequence of consensus-10

To improve the alignment, we added the original consensus sequence of five phytases from four different *Basidiomycetes*, called Basidio, still containing the undefined sequence positions (see Figure 3), nearly all phytase sequences used for calculation of the original consensus phytase and one new phytase sequence from the *Ascomycete* *Thermomyces lanuginosa* to a larger alignment. Using the consensus sequence of the basidiomycetal phytase sequences, does not pay regard to the diversity among the five amino acid sequences, but pays regard to the common and different amino acid residues between the phytases from the *Ascomycetes* and the *Basidiomycetes*.

We set plurality on 2.0 and threshold on 3. The used vote weight are listed in Table 3. The alignment and the corresponding consensus sequence is presented in Figure 4. The new consensus phytase sequence has 32 different amino acids in comparison to the original consensus phytase. Positions for which the program PRETTY was not able to calculate a
5 consensus amino acid residue were filled according to rules mentioned in Example 1. None of the residues suggested by the program was replaced.

Furthermore, we included all *Basidiomycetes* phytases as single amino acid sequences but assigning a vote weight of 0.2 in the alignment. The corresponding alignment is shown in Figure 6. The calculated consensus amino acid sequence (consensus phytase-11) has the
10 following differences to the sequence of consensus phytase-10. Letter X means that the program was not able to calculate a consensus amino acid; the amino acid in parenthesis corresponds to the amino acid finally included into the consensus phytase-10.

D35X, X(K)69K, X(E)100E, A101R, Q134N, X(K)153N, X(H)190H, X(A)204S,
X(E)220D, E222T, V227A, X(R)271R, H287A, X(D)288D, X(K)379K, X(T)389I, E390X,
15 X(E)415E, X(A)416A, X(R)446L, E463A, whereas the numbering is as in Fig. 5.

We also checked single amino acid replacements suggested by the improved consensus sequences 10 and 11 on their influence on the stability of the original consensus phytase. The approach is described in example 3.

20 **Conversion of consensus phytase-10 amino acid sequence to a DNA sequence**

The first 26 amino acid residues of *A. terreus* cbs116.46 phytase were used as signal peptide and, therefore, fused to the *N*-terminus of consensus phytase-10. The used procedure is further described in Example 1.

The resulting sequence of the *fcp10* gene is shown in Figure 5.

25

Construction and cloning of the consensus phytase-10 gene (*fcp10*)

The calculated DNA sequence of *fcp10* was divided into oligonucleotides of 85 bp, alternately using the sequence of the sense and the anti-sense strand. Every oligonucleotide overlaps 20 bp with its previous and its following oligonucleotide of the opposite strand.

30 The location of all primers, purchased by Microsynth, Balgach (Switzerland) and obtained in a PAGE-purified form, is indicated in Figure 5.

PCR-Reactions

In three PCR reactions, the synthesized oligonucleotides were composed to the entire gene. For the PCR, the High Fidelity Kit from Boehringer Mannheim (Boehringer Mannheim, Mannheim, Germany) and the thermo cycler The ProtokolTM from AMS Biotechnology (Europe) Ltd. (Lugano, Switzerland) was used. The following oligonucleotides were used in a concentration of 0.2 pMol/ml.

Mix 1.10: CP-1, CP-2, CP-3.10, CP-4.10, CP-5.10, CP-6, CP-7.10, CP-8.10, CP-9.10, CP-10.10

Mix 2.10: CP-9.10, CP-11.10, CP-12.10, CP-13.10, CP-14.10, CP-15.10, CP-16.10, CP-17.10, CP18.10, CP-19.10, CP-20.10, CP-21.10, CP-22.10

The newly synthesized oligonucleotides are marked by number 10. The phytase contains the following 32 exchanges, which are underlined in Figure 5, in comparison to the original consensus phytase: Y54F, E58A, D69K, D70G, A94K, N134Q, I158V, S187A, Q188N, D197N, S204A, T214L, D220E, L234V, A238P, D246H, T251N, Y259N, E267D, E277Q, A283D, R291I, A320V, R329H, S364T, I366V, A379K, S396A, G404A, Q415E, A437G, A463E.

Four short PCR primer were used for the assembling of the oligonucleotides:

CP-a: *Eco* RI
5'-TATATGAATTCATGGGCGTGTCGTC-3'

CP-b: 5'-TGAAAAGTTCATTGAAGGTTTC-3'

CP-c.10: 5'-TCTTCGAAAGCAGTACACAAAC-3'

CP-e: *Eco* RI
5'-TATATGAATTCTTAAGCGAAAC-3'

PCR reaction α :
10 μ l Mix 1.10 (2.0 pmol of each oligonucleotide)
2 μ l nucleotides (10 mM each nucleotide)
2 μ l primer CP-a (10 pmol/ml)
2 μ l primer CP-c.10 (10 pmol/ml)
10,0 μ l PCR buffer
0.75 μ l polymerase mixture
73.25 μ l H₂O

5 PCR reaction *b*: 10 µl Mix 2.10 (2.0 pmol of each oligonucleotide)
 2 µl nucleotides (10 mM each nucleotide)
 2 µl primer CP-b (10 pmol/ml)
 2 µl primer CP-e (10 pmol/ml)
 10,0 µl PCR buffer
 0.75 µl polymerase mixture (2.6 U)
 73.25 µl H₂O

10 Reaction conditions for PCR reaction *a* and *b*:
 step 1 2 min - 45 °C
 step 2 30 sec - 72 °C
 step 3 30 sec - 94 °C
 step 4 30 sec - 52 °C
 step 5 1 min - 72 °C

Step 3 to 5 were repeated 40-times.

15 The PCR products (670 and 905 bp) were purified by an agarose gel electrophoresis (0.9% agarose) and a following gel extraction (QIAEX II Gel Extraction Kit, Qiagen, Hilden, Germany). The purified DNA fragments were used for the PCR reaction *c*.

20 PCR reaction *c*: 6 µl PCR product of reaction *a* ≈50 ng)
 6 µl PCR product of reaction *b* ≈50 ng)
 2 µl primer CP-a (10 pmol/ml)
 2 µl primer CP-e (10 pmol/ml)
 10,0 µl PCR buffer
 0.75 µl polymerase mixture (2.6 U)
 73.25 µl H₂O

25 Reaction conditions for PCR reaction *c*:
 step 1 2 min - 94 °C
 step 2 30 sec - 94 °C
 step 3 30 sec - 55 °C
 step 4 1 min - 72 °C

30 Step 2 to 4 were repeated 31-times.

35 The resulting PCR product (1.4 kb) was purified as mentioned above, digested with *Eco* RI, and ligated in an *Eco* RI-digested and dephosphorylated pBsk(-)-vector (Stratagene, La Jolla, CA, USA). 1 µl of the ligation mixture was used to transform *E. coli* XL-1 competent cells (Stratagene, La Jolla, CA, USA). All standard procedures were carried out as described by Sambrook *et al.* (1987). The DNA sequence of the constructed gene (*fcp10*) was checked by sequencing as known in the art.

Example 3

Increasing the thermostability of consensus phytase-1 by introduction of single mutations suggested by the amino acid sequence of consensus phytase-10 and consensus phytase-11

5 In order to increase the thermostability of homologous genes, it is also possible to test the stability effect of each differing amino acid residue between the protein of interest and the calculated consensus sequence and to combine all stabilizing mutations into the protein of interest. We used the consensus phytase as protein of interest and tested the effect on the protein stability of 34 amino acid residues, differing to consensus phytase 10
10 and/or 11 as single mutations.

To construct muteins for expression in *A. niger*, *S. cerevisiae*, or *H. polymorpha*, the corresponding expression plasmid containing the consensus phytase gene was used as template for site-directed mutagenesis (see Example 6-8). Mutations were introduced using the "quick exchangeTM site-directed mutagenesis kit" from Stratagene (La Jolla, CA,
15 USA) following the manufacturer's protocol and using the corresponding primers. All mutations made and their corresponding primers are summarized in Table 4. Plasmids harboring the desired mutation were identified by DNA sequence analysis as known in the art.

Table 4: Primers used for site-directed mutagenesis of consensus phytase

20 (Exchanged bases are highlighted in bold. The introduction of a restriction site is marked above the sequence. When a restriction site is written in parenthesis, the mentioned site was destroyed by introduction of the mutation.)

| mutation | Primer set |
|----------|---|
| 25 Q50T | <p style="text-align: center;"><i>Kpn</i> I</p> <p>5'-CACTTGTGGGGTACCTACTCTCCATACTTCTC-3'</p> <p>5'-GAGAAGTATGGAGAGTAGGTACCCCAAGTG-3'</p> |
| 30 Y54F | <p>5'-GGTCAATACTCTCCATTCTTCTTTGGAAG-3'</p> <p>5'-CTTCCAAAGAGAAGAATGGAGAGTATTGACC-3'</p> |
| E58A | <p>5'-CATACTTCTCTTTGGCAGACGAATCTGC-3'</p> <p>5'-GCAGATTCGTCTGCCAAAGAGAAGTATG-3'</p> |

| | | |
|----|-------|--|
| | | <i>Aat</i> II |
| | D69K | 5'-CTCCAGACGTCCCAAAGGACTGTAGAGTTAC-3' 5'-GTA ACTCTACAGTCCTTTGGGACGTCTGGAG-3' |
| | | <i>Aat</i> II |
| 5 | D70G | 5'-CTCCAGACGTCCAGACGGCTGTAGAGTTAC-3' 5'-GTA ACTCTACAGCCGTCTGGGACGTCTGGAG-3' |
| | K91A | 5'-GATACCCA ACTTCTTCTGCGTCTAAGGCTTACTCTG-3' 5'-CAGAGTAAGCCTTAGACGCAGAAGAAGTTGGGTATC-3' |
| 10 | | <i>Sca</i> I |
| | A94K | 5'-CTTCTAAGTCTAAGAAGTACTCTGCTTTG-3' 5'-CAAAGCAGAGTACTTCTTAGACTTAGAAG-3' |
| | A101R | 5'-GCTTACTCTGCTTTGATTGAACGGATTCAAAAGAACGCTAC-3' 5'-GTAGCGTTCTTTTGAATCCGTTCAATCAAAGCAGAGTAAGC-3' |
| 15 | | |
| | N134Q | 5'-CCATTCGGTGAACAGCAAATGGTTAACTC-3' 5'-GAGTTAACCATTGCTGTTACCGAATGG-3' |
| | | <i>Nru</i> I |
| 20 | K153N | 5'-GATACAAGGCTCTCGCGAGAAACATTGTTC-3' 5'-GGAACAATGTTTCTCGCGAGAGCCTTGTATC-3' |
| | | <i>Bss</i> HI |
| | I158V | 5'-GATTGTTCCATTTCGTGCGCGCTTCTGGTTC-3' 5'-GAACCAGAAGCGCGCACGAATGGAACAATC-3' |
| | | <i>Bcl</i> I |
| 25 | D197N | 5'-CTCCAGTTATTAACGTGATCATTCCAGAAGG-3' 5'-CCTTCTGGAATGATCACGTTAATAACTGGAG-3' |
| | | <i>Apa</i> I |
| | S187A | 5'-GGCTGACCCAGGGGCCCCAACCACACCAAGC-3' 5'-GCTTGGTGTGGTTGGGCCCCTGGGTCAGCC-3' |
| 30 | | |
| | T214L | 5'-CACTTTGGACCATGGTCTTTGTACTGCTTTTCG-3' 5'-CGAAAGCAGTACAAAGACCATGGTCCAAAGTG-3' |
| | | <i>Avr</i> II |
| 35 | E222T | 5'-GCTTTCGAAGACTCTACCCTAGGTGACGACGTTG-3' 5'-CAACGTCGTCACCTAGGGTAGAGTCTTCGAAAGC-3' |

| | | |
|----|-------|---|
| | V227A | 5'-GGTGACGACGCTGAAGCTAACTTCAC-3' 5'-GTGAAGTTAGCTTCAGCGTCGTCACC-3' |
| | | <i>Sac II</i> |
| 5 | L234V | 5'-CTAACTTCACCGCGGTGTTTCGCTCCAG-3' 5'-CTGGAGCGAACACCGCGGTGAAGTTAG-3' |
| | A238P | 5'-GCTTTGTTTCGCTCCACCTATTAGAGCTAGATTGG-3' 5'-CCAATCTAGCTCTAATAGGTGGAGCGAACAAAGC-3' |
| | | <i>Hpa I</i> |
| 10 | T251N | 5'-GCCAGGTGTTAACTTGACTGACGAAG-3' 5'-TTCGTCAGTCAAGTTAACACCTGGC-3' |
| | | <i>Aat II</i> |
| | Y259N | 5'-GACGAAGACGTCGTAACTTGATGGAC-3' 5'-GTCCATCAAGTTAACGACGTCTTCGTC-3' |
| 15 | | |
| | | <i>Asp I</i> |
| | E267D | 5'-GTCCATTGACACTGTCGCTAGAACTT C-3' 5'-GAAGTTCTAGCGACAGTGTCGAATGGAC-3' |
| | | |
| 20 | E277Q | 5'-CTGACGCTACTCAGCTGTCTCCATT C-3' 5'-GAATGGAGACAGCTGAGTAGCGTCAG-3' |
| | | |
| | A283D | 5'-GTCTCCATTCTGTGATTTGTTCACTCAC-3' 5'-GTGAGTGAACAAATCACAGAATGGAGAC-3' |
| | | |
| 25 | | <i>Ksp I</i> |
| | H287A | 5'-GCTTTGTTCAACCGCGGACGAATGGAG-3' 5'-CTCCATTTCGTCGCGGTGAACAAAGC-3' |
| | | |
| | | <i>Bam HI</i> |
| | R291I | 5'-CACGACGAATGGATCCAATACGACTAC-3' 5'-GTAGTCGTATTGGATCCATTTCGTCGTG-3' |
| 30 | | |
| | | <i>Bsi WI</i> |
| | Q292A | 5'-GACGAATGGAGAGCGTACGACTACTTG-3' 5'-CAAGTAGTCGTACGCTCTCCATTTCGTC-3' |
| | | |
| | | <i>Hpa I</i> |
| 35 | A320V | 5'-GGTGTTGGTTTCGTTAACGAATTGATTGC-3' 5'-GCAATCAATTTCGTTAACGAACCAACACC-3' |
| | | |
| | | (<i>Bgl II</i>) |
| | R329H | 5'-GCTAGATTGACTCACTCTCCAGTTCAAG-3' 5'-CTTGAAGTGGAGAGTGAGTCAATCTAGC-3' |

| | | |
|----|-------|---|
| | | <i>Eco</i> RV |
| | S364T | 5'-CTCACGACAACACTATGATATCTATTTTCTTC-3' 5'-GAAGAAAATAGATATCATAGTGTGTCGTGAG-3' |
| | | <i>Nco</i> I |
| 5 | I366V | 5'-CGACAACTCCATGGTTTCTATTTTCTTCGC-3' 5'-GCGAAGAAAATAGAAACCATGGAGTTGTCG-3' |
| | | <i>Kpn</i> I |
| | A379K | 5'-GTACAACGGTACCAAGCCATTGTCTAC-3' 5'-GTAGACAATGGCTTGGTACCGTTGTAC-3' |
| 10 | S396A | 5'-CTGACGGTTACGCTGCTTCTTGGAC-3' 5'-GTCCAAGAAGCAGCGTAACCGTCAG-3' |
| | G404A | 5'-CTGTTCCATTTCGCTGCTAGAGCTTAC-3' 5'-GTAAGCTCTAGCAGCGAATGGAACAG-3' |
| 15 | Q415E | 5'-GATGCAATGTGAAGCTGAAAAGGAACC-3' 5'-GGTTCCTTTTCAGCTTCACATTGCATC-3' |
| | | <i>Sal</i> I |
| 20 | A437G | 5'-CACGGTTGTGGTGTGCGACAAGTTGGG-3' 5'-CCCAACTTGTGCGACACCACAACCGTG-3' |
| | | <i>Mun</i> I |
| | A463E | 5'-GATCTGGTGGCAATTGGGAGGAATGTTTCG-3' 5'-CGAAACATTCCTCCCAATTGCCACCAGATC-3' |

25 and accordingly for other mutations.

The temperature optimum of the purified phytases, expressed in *Saccharomyces cerevisiae* (Example 7), was determined as outlined in Example 9. Table 5 shows the effect on the stability of consensus phytase for each mutation introduced.

30

Table 5: Stability effect of the individual amino acid replacements in consensus phytase-1
(+ or - means a positive, respectively, negative effect on the protein stability up to 1 °C, ++ and -- means a positive, respectively, negative effect on the protein stability between 1 and 3 °C; the number 10 or 11 corresponds to the consensus phytase sequence that suggests the
35 amino acid replacement.)

| stabilizing | | neutral | | destabilizing | |
|-------------|--------|------------|--------|---------------|--------|
| mutation | effect | mutation | effect | mutation | effect |
| E58A (10) | + | D69A | ± | Y54F (10) | - |
| D69K (11) | + | D70G (10) | ± | V73I | - |
| D197N (10) | + | N134Q (10) | ± | A94K (10) | - |
| T214L (10) | ++ | G186H | ± | A101R (11) | - |
| E222T (11) | ++ | S187A (10) | ± | K153N (11) | - |
| E267D (10) | + | T214V | ± | I158V (10) | -- |
| R291I* | + | T251N (10) | ± | G203A | -- |
| R329H (10) | + | Y259N (10) | ± | G205S | - |
| S364T (10) | ++ | A283D (10) | ± | A217V | - |
| A379K (11) | + | A320V (10) | ± | V227A (11) | -- |
| G404A (10) | ++ | K445T | ± | L234V (10) | - |
| | | A463E (10) | ± | A238P (10) | -- |
| | | | | E277Q (10) | - |
| | | | | H287A (11) | - |
| | | | | Q292A (10) | - |
| | | | | I366V (10) | - |
| | | | | S396A (10) | -- |
| | | | | Q415E (11) | - |
| | | | | A437G (10) | -- |
| | | | | E451R | -- |

*: This amino acid replacement was found in another round of mutations.

We combined eight positive mutations (E58A, D197N, E267D, R291I, R329H, S364T, A379K, G404A) in the consensus phytase using the primers and the technique mentioned above in this example. Furthermore, the mutations Q50T and K91A were introduced which mainly influence the catalytical characteristics of the phytase (see patent application EP 97810175.6 and EP 97112688 as well as Example 9). The DNA and amino acid sequence of the resulting phytase gene (consensus phytase-thermo[8]-Q50T-K91A) is shown in Figure 7. In this way, the temperature optimum and the melting point of the consensus phytase was increased by 7 °C (Figure 15, 16, 17).

Using the results of Table 5, we further improved the thermostability of consensus phytase 10 by the following back mutations K94A, V158I, and A396S that revealed a strong negative influence on the stability of consensus phytase. The resulting protein is phytase-10-thermo [3]. Furthermore, we introduced the mutations Q50T and K91A which mainly influence the catalytical characteristics of consensus phytase (see patent application EP 97810175.6 and EP 97112688 as well as Example 9 and Figure 14 and 15). The resulting DNA and amino acid sequence is shown in Figure 8. The optimized phytase showed a 4 °C higher temperature optimum and melting point than consensus phytase 10 (Figure 12 and 13). Furthermore, the phytase has also a strongly increased specific activity with phytate as substrate of 250 U/mg at pH 5.5 (Figure 14).

Example 4

Stabilization of the phytase of *A. fumigatus* ATCC 13073 by replacement of amino acid residues with the corresponding consensus phytase-1 and consensus phytase-10 residues

At six typical positions where the *A. fumigatus* 13073 is the only or nearly the only phytase in the alignment of Figure 1 that does not contain the corresponding consensus phytase amino acid residue, the non-consensus amino acid residue was replaced by the consensus one. In a first round, the following amino acids were substituted in *A. fumigatus* 13073 phytase, containing the Q27T substitution and the signal sequence of *A. terreus* cbs.116.46 phytase (see European Patent Application No. 97810175.6 and Figure 9):

F55(28)Y, V100(73)I, F114(87)Y, A243(220)L, S265(242)P, N294(282)D.

The numbers in parentheses confer to the numbering of Figure 1.

In a second round, four of the seven stabilizing amino acid exchanges (E59A, R329H, S364T, G404A) found in the consensus phytase-10 sequence and, tested as single mutation in consensus phytase-1 (Table 5), were additionally introduced into the *A. fumigatus* a-

mutant. Furthermore, the amino acid replacement S126N, shown to reduce the protease susceptibility of the phytase, was introduced.

The mutations were introduced as described in example 3 (see Table 6) and expressed as described in example 6 to 8. The resulting *A. fumigatus* 13073 phytase variants were called a-mutant and β -mutant-E59A-S126N-R329H-S364T-G404A.

The temperature optimum (60 °C, Figure 20) and the melting point (67.0 °C, Figure 19) of the *A. fumigatus* 13073 phytase β -mutant was increased by 5 °C in comparison to the values of the wild-type (temperature optimum: 55 °C, T_m : 60 °C). The five additional amino acid replacements further increased the temperature optimum by 3 °C (Figure 20).

10 Table 6: Mutagenesis primers for stabilization of *A. fumigatus* phytase ATCC 13073

| Mutation | Primer |
|----------|---|
| F55Y | 5'-CACGTA CT CGCCATACTTTTCGCTCGAG-3' 5'-CTCGAGCGAAAAGTATGGCGAGTACGTG-3' |
| | (<i>Xho</i> I) |
| 15 E58A | 5'-CCATACTTTTCGCTCGCGGACGAGCTGTCCGTG-3' 5'-CACGGACAGCTCGTCCGCGAGCGAAAAGTAGG-3' |
| V100I | 5'-GTATAAGAAGCTTATTACGGCGATCCAGGCC-3' 5'-GGCCTGGATCGCCGTAATAAGCTTCTTATAC-3' |
| 20 F114Y | 5'-CTTCAAGGGCAAGTACGCCTTTTTGAAGACG-3' 5'-CGTCTTCAAAAAGGCGTACTTGCCCTTGAAG-3' |
| A243L | 5'-CATCCGAGCTCGCCTCGAGAAGCATCTTC-3' |
| 25 | 5'-GAAGATGCTTCTCGAGGCGAGCTCGGATG-3' |
| S265P | 5'-CTAATGGA TGTGTCCGTTTGATACGGTAG-3' 5'-CTACCGTATCAAACGGACACATGTCCATTAG-3' |

N294D 5'-GTGGAAGAAGTACGACTACCTTCAGTC-3'
5'-GACTGAAGGTAGTCGTACTTCTTCCAC-3'

(Mlu I)

5 R329H 5'-GCCCCGGTTGACGCAATTCGCCAGTGCAGG-3'
5'-CCTGCACTGGCGAATGCGTCAACCGGGC-3'

Nco I

S364T 5'-CACACGACAACACCATGGTTTCCATCTTC-3'
5'-GAAGATGGAAACCATGGTGTGTCGTGTG-3'

10 (Bss HI)

G404A 5'-GTGGTGCCTTTCGCCGCGGAGCCTACTTC-3'
5'-GAAGTAGGCTCGCGCGCGAAAGGCACCAC-3'

Example 5

15 Introduction of the active site amino acid residues of the *A. niger* NRRL 3135
phytase into the consensus phytase-1

We used the crystal structure of the *Aspergillus niger* NRRL 3135 phytase to define all active site amino acid residues (see Reference Example and EP 97810175.6). Using the alignment of Figure 1, we replaced the following active site residues and additionally the not identical adjacent ones of the consensus phytase by that of the *A. niger* phytase:

20 S89D, S92G, A94K, D164S, P201S, G203A, G205S, H212P, G224A, D226T, E255T, D256E, V258T, P265S, Q292H, G300K, Y305H, A314T, S364G, M365I, A397S, S398A, G404A, and A405S

25 The new protein sequence consensus phytase -7 was backtranslated into a DNA sequence (Figure 10) as described in Example 1. The corresponding gene (*fcp7*) was generated as described in Example 1 using the following oligonucleotide mixes:

Mix 1.7: CP-1, CP-2, CP-3, CP-4.7, CP-5.7, CP-6, CP-7, CP-8.7, CP-9, CP-10.7

Mix 2.7: CP-9, CP-10.7, CP-11.7, CP-12.7, CP-13.7, CP-14.7, CP-15.7, CP-16, CP-17.7, CP-18.7, CP-19.7, CP-20, CP-21, CP-22.

30 The DNA sequences of the oligonucleotides are indicated in Figure 3. The newly synthesized oligonucleotides are additionally marked by number 7. After assembling of the

oligonucleotides using the same PCR primers as mentioned in Example 1, the gene was cloned into an expression vector as described in Examples 6-8.

The pH-profile determined after expression in *H. polymorpha* and purification was shifted into the acidic range of the pH-spectrum showing an optimum at pH 4.5-5.0 (see Figure 18). The enzyme had a broad pH-optimum reaching at least 60% of its maximum activity from pH 2.5 to pH 6.0. Up to pH 5.0, the profile resembled the profile of the *A. niger* NRRL 3135 phytase. However, below pH 5.0 it lacked the typical low at pH 4.0 of the profile of *A. niger* phytase.

Example 6

Expression of the consensus phytase genes in *Hansenula polymorpha*

The phytase expression vectors, used to transform *H. polymorpha* RB11 (Gellissen *et al.*, 1994), was constructed by inserting the *Eco* RI fragment of pBsk-*fcp* or variants thereof into the multiple cloning site of the *H. polymorpha* expression vector pFPMT121, which is based on an *ura3* selection marker from *S. cerevisiae*, a formate dehydrogenase (*FMD*) promoter element and a methanol oxidase (*MO*) terminator element from *H. polymorpha*. The 5' end of the *fcp* gene is fused to the *FMD* promoter, the 3' end to the *MOX* terminator (Gellissen *et al.*, 1996; EP 0299 108 B). The resulting expression vector are designated pFPMT*fcp*, pFPMT*fcp10*, pFPMT*fcp7*.

The constructed plasmids were propagated in *E. coli*. Plasmid DNA was purified using standard state of the art procedures. The expression plasmids were transformed into the *H. polymorpha* strain RP11 deficient in orotidine-5'-phosphate decarboxylase (*ura3*) using the procedure for preparation of competent cells and for transformation of yeast as described in Gellissen *et al.* (1996). Each transformation mixture was plated on YNB (0.14% w/v Difco YNB and 0.5% ammonium sulfate) containing 2% glucose and 1.8% agar and incubated at 37 °C. After 4 to 5 days individual transformant colonies were picked and grown in the liquid medium described above for 2 days at 37 °C. Subsequently, an aliquot of this culture was used to inoculate fresh vials with YNB-medium containing 2% glucose. After seven further passages in selective medium, the expression vector integrates into the yeast genome in multimeric form. Subsequently, mitotically stable transformants were obtained by two additional cultivation steps in 3 ml non-selective liquid medium (YPD, 2% glucose, 10 g yeast extract, and 20 g peptone). In order to obtain genetically homogeneous recombinant strains an aliquot from the last stabilization culture was plated on a selective plate. Single colonies were isolated for analysis of phytase expression in YNB containing 2% glycerol instead of glucose to derepress the *fmd* promoter. Purification of the consensus phytases was done as described in Example 7.

Example 7

Expression of the consensus phytase genes in *Saccharomyces cerevisiae* and purification of the phytases from culture supernatant

The consensus phytase genes were isolated from the corresponding Bluescript-plasmid (pBsk \overline{fcp} , pBSK $\overline{fcp10}$, pBsk $\overline{fcp7}$) and ligated into the *Eco* RI sites of the expression cassette of the *Saccharomyces cerevisiae* expression vector pYES2 (Invitrogen, San Diego, CA, USA) or subcloned between the shortened GAPFL (glyceraldehyde-3-phosphate dehydrogenase) promoter and the *pho5* terminator as described by Janes *et al.* (1990). The correct orientation of the gene was checked by PCR. Transformation of *S. cerevisiae* strains. e. g. INVSc1 (Invitrogen, San Diego, CA, USA) was done according to Hinnen *et al.* (1978). Single colonies harboring the phytase gene under the control of the GAPFL promoter were picked and cultivated in 5 ml selection medium (SD-uracil, Sherman *et al.*, 1986) at 30°C under vigorous shaking (250 rpm) for one day. The preculture was then added to 500 ml YPD medium (Sherman *et al.*, 1986) and grown under the same conditions. Induction of the *gal1* promoter was done according to manufacturer's instruction. After four days of incubation cell broth was centrifuged (7000 rpm, GS3 rotor, 15 min, 5°C) to remove the cells and the supernatant was concentrated by way of ultrafiltration in Amicon 8400 cells (PM30 membranes) and ultrafree-15 centrifugal filter devices (Biomax-30K, Millipore, Bedford, MA, USA). The concentrate (10 ml) was desalted on a 40 ml Sephadex G25 Superfine column (Pharmacia Biotech, Freiburg, Germany), with 10 mM sodium acetate, pH 5.0, serving as elution buffer. The desalted sample was brought to 2 M (NH₄)₂SO₄ and directly loaded onto a 1 ml Butyl Sepharose 4 Fast Flow hydrophobic-interaction chromatography column (Pharmacia Biotech, Freiburg, Germany) which was eluted with a linear gradient from 2 M to 0 M (NH₄)₂SO₄ in 10 mM sodium acetate, pH 5.0. Phytase was eluted in the break-through, concentrated and loaded on a 120 ml Sephacryl S-300 gel permeation chromatography column (Pharmacia Biotech, Freiburg, Germany). Consensus phytase and consensus phytase -7 eluted as a homogeneous symmetrical peak and was shown by SDS-PAGE to be approx. 95% pure.

Example 8

Expression of the consensus phytase genes in *Aspergillus niger*

The Bluescript-plasmids pBsk \overline{fcp} , pBSK $\overline{fcp10}$, and pBsk $\overline{fcp7}$ were used as template for the introduction of a *Bsp* HI-site upstream of the start codon of the genes and an *Eco* RV-site downstream of the stop codon. The Expand™ High Fidelity PCR Kit (Boehringer Mannheim, Mannheim, Germany) was used with the following primers:

Primer Asp-1:

Bsp HI

5'-TATATCATGAGCGTGTTCGTCGTGCTACTGTTC-3'

Primer Asp-2 used for cloning of *fcp* and *fcp7*:

5

Eco RV

3'-ACCCGACTTACAAAGCGAATTCTATAGATATAT-5'

Primer Asp-3 used for cloning of *fcp10*:

Eco RV

3'-ACCCTTCTTACAAAGCGAATTCTATAGATATAT-5'

- 10 The reaction was performed as described by the supplier. The PCR-amplified *fcp*-genes had a new *Bsp* HI site at the start codon, introduced by primer Asp-1, which resulted in a replacement of the second amino acid residue glycine by serine. Subsequently, the DNA-fragment was digested with *Bsp* HI and *Eco* RV and ligated into the *Nco* I site downstream of the glucoamylase promoter of *Aspergillus niger* (*glaA*) and the *Eco* RV site
- 15 upstream of the *Aspergillus nidulans* tryptophan C terminator (*trpC*) (Mullaney *et al.*, 1985). After this cloning step, the genes were sequenced to detect possible failures introduced by PCR. The resulting expression plasmids which basically corresponds to the pGLAC vector as described in Example 9 of EP 684 313, contained the orotidine-5'-phosphate decarboxylase gene (*pyr4*) of *Neurospora crassa* as a selection marker.
- 20 Transformation of *Aspergillus niger* and expression of the consensus phytase genes was done as described in EP 684 313. The consensus phytases were purified as described in Example 7.

Example 9

Determination of phytase activity and of temperature optimum

- 25 Phytase activity was determined basically as described by Mitchell *et al* (1997). The activity was measured in an assay mixture containing 0.5% phytic acid (≈ 5 mM) in 200 mM sodium acetate, pH 5.0. After 15 min of incubation at 37 °C, the reaction was stopped by addition of an equal volume of 15% trichloroacetic acid. The liberated phosphate was quantified by mixing 100 μ l of the assay mixture with 900 μ l H₂O and 1 ml of 0.6 M
- 30 H₂SO₄, 2% ascorbic acid and 0.5% ammonium molybdate. Standard solutions of potassium phosphate were used as reference. One unit of enzyme activity was defined as the amount of enzyme that releases 1 μ mol phosphate per minute at 37 °C. The protein

concentration was determined using the enzyme extinction coefficient at 280 nm calculated according to Pace et al (1995): consensus phytase, 1.101; consensus phytase 7, 1.068; consensus phytase 10, 1.039.

5 In case of pH-optimum curves, purified enzymes were diluted in 10 mM sodium acetate, pH 5.0. Incubations were started by mixing aliquots of the diluted protein with an equal volume of 1% phytic acid (≈ 10 mM) in a series of different buffers: 0.4 M glycine/HCl, pH 2.5; 0.4 M acetate/NaOH, pH 3.0, 3.5, 4.0, 4.5, 5.0, 5.5; 0.4 M imidazole/HCl, pH 6.0, 6.5; 0.4 M Tris/HCl pH 7.0, 7.5, 8.0, 8.5, 9.0. Control experiments showed that pH was only slightly affected by the mixing step. Incubations were performed for 15 min at 37 °C as
10 described above.

For determinations of the substrate specificities of the phytases, phytic acid in the assay mixture was replaced by 5 mM concentrations of the respective phosphate compounds. The activity tests were performed as described above.

For determination of the temperature optimum, enzyme (100 μ l) and substrate
15 solution (100 μ l) were pre-incubated for 5 min at the given temperature. The reaction was started by addition of the substrate solution to the enzyme. After 15 min incubation, the reaction was stopped with trichloroacetic acid and the amount of phosphate released was determined.

The pH-optimum of the original consensus phytase was around pH 6.0-6.5 (70
20 U/mg). By introduction of the Q50T mutation, the pH-optimum shifted to pH 6.0 (130 U/mg). After introduction of K91A, the pH optimum shifted one pH-unit into the acidic pH-range showing a higher specific activity between pH 2.5 and pH 6.0. That was shown for the stabilized mutants and for consensus phytase-10, too (Figure 14 and 15).

Consensus phytase-7, which was constructed to transfer the catalytic characteristics of the
25 *A. niger* phytase NRRL 3135 into the consensus phytase, had a pH-profile which is shifted into the acidic range of the pH-spectrum showing an optimum between pH 4.5 and 5.0 (see Figure 19). The enzyme had a broad pH-optimum reaching at least 60% of its increased maximum activity from pH 2.5 to pH 6.0. The substrate spectrum, too, resemble more to that of the *A. niger* NRRL 3135 phytase than to the consensus phytase-1.

30 The temperature optimum of consensus phytase-1 (71 °C) was 16-26 °C higher than the temperature optimum of the wild-type phytases (45-55 °C, Table 7) which were used to calculate the consensus sequence. The improved consensus phytase-10 showed a further

increase of its temperature optimum to 80 °C (Figure 11). The temperature optimum of the consensus phytase-1-thermo[8] was found in the same range (78 °C) using the supernatant of an overproducing *S. cerevisiae* strain. The highest temperature optimum reached of 82 °C was determined for consensus phytase-10-thermo-Q50T-K91A.

- 5 **Table 7:** Temperature optimum and T_m -value of consensus phytase and of the phytases from *A. fumigatus*, *A. niger*, *E. nidulans*, and *M. thermophila*. The determination of the temperature optimum was performed as described in Example 9. The T_m -values were determined by differential scanning calorimetry as described in Example 10.

| phytase | temperature optimum [°C] | T_m [°C] |
|--|--------------------------|------------|
| Consensus phytase-10-thermo-Q50T-K91A | 82 | 89.3 |
| Consensus phytase-10-thermo-Q50T | 82 | 88.6 |
| Consensus phytase-10 | 80 | 85.4 |
| Consensus phytase-1-thermo[8]-Q50T | 78 | 84.7 |
| Consensus phytase-1-thermo[8]-Q50T-K91A | 78 | 85.7 |
| Consensus phytase-1 | 71 | 78.1 |
| <i>A. niger</i> NRRL3135 | 55 | 63.3 |
| <i>A. fumigatus</i> 13073 | 55 | 62.5 |
| <i>A. fumigatus</i> 13073 α -mutant | 60 | 67.0 |
| <i>A. fumigatus</i> 13073 α -mutant (optimized) | 63 | - |
| <i>A. terreus</i> 9A-1 | 49 | 57.5 |
| <i>A. terreus</i> cbs.116.46 | 45 | 58.5 |
| <i>E. nidulans</i> | 45 | 55.7 |
| <i>M. thermophila</i> | 55 | n. d. |
| <i>T. thermophilus</i> | 45 | n. d. |

Example 10

Determination of the melting point by differential scanning calorimetry (DSC)

In order to determine the unfolding temperature of the phytases, differential scanning
5 calorimetry was applied as previously published by Brugger et al (1997). Solutions of 50-
60 mg/ml homogeneous phytase were used for the tests. A constant heating rate of 10 °
C/min was applied up to 90-95 °C.

The determined melting points reflect the results obtained for the temperature
optimums (Table 7). The most stable consensus phytase designed is consensus phytase-10-
10 thermo-Q50T-K91A showing a melting temperature under the choosen condition of 89.3 °
C. This is 26 to 33.6 °C higher than the melting point of the wild-type phytases used.

Example 11

Transfer of basidiomycete phytase active site into consensus phytase-10-thermo- Q50T-K91A

15 As described previously (Example 3), mutations derived from the basidiomycete
phytase active site were introduced into the consensus phytase 10. The following five
constructs a) to e) were prepared:

- a) This construct is called consensus phytase 12, and it comprises a selected number of
active site residues of the basidio consensus sequence, its amino acid sequence
20 (consphy12) is shown in Fig. 21 (the first 26 amino acids forms the signal peptide,
amended positions are underlined);
- b) a cluster of mutations (Cluster II) was transferred to the consensus 10 sequence, viz.:
S80Q, Y86F, S90G, K91A, S92A, K93T, A94R, Y95I;
- c) analogously, another cluster of mutations (Cluster III) was transferred, viz.: T129V,
25 E133A, Q143N, M136S, V137S, N138Q, S139A;
- d) analogously, a further cluster of mutations (Cluster IV) was transferred, viz.: A168D,
E171T, K172N, F173W;

e) and finally, a further cluster of mutations (Cluster V) was transferred, viz.: Q297G, S298D, G300D, Y305T.

These constructs were expressed as described in Examples 6 to 8.

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Claims

1. A process for the preparation of a consensus protein, whereby such process is characterized by the following steps:

- 5 a) at least three, preferably four amino acid sequences are aligned by any standard alignment program known in the art,
- b) amino acids at the same position according to such alignment are compared regarding their evolutionary similarity by any standard program known in the art, whereas the degree of similarity provided by such a program which defines the least similarity of the amino acids that is used for the determination of an amino acid of corresponding positions is set to a less stringent number and the parameters are set in such a way that it is possible for the program to determine from only 2 identical amino acids at a corresponding position an amino acid for the consensus protein; however, if among the compared amino acid sequences are sequences that show a much higher degree of similarity to each other than to the residual sequences, these sequences are represented by their consensus sequence determined as defined in the same way as in the present process for the consensus sequence of the consensus protein or a vote weight of 1 divided by the number of such sequences is assigned to every of -those sequences,
- 10 c) in case no common amino acid at a defined position is identified by the program, any of the amino acids, preferably the most frequent amino acid of all such sequences is selected,
- 20 d) once the consensus sequence has been defined, such sequence is back-translated into a DNA sequence, preferably by using a codon frequency table of the organism in which expression should take place,
- 25 e) the DNA sequence is synthesized by methods known in the art and used either integrated into a suitable expression vector or by itself to transform an appropriate host cell,
- f) the transformed host cell is grown under suitable culture conditions and the consensus protein is isolated from the host cell or its culture medium by methods known in the art.
- 30

2. A process as claimed in claim 1 wherein the program used for the comparison of amino acids at a defined position regarding their evolutionary similarity is the program "PRETTY".

3. A process as claimed in claims 1 or 2, wherein

in a first step a consensus sequence is determined from a number of highly homologous sequences according to steps a), b) and c) of claim 1,

in a second step the amino acid sequence of another protein which is homologous to the
5 consensus sequence is compared with the consensus sequence and

in a third step only those amino acid residues are replaced in the amino acid sequence of the other protein which clearly differ from the consensus sequence of this protein family calculated under moderately stringent conditions whereas at all positions of the alignment where no preferred single amino acid can be determined under moderately stringent
10 conditions the amino acids of the other protein remain unchanged.

4. A process as claimed in any one of claims 1-3, wherein

in a first step a consensus sequence is determined from homologous sequences according to steps a), b) and c) of claim 1,

in a second step the active center of the protein comprising all amino acid residues that are
15 involved in forming the active center is determined in the consensus sequence and in the sequence of a homologous protein as well and

in a third step some or all of the amino acids that form the active center of the homologous protein are inserted in the backbone of the consensus sequence.

5. A process as claimed in claim 4, wherein the active center of the protein is
20 determined by using an analysis of the three-dimensional structure of the protein.

6. A process as claimed in claims 4 and 5, wherein the homologous protein is an enzyme.

7. A process as claimed in claims 1 to 6, wherein the defined protein family is the family of phytases.

25 8. A process as claimed in claim 7, wherein the phytases are of fungal origin.

9. A process as claimed in claims 7 or 8, wherein the amino acid sequence of the phytase is changed by the introduction of at least one mutation selected from the group consisting of

| | |
|-------|-------|
| E58A | F54Y |
| D69K | I73V |
| D197N | K94A |
| T214L | R101A |
| E222T | N153K |
| E267D | V158I |
| R291I | A203G |
| R329H | S205G |
| S364T | V217A |
| A379K | A227V |
| G404A | V234L |
| | P238A |
| | Q277E |
| | A287H |
| | A292Q |
| | V366I |
| | A396S |
| | E415Q |
| | G437A |
| | E451R |

whereby the number represents the position in the consensus phytase sequence or a corresponding residue according to an alignment as shown in Fig. 1 when 26 amino acids (signal sequence) are added to the sequences shown in Fig. 1 and the letter before the number represents the amino acid in the phytase which is replaced by the amino acid
5 behind the number.

10. A process as claimed in any one of claims 1 to 9, wherein the host cell is of eukaryotic origin.

11. A process as claimed in claim 10, wherein eukaryotic means fungal, preferably *Aspergillus* or yeast, preferably *Saccharomyces* or *Hansenula*.

12. A consensus protein obtainable preferably obtained by a process as claimed in any one of claims 1 to 11.

5 13. A consensus protein which comprises the amino acid sequence shown in Figure 2 or any variants or muteins thereof (consensus phytase-1).

14. A mutein of the consensus protein of claim 13 characterized therein that in the amino acid sequence of Figure 2 the following replacements have been effected Q50L, Q50T, Q50G, Q50T-Y51N, Q50L-Y51N or Q50T-K91A.

10 15. A consensus protein which comprises the amino acid sequence shown in Figure 4 having the designation consensus phytase 10 (Fcp10) and any variants or muteins thereof.

16. A consensus protein which comprises the amino acid sequence shown in Figure 6 having the designation Consensus seq. 11 and any variant or mutein thereof.

15 17. A consensus protein which comprises the amino acid sequence shown in Figure 10 (consensus phytase 7) and any variant or mutein thereof.

18. A consensus protein which comprises the amino acid sequence shown in Figure 21 (consensus phytase 12) and any variant or mutein thereof.

19. A consensus protein which comprises the amino acid sequence shown in Figure 3 (basidio consensus) and any variant or mutein thereof.

20 20. A phytase being selected from amongst: *A. fumigatus* ATCC 13073 alpha-mutant; *A. fumigatus* ATCC 13073 alpha-mutant-(E59A-S126N-R329H, S364T-G404A)-Q27T; *A. fumigatus* ATCC 13073 alpha-mutant-(E59A, S126N-R329H-S364T-G404A)-Q27T-K68A, preferably the latter.

25 21. A food, feed or pharmaceutical composition comprising a consensus protein as claimed in any of the claims 12 to 17.

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Figure 1

| | | |
|-----|-----------------------|----------------------------------|
| | | 1 |
| 50 | | |
| 5 | A. terreus 9A-1 | KhsDCNSVDh GYQCFPELSH kWGLYAPYFS |
| | LQDESPFPID VPEDChITFV | |
| | A. terreus cbs | NhsDCTSVDr GYQCFPELSH kWGLYAPYFS |
| | LQDESPFPID VPDDChITFV | |
| | A. niger var. awamori | NqsTCDTVDQ GYQCFSETSH LWGQYAPFFS |
| 10 | LANESAISPD VPAGCrVTFA | |
| | A. niger T213 | NqsSCDTVDQ GYQCFSETSH LWGQYAPFFS |
| | LANESVISPD VPAGCrVTFA | |
| | A. niger NRRL3135 | NqsSCDTVDQ GYQCFSETSH LWGQYAPFFS |
| | LANESVISPE VPAGCrVTFA | |
| 15 | A. fumigatus 13073 | GskSCDTVDl GYQCsPATSH LWGQYSPFFS |
| | LEDELSVSSK LPKDCrITLV | |
| | A. fumigatus 32722 | GskSCDTVDl GYQCsPATSH LWGQYSPFFS |
| | LEDELSVSSK LPKDCrITLV | |
| | A. fumigatus 58128 | GskSCDTVDl GYQCsPATSH LWGQYSPFFS |
| 20 | LEDELSVSSK LPKDCrITLV | |
| | A. fumigatus 26906 | GskSCDTVDl GYQCsPATSH LWGQYSPFFS |
| | LEDELSVSSK LPKDCrITLV | |
| | A. fumigatus 32239 | GskACDTVEl GYQCsPGTSH LWGQYSPFFS |
| | LEDELSVSSD LPKDCrVTFV | |
| 25 | E. nidulans | QNHSCNTADG GYQCFPNVSH VWGQYSPYFS |
| | IEQESAISd VPHGCeVTFV | |
| | T. thermophilus | DSHSCNTVEG GYQCrPEISH sWGQYSPFFS |
| | LADQSEISPD VPQNCKITFV | |
| | M. thermophila | ESRPCDTpDl GFQCgTAISH FWGQYSPYFS |
| 30 | VpSElDaS.. IPDDCeVTFA | |
| | Consensus | NSHSCDTVDG GYQCFPEISH LWGQYSPYFS |
| | LEDESAISPD VPDDC-VTFV | |
| | Consensus phytase | NSHSCDTVDG GYQCFPEISH LWGQYSPYFS |
| 35 | LEDESAISPD VPDDCrVTFV | |
| | | 51 |
| 100 | | |
| 40 | A. terreus 9A-1 | QVLARHGARs PThSKtKAYA AtIAAIQKSA |
| | TaFpGKYAFL QSYNYSLDSE | |
| | A. terreus cbs | QVLARHGARs PTDSKtKAYA AtIAAIQKNA |
| | TaLpGKYAFL KSYNYSMGSE | |
| | A. niger var. awamori | QVLSRHGARY PTESKgKkYS ALIEEIQQNV |
| 45 | TtFDGKYAFL KTYNYSLGAD | |
| | A. niger T213 | QVLSRHGARY PTESKgKkYS ALIEEIQQNV |
| | TtFDGKYAFL KTYNYSLGAD | |
| | A. niger NRRL3135 | QVLSRHGARY PTDSKgKkYS ALIEEIQQNA |
| | TtFDGKYAFL KTYNYSLGAD | |
| 50 | A. fumigatus 13073 | QVLSRHGARY PTSSKsKkYK kLVTAIQaNA |
| | TdFKGKFAFL KTYNYTLGAD | |
| | A. fumigatus 32722 | QVLSRHGARY PTSSKsKkYK kLVTAIQaNA |
| | TdFKGKFAFL KTYNYTLGAD | |
| | A. fumigatus 58128 | QVLSRHGARY PTSSKsKkYK kLVTAIQaNA |
| 55 | TdFKGKFAFL KTYNYTLGAD | |

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- 50 -

A. fumigatus 26906 QVLSRHGARY PTSSKsKkYK kLVTAIQaNA
 TdFKGKFAFL KTYNYTLGAD
A. fumigatus 32239 QVLSRHGARY PTASKsKkYK kLVTAIQKNA
 TeFKGKFAFL ETYNYTLGAD
 5 *E. nidulans* QVLSRHGARY PTESKsKAYS GLIEAIQKNA
 TsFwGQYAFI ESYNITLGAD
T. thermophilus QLLSRHGARY PTSSKtELYS QLISrIQKTA
 TaYKGyYAFI KDYrYqLGAN
M. thermophila QVLSRHGARA PtlKRaaSYv DLIDrIHhGA
 10 IsYgPgYEFL RTYDYTLGAD

 Consensus QVLSRHGARY PTSSK-KAYS ALIEAIQKNA T-
 FKGYAFI KTYNYTLGAD
 Consensus phytase QVLSRHGARY PTSSKSKAYS ALIEAIQKNA
 15 TAFKGYAFI KTYNYTLGAD

101

150
 20 *A. terreus* 9A-1 ELTPFGrNQL rDlGaQFYeR YNALTRhInP
 FVRATDASRV hESAekFVEG
A. terreus cbs NLTPFGrNQL qDlGaQFYRR YDTLTRhInP
 FVRAADSSRV hESAekFVEG
A. niger var. *awamori* DLTPFGEQEL VNSGIKFYQR YESLTRNIIP
 25 FIRSSGSSRV IASGEKFIEG
A. niger T213 DLTPFGEQEL VNSGIKFYQR YESLTRNIIP
 FIRSSGSSRV IASGEKFIEG
A. niger NRRL3135 DLTPFGEQEL VNSGIKFYQR YESLTRNIVP
 FIRSSGSSRV IASGKKFIEG
 30 *A. fumigatus* 13073 DLTPFGEQQL VNSGIKFYQR YKALARSVVP
 FIRASGSDRV IASGEKFIEG
A. fumigatus 32722 DLTPFGEQQL VNSGIKFYQR YKALARSVVP
 FIRASGSDRV IASGEKFIEG
A. fumigatus 58128 DLTPFGEQQL VNSGIKFYQR YKALARSVVP
 35 FIRASGSDRV IASGEKFIEG
A. fumigatus 26906 DLTAfGEQQL VNSGIKFYQR YKALARSVVP
 FIRASGSDRV IASGEKFIEG
A. fumigatus 32239 DLTPFGEQQM VNSGIKFYQK YKALAgSVVP
 FIRSSGSDRV IASGEKFIEG
 40 *E. nidulans* DLTiFGENQM VDsgaKFYRR YKNLARKnTP
 FIRASGSDRV VASAEKFIEG
T. thermophilus DLTPFGENQM IQlGIKFYnH YKSLARNAVP
 FVRCSGSDRV IASGrIFIEG
M. thermophila ELTRtGQQQM VNSGIKFYRR YRALARKsIP
 45 FVRTAGqDRV VhSAENFTQG

 Consensus DLTPFGENQM VNSGIKFYRR YKALARK-VP
 FVRASGSDRV IASAEKFIEG
 Consensus phytase DLTPFGENQM VNSGIKFYRR YKALARKIVP
 50 FIRASGSDRV IASAEKFIEG

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| | | |
|----|-------------------------------------|----------------------------------|
| | | 151 |
| | 200 | |
| | <i>A. terreus</i> 9A-1 | FQTARqDDHh ANpHQSPPrV DVaIPEGSAY |
| | NNTLEHSICT AFES...STV | |
| 5 | <i>A. terreus</i> cbs | FQNARqGDPH ANpHQSPPrV DVVIPEGTAY |
| | NNTLEHSICT AFEA...STV | |
| | <i>A. niger</i> var. <i>awamori</i> | FQSTKLkDPr AqpgQSSPkI DVVISEASSs |
| | NNTLDPGTCT VFED...SEL | |
| | <i>A. niger</i> T213 | FQSTKLkDPr AqpgQSSPkI DVVISEASSs |
| 10 | NNTLDPGTCT VFED...SEL | |
| | <i>A. niger</i> NRRL3135 | FQSTKLkDPr AqpgQSSPkI DVVISEASSs |
| | NNTLDPGTCT VFED...SEL | |
| | <i>A. fumigatus</i> 13073 | FQqAKLADPG A.TNRAAPAI SVIIPESETF |
| | NNTLDHGVCT kFEA...SQL | |
| 15 | <i>A. fumigatus</i> 32722 | FQqAKLADPG A.TNRAAPAI SVIIPESETF |
| | NNTLDHGVCT kFEA...SQL | |
| | <i>A. fumigatus</i> 58128 | FQqAKLADPG A.TNRAAPAI SVIIPESETF |
| | NNTLDHGVCT kFEA...SQL | |
| | <i>A. fumigatus</i> 26906 | FQqAKLADPG A.TNRAAPAI SVIIPESETF |
| 20 | NNTLDHGVCT kFEA...SQL | |
| | <i>A. fumigatus</i> 32239 | FQqANVADPG A.TNRAAPVI SVIIPESETY |
| | NNTLDHSVCT NFEA...SEL | |
| | <i>E. nidulans</i> | FRKAQLhDHG S..gQATPVV NVIIPeIDGF |
| | NNTLDHSTCV SFEN...DER | |
| 25 | <i>T. thermophilus</i> | FQSAKvldPh SDkHDAPPTI NVIIeEGPSY |
| | NNTLDtGSCP VFED...SSg | |
| | <i>M. thermophila</i> | FHSALLADRG STvRPTlPyd mVVIPETAGa |
| | NNTLHNDlCT AFEEgpySTI | |
| 30 | Consensus | FQSAKLADPG S-PHQASPVI NVIIPESGy |
| | NNTLDHGTCT AFED---SEL | |
| | Consensus phytase | FQSAKLADPG SQPHQASPVI DVIIPESGy |
| | NNTLDHGTCT AFED...SEL | |
| 35 | | |
| | | 201 |
| | 250 | |
| | <i>A. terreus</i> 9A-1 | GDDAvANFTA VFAPAIaQRL EADLPGVqLS |
| | TDDVvnlMAM CPFETVSlTD | |
| 40 | <i>A. terreus</i> cbs | GDAAADNFTA VFAPAIakRL EADLPGVqLS |
| | ADDVvnlMAM CPFETVSlTD | |
| | <i>A. niger</i> var. <i>awamori</i> | ADTVEANFTA TFAPSIRQRL ENDLsgVTLT |
| | DTEVTyLMDM CSFDTIstST | |
| | <i>A. niger</i> T213 | ADTVEANFTA TFAPSIRQRL ENDLsgVTLT |
| 45 | DTEVTyLMDM CSFDTIstST | |
| | <i>A. niger</i> NRRL3135 | ADTVEANFTA TFVPSIRQRL ENDLsgVTLT |
| | DTEVTyLMDM CSFDTIstST | |
| | <i>A. fumigatus</i> 13073 | GDEVAANFTA lFAPDIRARA EkHLPGVTLT |
| | DEDVVsLMDM CSFDTVARTS | |
| 50 | <i>A. fumigatus</i> 32722 | GDEVAANFTA lFAPDIRARA EkHLPGVTLT |
| | DEDVVsLMDM CSFDTVARTS | |
| | <i>A. fumigatus</i> 58128 | GDEVAANFTA lFAPDIRARA EkHLPGVTLT |
| | DEDVVsLMDM CSFDTVARTS | |
| | <i>A. fumigatus</i> 26906 | GDEVAANFTA lFAPDIRARA KkHLPGVTLT |
| 55 | DEDVVsLMDM CSFDTVARTS | |
| | <i>A. fumigatus</i> 32239 | GDEVEANFTA lFAPAIRARI EkHLPGVqLT |
| | DDDVVsLMDM CSFDTVARTA | |

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E. nidulans      ADEiEANFTA IMGPPiRkRL ENDLPGIKLT
NENViYLMDM CSFDTMARTA
T. thermophilus  GHDAQEKFAK qFAPAiLEKI KDHLPGVDLA
vSDVpyLMDL CPFETLARNh
5 M. thermophila  GDDAQDTYiS TFAGPiTARV NANLPGANLT
DADTVaLMDL CPFETVAsSS

Consensus        GDDAEANFTA TFAPAiRARL EADLPGVTiLT DEDVV-
LMDM CPFETVARTS
10 Consensus phytase  GDDVEANFTA LFAPAiRARL EADLPGVTiLT
DEDVVYLMDM CPFETVARTS

251
15 300
A. terreus 9A-1 ..... DAhTLSPFC DLFTATeWtq
YNYLlSLDKY YGYGGGNPLG
A. terreus cbs ..... DAhTLSPFC DLFTAaEWtq
YNYLlSLDKY YGYGGGNPLG
20 A. niger var. awamori ..... vDTKLSPFC DLFTHdEWih
YDYlQSLkKY YGHGAGNPLG
A. niger T213 ..... vDTKLSPFC DLFTHdEWih
YDYlRSLkKY YGHGAGNPLG
A. niger NRRL3135 ..... vDTKLSPFC DLFTHdEWin
YDYlQSLkKY YGHGAGNPLG
25 A. fumigatus 13073 ..... DASQLSPFC QLFTHnEWkk
YNYLQSLGKY YGYGAGNPLG
A. fumigatus 32722 ..... DASQLSPFC QLFTHnEWkk
YNYLQSLGKY YGYGAGNPLG
30 A. fumigatus 58128 ..... DASQLSPFC QLFTHnEWkk
YNYLQSLGKY YGYGAGNPLG
A. fumigatus 26906 ..... DASQLSPFC QLFTHnEWkk
YNYLQSLGKY YGYGAGNPLG
A. fumigatus 32239 ..... DASELSPFC AIFTHnEWkk
35 YDYlQSLGKY YGYGAGNPLG
E. nidulans ..... HGTELSPFC AIFTEkEWlq
YDYlQSLSKY YGYGAGSPLG
T. thermophilus ..... TDT.LSPFC ALStQeEWqa
YDYYQSLGKY YGnGGGNPLG
40 M. thermophila  sdpatadagg gNGrPLSPFC rLFSEsEWra
YDYlQSVGKW YGYGPGNPLG

Consensus        ----- DATELSPFC ALFTE-EW--
YDYlQSLGKY YGYGAGNPLG
45 Consensus phytase ..... DATELSPFC ALFTHDEWRQ
YDYlQSLGKY YGYGAGNPLG

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| | | | |
|----|-------------------------------------|------------|-----------------------|
| | 350 | | 301 |
| | <i>A. terreus</i> 9A-1 | PVQGVGWaNE | LMARLTRAPV HDHTCVNNTL |
| | DASPATFPLN ATLYADFSHD | | |
| 5 | <i>A. terreus</i> cbs | PVQGVGWaNE | LIARLTRSPV HDHTCVNNTL |
| | DANPATFPLN ATLYADFSHD | | |
| | <i>A. niger</i> var. <i>awamori</i> | PTQGVGYaNE | LIARLTHSPV HDDTSSNHTL |
| | DSNPATFPLN STLYADFSHD | | |
| | <i>A. niger</i> T213 | PTQGVGYaNE | LIARLTHSPV HDDTSSNHTL |
| 10 | DSNPATFPLN STLYADFSHD | | |
| | <i>A. niger</i> NRRL3135 | PTQGVGYaNE | LIARLTHSPV HDDTSSNHTL |
| | DSSPATFPLN STLYADFSHD | | |
| | <i>A. fumigatus</i> 13073 | PAQGIGFtNE | LIARLTRSPV QDHTSTNStL |
| | vSNPATFPLN ATMYVDFSHD | | |
| 15 | <i>A. fumigatus</i> 32722 | PAQGIGFtNE | LIARLTRSPV QDHTSTNStL |
| | vSNPATFPLN ATMYVDFSHD | | |
| | <i>A. fumigatus</i> 58128 | PAQGIGFtNE | LIARLTRSPV QDHTSTNStL |
| | vSNPATFPLN ATMYVDFSHD | | |
| | <i>A. fumigatus</i> 26906 | PAQGIGFtNE | LIARLTRSPV QDHTSTNStL |
| 20 | vSNPATFPLN ATMYVDFSHD | | |
| | <i>A. fumigatus</i> 32239 | PAQGIGFtNE | LIARLTNSPV QDHTSTNStL |
| | DSDPATFPLN ATLYVDFSHD | | |
| | <i>E. nidulans</i> | PAQGIGFtNE | LIARLTQSPV QDNTSTNHTL |
| | DSNPATFPLD rKLYADFSHD | | |
| 25 | <i>T. thermophilus</i> | PAQGVGFvNE | LIARMTSPV QDYTTVNHTL |
| | DSNPATFPLN ATLYADFSHD | | |
| | <i>M. thermophila</i> | PTQGVGFvNE | LLARLAgvPV RDgTSTNRTL |
| | DGDPrTFPLG rPLYADFSHD | | |
| 30 | Consensus | PAQGVGF-NE | LIARLTHSPV QDHTSTNHTL |
| | DSNPATFPLN ATLYADFSHD | | |
| | Consensus phytase | PAQGVGFANE | LIARLTRSPV QDHTSTNHTL |
| | DSNPATFPLN ATLYADFSHD | | |
| 35 | | | |
| | 400 | | 351 |
| | <i>A. terreus</i> 9A-1 | SNLVSIFWAL | GLYNGTAPLS qTSVESVSQT |
| | DGYAAAWTVP FAARAYVEMM | | |
| 40 | <i>A. terreus</i> cbs | SNLVSIFWAL | GLYNGTkPLS qTTVEDITrT |
| | DGYAAAWTVP FAARAYIEMM | | |
| | <i>A. niger</i> var. <i>awamori</i> | NGIISILFAL | GLYNGTkPLS TTTVENITQT |
| | DGFSSAWTVP FASRLYVEMM | | |
| | <i>A. niger</i> T213 | NGIISILFAL | GLYNGTkPLS TTTVENITQT |
| 45 | DGFSSAWTVP FASRLYVEMM | | |
| | <i>A. niger</i> NRRL3135 | NGIISILFAL | GLYNGTkPLS TTTVENITQT |
| | DGFSSAWTVP FASRLYVEMM | | |
| | <i>A. fumigatus</i> 13073 | NSMVSIFFAL | GLYNGTEPLS rTSVESaKEl |
| | DGYSASWVVP FGARAYFetM | | |
| 50 | <i>A. fumigatus</i> 32722 | NSMVSIFFAL | GLYNGTGPLS rTSVESaKEl |
| | DGYSASWVVP FGARAYFetM | | |
| | <i>A. fumigatus</i> 58128 | NSMVSIFFAL | GLYNGTEPLS rTSVESaKEl |
| | DGYSASWVVP FGARAYFetM | | |
| | <i>A. fumigatus</i> 26906 | NSMVSIFFAL | GLYNGTEPLS rTSVESaKEl |
| 55 | DGYSASWVVP FGARAYFetM | | |
| | <i>A. fumigatus</i> 32239 | NGMIPIFFAM | GLYNGTEPLS qTSeESTKES |
| | NGYSASWAVP FGARAYFetM | | |

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E. nidulans NSMISIFFAM GLYNGTQPLS mDSVESIQEm
 DGYAASWTVP FGARAYFELM
T. thermophilus NTMTSIFaAL GLYNGTAKLS TTEIKSIEET
 DGYSAAWTVP FGGRAYIEMM
 5 *M. thermophila* NDMMGVLGAL GaYDGVPPLD KTArrDpEEI
 GGYAASWAVP FAARiYVEKM

 Consensus NSMISIFFAL GLYNGTAPLS TTSVESIEET
 DGYAASWTVP FGARAYVEMM
 10 Consensus phytase NSMISIFFAL GLYNGTAPLS TTSVESIEET
 DGYSASWTVP FGARAYVEMM

 401
 15 450
A. terreus 9A-1 QC..... RAEKE PLVRVLVNDR
 VMPLHGCPD KLGRCKrDAF
A. terreus cbs QC..... RAEKQ PLVRVLVNDR
 VMPLHGCAVD NLGRCKrDDF
 20 *A. niger* var. *awamori* QC..... QAEQE PLVRVLVNDR
 VVPLHGCPID aLGRCTrDSF
A. niger T213 QC..... QAEQE PLVRVLVNDR
 VVPLHGCPID aLGRCTrDSF
A. niger NRRL3135 QC..... QAEQE PLVRVLVNDR
 25 VVPLHGCPVD aLGRCTrDSF
A. fumigatus 13073 QC..... KSEKE PLVRALINDR
 VVPLHGCDVD KLGRCKLNDF
A. fumigatus 32722 QC..... KSEKE PLVRALINDR
 VVPLHGCDVD KLGRCKLNDF
 30 *A. fumigatus* 58128 QC..... KSEKE SLVRALINDR
 VVPLHGCDVD KLGRCKLNDF
A. fumigatus 26906 QC..... KSEKE PLVRALINDR
 VVPLHGCDVD KLGRCKLNDF
A. fumigatus 32239 QC..... KSEKE PLVRALINDR
 35 VVPLHGCAVD KLGRCKLNDF
E. nidulans QC..... E.KKE PLVRVLVNDR
 VVPLHGCAVD KFGRCTLDDW
T. thermophilus QC..... DDSDE PVVRVLVNDR
 VVPLHGCEVD SLGRCKrDDF
 40 *M. thermophila* RCsggggggg gggegrQEKDE eMVRVLVNDR
 VMTLkGCGAD ErGMCTLErF

 Consensus QC----- QAEKE PLVRVLVNDR
 VVPLHGCAVD KLGRCKLDDF
 45 Consensus phytase QC..... QAEKE PLVRVLVNDR
 VVPLHGCAVD KLGRCKRDDF

 451
 471
 50 *A. terreus* 9A-1 VAGLSFAQAG GNWADCF---
A. terreus cbs VEGLSFARAG
 GNWAECF---
A. niger var. *awamori* VrGLSFARSG GDWAECsA--
A. niger T213 VrGLSFARSG GDWAECFA--
 55 *A. niger* NRRL3135 VrGLSFARSG
 GDWAECFA--
A. fumigatus 13073 VKGLSWARSG GNWGECS--
A. fumigatus 32722 VKGLSWARSG GNWGECS--

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| | | |
|----|--------------------|-------------------------|
| | A. fumigatus 58128 | VKGLSWARSG GNWGECS-- - |
| | A. fumigatus 26906 | VKGLSWARSG GNWGECS-- - |
| | A. fumigatus 32239 | VKGLSWARSG |
| | GNSEQSFS-- - | |
| 5 | E. nidulans | VEGLNFARSG GNWkTCFTl~ - |
| | T. thermophilus | VrGLSFARqG GNWEGCYAas e |
| | M. thermophila | IESMAFARGN GKWDlCFA-- - |
| | Consensus | VEGLSFARSG GNWAECS-- - |
| 10 | Consensus phytase | VEGLSFARSG GNWAECS... . |

Figure 2

CP-1

15 Eco RI M G V F V V L L S I A T L F G S T
TATATGAATTCA~~TGGGCGTGTTCGTCGTC~~ACTGTCCATTGCCACCTTGTTCCGGTTCCA

1 -----+-----+-----+-----+-----+-----+ 60

ATATACTTAAGTACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAACAAGCCAAGGT

S G T A L G P R G N S H S C D T V D G G

20 CATCCGGTACCGCCTTGGGTCCTCGTGGTAATTCTCACTCTTGACACTGTTGACGGTG

61 -----+-----+-----+-----+-----+-----+ 120

GTAGGCCATGGCGGAACCCAGGAGCACCATTAAAGAGTGAGAACAACCTGTGACAACCTGCCAC

CP-2

CP-3

25 Y Q C F P E I S H L W G Q Y S P Y F S L
GTTACCAATGTTTCCAGAAATTTCTCACTTGTTGGGGTCAATACTCTCCATACTTCTCTT

121 -----+-----+-----+-----+-----+-----+ 180

CAATGGTTACAAAGGGTCTTTAAAGAGTGAACACCCAGTTATGAGAGGTATGAAGAGAA

E D E S A I S P D V P D D C R V T F V Q

30 TGGAAGACGAATCTGCTATTTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTCGTTT

181 -----+-----+-----+-----+-----+-----+ 240

ACCTTCTGCTTAGACGATAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAG

CP-4

CP-5

35 V L S R H G A R Y P T S S K S K A Y S A

AAGTTTGTCTAGACACGGTGCTAGATACCCAACTTCTTCTAAGTCTAAGGCTTACTCTG

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241 -----+-----+-----+-----+-----+-----+ 300

TTCAAAACAGATCTGTGCCACGATCTATGGGTTGAAGAAGATTGAGATTCCGAATGAGAC

L I E A I Q K N A T A F K G K Y A F L K

5 CTTTGATTGAAGCTATTCAAAGAACGCTACTGCTTCAAGGGTAAGTACGCTTTCTTGA

301 -----+-----+-----+-----+-----+-----+ 360

GAAACTAACTTCGATAAGTTTCTTGCGATGACGAAAGTTCCCATTTCATGCGAAAGAACT

CP-6

CP-7

10 T Y N Y T L G A D D L T P F G E N Q M V

AGACTTACAACCTACACTTTGGGTGCTGACGACTTGACTCCATTGCGTGAAAACCAAATGG

361 -----+-----+-----+-----+-----+-----+ 420

TCTGAATGTTGATGTGAAAACCCACGACTGCTGAACTGAGGTAAGCCACTTTTGGTTTACC

15 N S G I K F Y R R Y K A L A R K I V P F

TTAACTCTGGTATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCAT

421 -----+-----+-----+-----+-----+-----+ 480

AATTGAGACCATAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTA

CP-8

20

CP-9

I R A S G S D R V I A S A E K F I E G F

TCATTAGAGCTTCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTTCATTGAAGGTT

481 -----+-----+-----+-----+-----+-----+ 540

AGTAATCTCGAAGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAACTTCCAA

25

Q S A K L A D P G S Q P H Q A S P V I D

TCCAATCTGCTAAGTTGGCTGACCCAGGTTCTCAACCACACCAAGCTTCTCCAGTTATTG

541 -----+-----+-----+-----+-----+-----+ 600

AGGTTAGACGATTCAACCGACTGGGTCCAAGAGTTGGTGTGGTTCGAAGAGGTCAATAAC

30

CP-10

CP-11

V I I P E G S G Y N N T L D H G T C T A

ACGTTATTATTCCAGAAGGATCAGGTTACAACACACTTTGGACCACGGTACTTGTACTG

601 -----+-----+-----+-----+-----+-----+ 660

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TGCAATAATAAGGTCTTCctAGgCCAATGTTGTTGTGAAACCTGGTGCCATGAACATGAC

F E D S E L G D D V E A N F T A L F A P

CTTTCGAAGACTCTGAATTGGGTGACGACGTTGAAGCTAACTTCACTGCTTTGTTGCTC

5 661 -----+-----+-----+-----+-----+-----+ 720

GAAAGCTTCTGAGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGAAACAAGCGAG

CP-12

A I R A R L E A D L P G V T L T D E D V

10 CAGCTATTAGAGCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTTGACTGACGAAGACG

721 -----+-----+-----+-----+-----+-----+ 780

GTCGATAATCTCGATCTAACCTTCGACTGAACGGTCCACAATGAACTGACTGCTTCTGC

CP-13

15 V Y L M D M C P F E T V A R T S D A T E

TTGTTTACTTGATGGACATGTGTCCATTGAACTGTTGCTAGAACTTCTGACGCTACTG

781 -----+-----+-----+-----+-----+-----+ 840

AACAAATGAACTACCTGTACACAGGTAAGCTTTGACAACGATCTTGAAGACTGCGATGAC

20 L S P F C A L F T H D E W R Q Y D Y L Q

AATTGTCTCCATTCTGTGCTTTGTTCACTCACGACGAATGGAGACAATACGACTACTTGC

841 -----+-----+-----+-----+-----+-----+ 900

TTAACAGAGGTAAGACACGAAACAAGTGAGTGCTGCTTACCTCTGTTATGCTGATGAACG

CP-14

25

CP-15

S L G K Y Y G Y G A G N P L G P A Q G V

AATCTTTGGGTAAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTG

901 -----+-----+-----+-----+-----+-----+ 960

TTAGAAACCCATTTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCAC

30

G F A N E L I A R L T R S P V Q D H T S

TTGGTTTCGCTAACGAATTGATTGCTAGATTGACTAGATCTCCAGTTCAAGACCACACTT

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961 -----+-----+-----+-----+-----+-----+
1020
AACCAAAGCGATTGCTTAACTAACGATCTAACTGATCTAGAGGTCAAGTTCTGGTGTGAA
CP-16
5 CP-17
T N H T L D S N P A T F P L N A T L Y A
CTACTAACCACACTTTGGACTCTAACCAGCTACTTTCCCATTTGAACGCTACTTTGTACG
1021 -----+-----+-----+-----+-----+-----+
1080
10 GATGATTGGTGTGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGCGATGAAACATGC
D F S H D N S M I S I F F A L G L Y N G
CTGACTTCTCTCACGACAACTCTATGATTCTATTTTCTTCGCTTTGGGTTTGTACAACG
1081 -----+-----+-----+-----+-----+-----+
15 1140
GACTGAAGAGAGTGCTGTTGAGATACTAAAGATAAAAGAAGCGAAACCCAAACATGTTGC
CP-18
CP-19
T A P L S T T S V E S I E E T D G Y S A
20 GTACTGCTCCATTGTCTACTACTTCTGTTGAATCTATTGAAGAAACTGACGGTTACTCTG
1141 -----+-----+-----+-----+-----+-----+
1200
CATGACGAGGTAACAGATGATGAAGACAACTTAGATAACTTCTTTGACTGCCAATGAGAC
25 S W T V P F G A R A Y V E M M Q C Q A E
CTTCTTGGACTGTTCCATTGCGTGCTAGAGCTTACGTTGAAATGATGCAATGTCAAGCTG
1201 -----+-----+-----+-----+-----+-----+
1260
GAAGAACCTGACAAGGTAAGCCACGATCTCGAATGCAACTTTACTACGTTACAGTTCGAC
30 CP-20
CP-21
K E P L V R V L V N D R V V P L H G C A
AAAAGGAACCATTGGTTAGAGTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG
1261 -----+-----+-----+-----+-----+-----+
35 1320

CTAGACCACCATTTGACCCGACTTACAAAGCGAATTCTTAAGTATAT

15

Figure 3

| | | | |
|----|---|--|--|
| | | 1 | |
| 50 | | | |
| 20 | <i>P. involutus</i> (phyA1) QInQVNIIQR | SvP.KnTAPt FPIPeseQrn WSPYSPYFPL AeYkAPPAGC | |
| | <i>P. involutus</i> (phyA2) EInQVNIIQR | SvP.RniAPK FSIPeseQrn WSPYSPYFPL AeYkAPPAGC | |
| | <i>T. pubescens</i> QInQVHIIQR | hiPlRdTSAc LdVTrDvQqs WSmYSPYFPA AtYvAPPASC | |
| 25 | <i>A. pediades</i> KitQVNIIQR | GgvvQaTfvQ pfFPpQiQds WAAYTPYYPV qaYtPPPkDC | |
| | <i>P. lycii</i> tVtQVNLIQR | StQfsfvAAQ LPIPaQntsn WGPYdPFFPV EpYaAPPEGC | |
| 30 | Basidio QVNIIQR | S-P-R-TAAQ LPIP-Q-Q-- WSPYSPYFPV A-Y-APPAGC QI- | |

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51

100

P. involutus (phyA1) HGARFPTSGA TTRIKAGLTK LQGVqnfTDA KFNFIksfky
dLGnsDLVPF

5 *P. involutus* (phyA2) HGARFPTSGA ATRIKAGLSK LQSVqnfTDP KFDFIksfTY
dLGtsDLVPF

T. pubescens HGARFPTSGA AKRIQTAVAK LKAAsnyTDP lLAFVtNyTY
sLGqDsLveL

10 *A. pediades* HGARFPTSGA GTRIQAaVKK LQSAktyTDP RLDFLtnTY
tLGhDDLVPF

P. lycii HGARWPTSGA rSRqvAAVAK IQmArpfTDP KYEFLnDfvY
kFGvADLLPF

15 *Basidio* HGARFPTSGA ATRIQAaVAK LQSA---TDP KLDfL-N-TY -LG-
DDLVPF

101

150

20 *P. involutus* (phyA1) GAaQSfDAGQ EAFARYSkLV SkNNLPFIRA dGSDRVVDSA
TNWTAGFAsA

P. involutus (phyA2) GAaQSfDAGl EvFARYSkLV SsDNLPFIRS dGSDRVVDTA
TNWTAGFAsA

25 *T. pubescens* GAtQSSEAGQ EAFTRYsLV SaDELpFVRA SGSDRVVATA
nNWTAGFAlA

A. pediades GAlQSSQAGE ETfQRYsflV SkENLPFVRA SSSNRVVDSA
TNWTEGFSaA

P. lycii GAnQShQTGt DmYTRYStLf egGDVPFVRA AGdQRVVDSS
TNWTAGFGdA

30 *Basidio* GA-QSSQAGQ EAFTRYs-LV S-DNLpFVRA SGSDRVVDSA
TNWTAGFA-A

35

151

200

P. involutus (phyA1) ShNTvqPkLn LILPQtGNDT LEDNMCPaAG DSDPQvNaWL
AVafPSITAR

40 *P. involutus* (phyA2) SrNAiqPkLd LILPQtGNDT LEDNMCPaAG ESDPQvDaWL
AsafPSVTAQ

| | | |
|----|---|--|
| | <i>T. pubescens</i> AqFAPPMTAR | SsNSitPvLs VIISEaGNDT LDDNMCPaAG DSDPQvNqWL |
| | <i>A. pediades</i> SIYGTPIAnR | ShHvlnPiLf VILSEslNDT LDDaMCPnAG sSDPQtGiWt |
| 5 | <i>P. lycii</i> GVFAPnITAR | SgETvlPtLq VVLqEeGNcT LcNNMCPnEv DGDest.tWL |
| | Basidio AVFAPPITAR | S-NT--P-L- VILSE-GNDT LDDNMCP-AG DSDPQ-N-WL |
| 10 | | |
| | 250 | 201 |
| 15 | <i>P. involutus</i> (phyA1) giPGsFeAFa | LNAAAPSVNL TDtDAfNLvs LCAFlTVSke kksdFcTLFE |
| | <i>P. involutus</i> (phyA2) giPGsFeAFa | LNAAAPGANL TDaDAfNLvs LCPFmTVSke qksdFcTLFE |
| | <i>T. pubescens</i> elQAE.dAFa | LNAGAPGANL TDtDTyNLlt LCPFETVatE rrSeFCDIYE |
| 20 | <i>A. pediades</i> .tPEEFaqFe | LNqqAPGANI TAAdvsNLip LCAFETivke tpSpFCNLf. |
| | <i>P. lycii</i> .tABEYvSYe | LNAAAPSANL SDsDAItLmd MCPFDTLsG naSpFCDLf. |
| 25 | Basidio AF- | LNAAAPGANL TD-DA-NL-- LCPFETVS-E --S-FCDLFE --PEEF- |
| | | |
| | 300 | 251 |
| 30 | | |
| | <i>P. involutus</i> (phyA1) NTQTNRtLDA | YgGDLDKfYG TGYGQeLGPV QGVGYVNELI ARLTnsAVRD |
| | <i>P. involutus</i> (phyA2) NTQTNRtLDA | YaGDLDKfYG TGYGQALGPV QGVGYINELL ARLTnsAVnD |
| 35 | <i>T. pubescens</i> HTQTNsTLDS | YnADLDKfYG TGYGQPLGPV QGVGYINELI ARLTaQnVsD |
| | <i>A. pediades</i> NTQTNRtLDS | YfGDLDKfYG TGYGQPLGPV QGVGYINELL ARLTemPVrd |
| 40 | <i>P. lycii</i> ETQTNRtLDS | YyyDLdkYYG TGpGNALGPV QGVGYVNELL ARLTgQAVRD |

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Basidio Y-GDLDFYFG TGYGQPLGPV QGVGYINELL ARLT-QAVRD
 NTQTNRTLDS

5

301

350

P. involutus (phyA1) SPvTFPLNKT FYADFSHDN1 MVAVFSAMGL FrQPAPLsTS
 vPNPwRTWrT

10 *P. involutus* (phyA2) APdTFPLNKT MYADFSHDN1 MVAVFSAMGL FrQSAPLsTS
 tPDPNRTWLT

T. pubescens SPeTFPLNRT LYADFSHDNQ MVAIFSAMGL FNQSAPLDPT
 tPDPaRTFLv

15 *A. pediades* SPtTFPLDRS IYADLSHDNQ MIAIFSAMGL FNQSSPLDPS
 fPNPKRTWVT

P. lycii dPaTFPLNRT FYADFSHDNt MVPIFAALGL FNaTA.LDPl
 kPDeNRLwVd

20 Basidio SP-TFPLNRT FYADFSHDNQ MVAIFSAMGL FNQSAPLDPS -
 PDPNRTWVT

351

400

25 *P. involutus* (phyA1) SsLVPFSGRM VVERLsC..f GT..... tkV
 RVLVQDqVQP

P. involutus (phyA2) SsVVPFSARM aVERLsC..a GT..... tkV
 RVLVQDqVQP

30 *T. pubescens* kKIVPFsARM VVERLdC..g GA..... qSv
 RLLVNDaVQP

A. pediades SRLtPFsARM VtERLlCqrd GTgsgggsri mrngnvqtfv
 RILVNDALQP

P. lycii SKLVPFSGHM tVEKLaC... sgkeaV
 RVLVNDaVQP

35 Basidio SKLVPFsARM VVERL-C--- GT-----V
 RVLVNDaVQP

40 401 441

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P. involutus (phyA1) LEFCGGDrNG lCTLakFVES QtFARsDGaG DFEKCFATSa ~
P. involutus (phyA2) LEFCGGDqDG lCALDkFVES QaYARsGGaG DFEKCLATTv ~
T. pubescens LAFCGADtsG vCTLDAFVES QaYARNDGEG DFEKCFAT-- ~
A. pediades LKFCGGDmDS lCTLEAFVES QkYAREDGQG DFEKCFD--- ~
5 *P. lycii* LEFCGG.vDG vCeLsAFVES QtYARENGQG DFAKCgfvPs e

Basidio LEFCGGD-DG -CTLDAFVES Q-YAREDGQG DFEKCFATP- -

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Figure 4

| | | | |
|----|---|---|--|
| | | 1 | |
| 50 | | | |
| 5 | <i>A. terreus</i> 9a1 VPeDCHITFV | KhsdCNSVDh GYQCfPELSH kWGLYAPYFS LqDESPFP1D | |
| | <i>A. terreus</i> cbs VPdDCHITFV | NhsdCtSVDr GYQCfPELSH kWGLYAPYFS LqDESPFP1D | |
| 10 | <i>A. niger</i> var. <i>awamori</i> VPaGCRVTFa | NqsTCDTVdG GYQCfSEtSH LWGQYAPFFS LANESAISPD | |
| | <i>A. niger</i> NRRL3135 VPaGCRVTFa | NqsSCDTVdG GYQCfSEtSH LWGQYAPFFS LANESvISPE | |
| | <i>A. fumigatus</i> 13073 LPkDCRITLV | GskSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDELSVSSK | |
| 15 | <i>A. fumigatus</i> 32722 LPkDCRITLV | GskSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDELSVSSK | |
| | <i>A. fumigatus</i> 58128 LPkDCRITLV | GskSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDELSVSSK | |
| 20 | <i>A. fumigatus</i> 26906 LPkDCRITLV | GskSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDELSVSSK | |
| | <i>A. fumigatus</i> 32239 LPkDCRVTFV | GskACDTVE1 GYQCSPGtSH LWGQYSPFFS LEDELSVSSD | |
| | <i>E. nidulans</i> VPhGCeVTFV | QNHSCNTaDG GYQCfPNVSH VWGQYSPYFS IEQESAISeD | |
| 25 | <i>T. thermophilus</i> VPqNCKITFV | DSHSCNTVEG GYQCrPEISH sWGQYSPFFS LADQSEISPD | |
| | <i>T. lanuginosa</i> VPkGCRVeFV | ----- ----nvDIAR hWGQYSPFFS LAEvSEISPA | |
| 30 | <i>M. thermophila</i> IPdDCeVTFa | ESRPCDTpD1 GFQCgTAISH FWGQYSPYFS VPSElDaS.. | |
| | Basidio pPaGCQIxqV | xSxPxrxTAA qLPipxQxqx xWSPYSPYFP VAXyxA.... | |
| 35 | Consensus GCRVTFV | NSHSCDTVdG GYQC-PEISH LWGQYSPFFS LADESAISPD VP- | |
| | Fcp10 VPKGCRVTFV | NSHSCDTVdG GYQCfPEISH LWGQYSPFFS LADESAISPD | |

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| | | | |
|-----|---|--|--|
| | | 51 | |
| 100 | | | |
| 5 | <i>A. terreus</i> 9a1 QSYNYSLDSE | QVLARHGARS PThSKTKaYA AtIaAIQKSA TaFpGKYAFL | |
| | <i>A. terreus</i> cbs KSYNYSMGSE | QVLARHGARS PTdSKTKaYA AtIaAIQKNA TaLpGKYAFL | |
| | <i>A. niger</i> var. <i>awamori</i> KTYNYSLGAD | QVLSRHGARY PTeSKGKKYS ALIeEIQQNv TtFDGKYAFL | |
| 10 | <i>A. niger</i> NRRL3135 KTYNYSLGAD | QVLSRHGARY PTdSKGKKYS ALIeEIQQNA TtFDGKYAFL | |
| | <i>A. fumigatus</i> 13073 KTYNYTLGAD | QVLSRHGARY PTSSSKKKYk kLVtAIQaNA TdFKGKFAFL | |
| 15 | <i>A. fumigatus</i> 32722 KTYNYTLGAD | QVLSRHGARY PTSSSKKKYk kLVtAIQaNA TdFKGKFAFL | |
| | <i>A. fumigatus</i> 58128 KTYNYTLGAD | QVLSRHGARY PTSSSKKKYk kLVtAIQaNA TdFKGKFAFL | |
| | <i>A. fumigatus</i> 26906 KTYNYTLGAD | QVLSRHGARY PTSSSKKKYk kLVtAIQaNA TdFKGKFAFL | |
| 20 | <i>A. fumigatus</i> 32239 ETYNITLGAD | QVLSRHGARY PTASKSKKYk kLVtAIQKNA TeFKGKFAFL | |
| | <i>E. nidulans</i> ESYNYTLGAD | QVLSRHGARY PTeSKSKaYS GLIeAIQKNA TsFwGQYAFL | |
| 25 | <i>T. thermophilus</i> KdYrYqLGAN | QLLSRHGARY PTSSKTELYS qLIsrIQKtA TaYKGyYAFL | |
| | <i>T. lanuginosa</i> RdYaYhLGAD | QVLSRHGARY PTAhKSEvYA ELLqrIQDtA TeFKGDFAFL | |
| | <i>M. thermophila</i> RTYDYTLGAD | QVLSRHGARA PtlkRAasYv DLIdrIHhGA isYgPgYEFL | |
| 30 | Basidio xnxtYxLGxD | NIIqRHGARF PTSGaAtRiq AaVakLQsax xxtDPKLDLFL | |
| | Consensus KTYNYTLGAD | QVLSRHGARY PTSSSKKKYS ALI-AIQKNA T-FKGKYAFL | |
| 35 | Fcp10 KTYNYTLGAD | QVLSRHGARY PTSSSKKKYS ALIEAIQKNA TAFKGKYAFL | |

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| | | | |
|----|-------------------------------------|--|--|
| | | 101 | |
| 5 | 150 | | |
| | <i>A. terreus</i> 9a1 | ELTPFGGrNQL rDlGaQFYeR YNAL.TRhIn PFVRATDAsR | |
| | VhESA EK FVE | | |
| | <i>A. terreus</i> cbs | NLTPFGGrNQL qDlGaQFYRR YDTL.TRhIn PFVRAADSsR | |
| | VhESA EK FVE | | |
| 10 | <i>A. niger</i> var. <i>awamori</i> | DLTPFGGEQEL VNSGIKFYQR YESL.TRnII PFIRSSGSsR | |
| | VIASGEKFIE | | |
| | <i>A. niger</i> NRRL3135 | DLTPFGGEQEL VNSGIKFYQR YESL.TRnIV PFIRSSGSsR | |
| | VIASGKKFIE | | |
| 15 | <i>A. fumigatus</i> 13073 | DLTPFGGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR | |
| | VIASGEKFIE | | |
| | <i>A. fumigatus</i> 32722 | DLTPFGGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR | |
| | VIASGEKFIE | | |
| | <i>A. fumigatus</i> 58128 | DLTPFGGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR | |
| | VIASGEKFIE | | |
| 20 | <i>A. fumigatus</i> 26906 | DLTAFGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR | |
| | VIASGEKFIE | | |
| | <i>A. fumigatus</i> 32239 | DLTPFGGEQQM VNSGIKFYQK YKAL.AgsVV PFIRSSGSDR | |
| | VIASGEKFIE | | |
| 25 | <i>E. nidulans</i> | DLTiFGENQM VDSGaKFYRR YKnL.ARknt PFIRASGSDR | |
| | VVASAEKFIN | | |
| | <i>T. thermophilus</i> | DLTPFGENQM IQlGIKFYnH YKSL.ARnaV PFVRCSGSDR | |
| | VIASGrIFIE | | |
| | <i>T. lanuginosa</i> | NLTRFGEEQM MESGrQFYHR YREq.AReIV PFVRAAGSAR | |
| | VIASAEfFnr | | |
| 30 | <i>M. thermophila</i> | ELTRtGQQQM VNSGIKFYRR YRAL.ARksI PFVRTAGqDR | |
| | VVhSAENftQ | | |
| | Basidio | DLvPFGAxQs sQAGqEaFtR YsxLvSxdnL PFVRASGSDR | |
| | VVDSAtNWtA | | |
| 35 | Consensus | DLTPFGGEQQM VNSGIKFYRR YKAL-AR-IV PFVRASGSDR | |
| | VIASAEKFIE | | |
| | Fcp10 | DLTPFGGEQQM VNSGIKFYRR YKAL.ARKIV PFVRASGSDR | |
| | VIASAEKFIE | | |

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| | | | |
|-----|---|---|--|
| | | 151 | |
| 200 | | | |
| 5 | <i>A. terreus</i> 9a1 TAFES...St | GFQTARqDDh hAnphQPSPr VDVaIPEGsA YNNTLEHSLC | |
| | <i>A. terreus</i> cbs TAFEa...St | GFQNARqGDP hAnphQPSPr VDVVIPEGtA YNNTLEHSIC | |
| | <i>A. niger</i> var. <i>awamori</i> TvFEd...SE | GFQSTKLkDP rAqpgQSSPk IDVVISEAs sNNTLDpGtC | |
| 10 | <i>A. niger</i> NRRL3135 TvFEd...SE | GFQSTKLkDP rAqpgQSSPk IDVVISEAs sNNTLDpGtC | |
| | <i>A. fumigatus</i> 13073 TkFEa...SQ | GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC | |
| 15 | <i>A. fumigatus</i> 32722 TkFEa...SQ | GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC | |
| | <i>A. fumigatus</i> 58128 TkFEa...SQ | GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC | |
| | <i>A. fumigatus</i> 26906 TkFEa...SQ | GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC | |
| 20 | <i>A. fumigatus</i> 32239 TnFEa...SE | GFQqANVADP gAt.nRAAPV ISVIIPESeT YNNTLDHSVC | |
| | <i>E. nidulans</i> vSFEn...dE | GFRKAQLhDh g.s.gQATPV VNVIPEidG FNNTLDHStC | |
| 25 | <i>T. thermophilus</i> PvFEd...Ss | GFQSAKVlDP hSdKhDAPPt INVIIeEGpS YNNTLDtGsC | |
| | <i>T. lanuginosa</i> PAaEe...Ap | GFQdAKdrDP rSnkdQAePV INVIISEETG sNNTLDgltC | |
| | <i>M. thermophila</i> TAFEegPySt | GFHSALLADR gStvrPTlPy dmVVIPETaG aNNTLHNDLC | |
| 30 | BasidioPxAG | GFaxA.....sxntxxPx LxVILSExg. .NDTLDDNMC | |
| | Consensus | GFQSAKLADP -A---QASPV INVIIPEG-G YNNTLDHGLC | |
| | TAFE--P-SE | | |
| 35 | Fcp10 | GFQSAKLADP GANPHQASPV INVIIPEGAG YNNTLDHGLC | |
| | TAFEE...SE | | |

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| | | | |
|----|--|-----|---|
| | <i>A. terreus</i> 9a1 MCPFETVSlT | | VGDDaVaNFT AVFAPAIaQR LEAdLPGVQL StDDVVNLMA |
| | <i>A. terreus</i> cbs MCPFETVSlT | | VGDAaADNFT AVFAPAIaKR LEAdLPGVQL SADDVVNLMA |
| 5 | <i>A. niger</i> var. <i>awamori</i> MCSFDTISs | | LADtVEANFT AtFAPSIRqR LEndLSGVtL TDtEVtyLMD |
| | <i>A. niger</i> NRRL3135 MCSFDTISs | | LADtVEANFT AtFvPSIRqR LEndLSGVtL TDtEVtyLMD |
| 10 | <i>A. fumigatus</i> 13073 MCSFDTVArT | | LGDEVAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD |
| | <i>A. fumigatus</i> 32722 MCSFDTVArT | | LGDEVAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD |
| | <i>A. fumigatus</i> 58128 MCSFDTVArT | | LGDEVAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD |
| 15 | <i>A. fumigatus</i> 26906 MCSFDTVArT | | LGDEVAANFT ALFAPdIRAR aKkhLPGVtL TDEDVVSLMD |
| | <i>A. fumigatus</i> 32239 MCSFDTVArT | | LGDEVEANFT ALFAPAIRAR IEkhLPGVQL TDDDVVSLMD |
| 20 | <i>E. nidulans</i> MCSFDTMArT | | rADEIEANFT AIMGPPIRkR LEndLPGIKL TNENViyLMD |
| | <i>T. thermophilus</i> LCPFETLArN | | gGHDAQEKFA kqFAPAIIEK IKDhLPGVDL AvsDVpyLMD |
| | <i>T. lanuginosa</i> LCPFDTVGSd | | .DptqpAEfI qVFGPRVlkK ItkhMPGVNL TLEDVplFMD |
| 25 | <i>M. thermophila</i> LCPFETVAss | | IGDDaQDtYl StFAGPItAR VNAnLPGaNL TDADtVaLMD |
| | Basidio LCPFETVS.. | | dSDpqxnXWl AVFAPPItAR LNAaaPGaNL TDxDaxNLxx |
| 30 | Consensus MCPFDTVA-T | | LGDDVEANFT AVFAPPiRAR LEA-LPGVNL TDEDVVNLMD |
| | Fcp10 MCPFDTVART | | LGDDVEANFT AVFAPPiRAR LEAHLPGVNL TDEDVVNLMD |
| 35 | 300 | 251 | |
| | <i>A. terreus</i> 9a1 dKYYGYGGGN | | dD..Aht... ..LSPF CDLFTa..tE WtQYNYLlSL |

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| | | |
|----|---|--|
| | <i>A. terreus</i> cbs | dD..Aht... ..LSPF CDLFTa..aE WtQYNYLlSL |
| | dKYYGYGGGN | |
| | <i>A. niger</i> var. <i>awamori</i> Tv..DTK... ..LSPF CDLFTH..dE WiHYDYLQSL | |
| | kKYYGHGAGN | |
| 5 | <i>A. niger</i> NRRL3135 | Tv..DTK... ..LSPF CDLFTH..dE WiNYDYLQSL |
| | kKYYGHGAGN | |
| | <i>A. fumigatus</i> 13073 | SD..ASQ... ..LSPF CQLFTH..nE WkKYNYLQSL |
| | gKYYGYGAGN | |
| 10 | <i>A. fumigatus</i> 32722 | SD..ASQ... ..LSPF CQLFTH..nE WkKYNYLQSL |
| | gKYYGYGAGN | |
| | <i>A. fumigatus</i> 58128 | SD..ASQ... ..LSPF CQLFTH..nE WkKYNYLQSL |
| | gKYYGYGAGN | |
| | <i>A. fumigatus</i> 26906 | SD..ASQ... ..LSPF CQLFTH..nE WkKYNYLQSL |
| | gKYYGYGAGN | |
| 15 | <i>A. fumigatus</i> 32239 | AD..ASE... ..LSPF CAIFTH..nE WkKYDYLQSL |
| | gKYYGYGAGN | |
| | <i>E. nidulans</i> | AH..GTE... ..LSPF CAIFTE..kE WlQYDYLQSL |
| | sKYYGYGAGS | |
| 20 | <i>T. thermophilus</i> | ht..DT.... ..LSPF CALsTQ..eE WqaYDYYQSL |
| | gKYYGnGGGN | |
| | <i>T. lanuginosa</i> | PvlfPrQ... ..LSPF CHLFTa..dD WmaYDYYyTL |
| | dKYYSHGGGS | |
| | <i>M. thermophila</i> | SsdpATadag ggnggrpLSPF CrLFSE..sE WrayDYLQSV |
| | gKWYGYGPGN | |
| 25 | Basidio |xexxSxF CDLFexxpeE FxaFxYxgdL |
| | dKFYGTgYgQ | |
| | Consensus | SD--ATQ--- ----LSPF CDLFTH---E W-QYDYLQSL - |
| | KYYGYGAGN | |
| 30 | Fcp10 | SD..ATQ... ..LSPF CDLFTH..DE WiQYDYLQSL |
| | GKYYGYGAGN | |
| | | 301 |
| | 350 | |
| 35 | <i>A. terreus</i> 9a1 | PLGPvQGVGW aNELMARLTR A.PVHDHTCv NNTLDASPAT |
| | FPLNATLYAD | |
| | <i>A. terreus</i> cbs | PLGPvQGVGW aNELIARLTR S.PVHDHTCv NNTLDANPAT |
| | FPLNATLYAD | |

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| | | |
|----|-------------------------------------|---|
| | <i>A. niger</i> var. <i>awamori</i> | PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSNPAT |
| | FPLNSTLYAD | |
| | <i>A. niger</i> NRRL3135 | PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSSPAT |
| | FPLNSTLYAD | |
| 5 | <i>A. fumigatus</i> 13073 | PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT |
| | FPLNATMYvD | |
| | <i>A. fumigatus</i> 32722 | PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT |
| | FPLNATMYvD | |
| 10 | <i>A. fumigatus</i> 58128 | PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT |
| | FPLNATMYvD | |
| | <i>A. fumigatus</i> 26906 | PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT |
| | FPLNATMYvD | |
| | <i>A. fumigatus</i> 32239 | PLGPAQGIGF tNELIARLTN S.PVQDHTST NsTLDSDPAT |
| | FPLNATIYvD | |
| 15 | <i>E. nidulans</i> | PLGPAQGIGF tNELIARLTQ S.PVQDNTST NHTLDSNPAT |
| | FPLDrkLYAD | |
| | <i>T. thermophilus</i> | PLGPAQGVGF vNELIARMTH S.PVQDYTTv NHTLDSNPAT |
| | FPLNATLYAD | |
| 20 | <i>T. lanuginosa</i> | AFGPSRGVGF vNELIARMTg NlPVKDHTTv NHTLDdNPET |
| | FPLDAvLYAD | |
| | <i>M. thermophila</i> | PLGPTQGVGF vNELLARLA. GvPVRDgTST NRTLdGDPRT |
| | FPLGrPLYAD | |
| | Basidio | PLGPvQGVGY iNELLARLTx qa.VRDNTqT NRTLdSSPxT |
| | FPLNrTFYAD | |
| 25 | Consensus | PLGPAQGVGF -NELIARLTH S-PVQDHTST NHTLDSNPAT |
| | FPLNATLYAD | |
| | Fcp10 | PLGPAQGVGF vNELIARLTH S.PVQDHTST NHTLDSNPAT |
| | FPLNATLYAD | |
| 30 | | |
| | | 351 |
| | 400 | |
| | <i>A. terreus</i> 9a1 | FSHDSnLVSI FWALGLYNGT aPLSqTSVE. .SvsQTDGYA |
| | AAWTVPFAR | |
| 35 | <i>A. terreus</i> cbs | FSHDSnLVSI FWALGLYNGT kPLSqTTVE. .ditrTDGYA |
| | AAWTVPFAR | |
| | <i>A. niger</i> var. <i>awamori</i> | FSHDNGIISI LFALGLYNGT kPLSTTTVE. .NitQTDGFS |
| | SAWTVPFASR | |

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| | | |
|----|---|---|
| | <i>A. niger</i> NRRL3135 SAWTVPFASR | FSHDNGIISI LFALGLYNGT kPLSTTTVE. .NitQTDGFS |
| | <i>A. fumigatus</i> 13073 ASWvVPFGAR | FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKElDGYS |
| 5 | <i>A. fumigatus</i> 32722 ASWvVPFGAR | FSHDNSMVSI FFALGLYNGT gPLSrTSVE. .SaKElDGYS |
| | <i>A. fumigatus</i> 58128 ASWvVPFGAR | FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKElDGYS |
| 10 | <i>A. fumigatus</i> 26906 ASWvVPFGAR | FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKElDGYS |
| | <i>A. fumigatus</i> 32239 ASWAVPFGAR | FSHDNGMIPI FFAMGLYNGT ePLSqTSeE. .StKESNGYS |
| | <i>E. nidulans</i> ASWTVPFGAR | FSHDNSMISI FFAMGLYNGT qPLSmdSVE. .SiQEmDGYA |
| 15 | <i>T. thermophilus</i> AAWTVPFGR | FSHDNTMtSI FaALGLYNGT akLSTTeIK. .SiEETDGYS |
| | <i>T. lanuginosa</i> ASWTVPFAAR | FSHDNTMtGI FsAMGLYNGT kPLSTSkIQP pTgAAADGYA |
| 20 | <i>M. thermophila</i> ASWAVPFAAR | FSHDNdMMGV LgALGaYDgV pPLdkTA..R rdpEELGGYA |
| | Basidio TSklVPFSAR | FSHDNqMVAI FsAMGLFNqS aPLdPSxpDP nrt.....Wv |
| 25 | Consensus ASWTVPFAAR | FSHDNTMVSI FFALGLYNGT -PLSTTSVEP -S-EETDGYS |
| | Fcp10 ASWTVPFAAR | FSHDNTMVSI FFALGLYNGT KPLSTTSVE. .SiEETDGYS |
| 30 | 450 | 401 |
| | <i>A. terreus</i> 9a1 PLHGCPtDKL | AYVEMMQC.. ra.....EKEPL VRVLVNDRVm |
| | <i>A. terreus</i> cbs PLHGCAVDNL | AYIEMMQC.. ra.....EKQPL VRVLVNDRVm |
| 35 | <i>A. niger</i> var. <i>awamori</i> PLHGCPIDaL | lyVEMMQC.. Qa.....EQEPL VRVLVNDRVV |
| | <i>A. niger</i> NRRL3135 PLHGCPVDaL | lyVEMMQC.. Qa.....EQEPL VRVLVNDRVV |

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| | | |
|----|---|---|
| | <i>A. fumigatus</i> 13073 PLHGCDVDKL | AYfEtMQC.. Ks..... EKEPL VRaLINDRVV |
| | <i>A. fumigatus</i> 32722 PLHGCDVDKL | AYfEtMQC.. Ks..... EKEPL VRaLINDRVV |
| 5 | <i>A. fumigatus</i> 58128 PLHGCDVDKL | AYfEtMQC.. Ks..... EKESL VRaLINDRVV |
| | <i>A. fumigatus</i> 26906 PLHGCDVDKL | AYfEtMQC.. Ks..... EKEPL VRaLINDRVV |
| 10 | <i>A. fumigatus</i> 32239 PLHGCAVDKL | AYfEtMQC.. Ks..... EKEPL VRaLINDRVV |
| | <i>E. nidulans</i> PLHGCAVDKF | AYfELMQC.. E..... KKEPL VRVLVNDRVV |
| | <i>T. thermophilus</i> PLHGCEVDsL | AYIEMMQC.. Dd..... sDEPV VRVLVNDRVV |
| 15 | <i>T. lanuginosa</i> PLHGCrVDRW | AYVELLRC.. Etetsseeee EG... EDEPF VRVLVNDRVV |
| | <i>M. thermophila</i> TLkGCGaDER | iYVEkMRC.. sggggggggg EGrqeKDEeM VRVLVNDRVM |
| 20 | Basidio PLEfCGgDxd | mvVErLxCxx xgtxxxxxxxx xxxxxxxxxxx VRVLVNDaVq |
| | Consensus PLHGCGVDKL | AYVEMMQC-- E----- EG---EKEPL VRVLVNDRVV |
| 25 | Fcp10 PLHGCGVDKL | AYVEMMQC.. EA..... EKEPL VRVLVNDRVV |

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| | 451 | 482 |
|-----------------------|-----------------------|---------------|
| A. terreus 9a1 | GRCKrDAFVA GLSFAQAG.. | GNWADCF--- -- |
| A. terreus cbs | GRCKrDDFVE GLSFARAG.. | GNWAECF--- -- |
| A. niger var. awamori | GRCtrDsFVr GLSFARSG.. | GDWAECsA-- -- |
| 5 A. niger NRRL3135 | GRCtrDsFVr GLSFARSG.. | GDWAECFA-- -- |
| A. fumigatus 13073 | GRCKlNDFVK GLSWARSG.. | GNWGECSF-- -- |
| A. fumigatus 32722 | GRCKlNDFVK GLSWARSG.. | GNWGECSF-- -- |
| A. fumigatus 58128 | GRCKlNDFVK GLSWARSG.. | GNWGECSF-- -- |
| A. fumigatus 26906 | GRCKlNDFVK GLSWARSG.. | GNWGECSF-- -- |
| 10 A. fumigatus 32239 | GRCKlKDFVK GLSWARSG.. | GNSEQSFS-- -- |
| E. nidulans | GRCtlDDWVE GLNFARSG.. | GNWktCFTl- -- |
| T. thermophilus | GRCKrDDFVr GLSFARqG.. | GNWEGCYAas e- |
| T. lanuginosa | GRCRrDEWIK GLTFARqG.. | GHWDrCF--- -- |
| M. thermophila | GmCtlErFIE SMAFARGN.. | GKWDlCFA-- -- |
| 15 Basidio | GxCtlDAFVE SqxYAReDgq | GDFEKCFAtp xx |
| Consensus | GRCK-DDFVE GLSFARSG-- | GNWEECFA-- -- |
| Fcp10 | GRCKRDDFVE GLSFARSG.. | GNWEECFA.. .. |

20

25

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Figure 5

CP-1

5 Eco RI M G V F V V L L S I A T L F G S T 17
TATATGAATTCATGGGCGTGTTCGTGCTACTGTCCATTGCCACCTTGTTTCGGTTCCA

1 -----+-----+-----+-----+-----+-----+ 60

ATATACTTAAGTACCCGCACAAGCAGCACGATGACAGGTAACGGTGAACAAGCCAAGGT

10 S G T A L G P R G N S H S C D T V D G G 37
CATCCGGTACCGCCTTGGGTCCTCGTGGTAATTCTCACTCTTGACACTGTTGACGGTG

61 -----+-----+-----+-----+-----+-----+ 120

GTAGGCCATGGCGGAACCCAGGAGCACCATTAAAGAGTGAGAACACTGTGACAACTGCCAC

CP-2

CP-3.10

15 Y Q C F P E I S H L W G Q Y S P E F S L 57
GTTACCAATGTTTCCAGAAATTTCTCACTTGTGGGGTCAATACTCTCCATTCTTCTCTT

121 -----+-----+-----+-----+-----+-----+ 180

CAATGGTTACAAAGGGTCTTTAAAGAGTGAACACCCCAGTTATGAGAGGTAAGAAGAGAA

20 A D E S A I S P D V P K Q C R V T F V Q 77
TGGCTGACGAATCTGCTATTTCTCCAGACGTTCCAAAGGGTTGTAGAGTTACTTTTCGTTT

181 -----+-----+-----+-----+-----+-----+ 240

ACCGACTGCTTAGACGATAAAGAGGTCTGCAAGGTTTCCCGACATCTCAATGAAAGCAAG

CP-4.10

CP-5.10

25 V L S R H G A R Y P T S S K S K K Y S A 97
AAGTTTGTCTAGACACGGTGCTAGATACCCAACTTCTTCTAAGTCTAAGAAGTACTCTG

241 -----+-----+-----+-----+-----+-----+ 300

TTCAAAACAGATCTGTGCCACGATCTATGGGTTGAAGAAGATTCAAGTTCTTCATGAGAC

30 L I E A I Q K N A T A F K G K Y A F L K 117
CTTTGATTGAAGCTATTCAAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTTCTTGA

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301 -----+-----+-----+-----+-----+-----+ 360

GAAACTAACTTCGATAAGTTTCTTGCATGACGAAAGTTCCCATTCATGCGAAAGAAGT

CP-6

CP-7.10

5 T Y N Y T L G A D D L T P F G E Q Q M V 137

AGACTTACAACACTACTTTGGGTGCTGACGACTTGACTCCATTCCGGTGAACAACAAATGG

361 -----+-----+-----+-----+-----+-----+ 420

TCTGAATGTTGATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTGTGTTTACC

10 N S G I K F Y R R Y K A L A R K I V P F 157

TTAACTCTGGTATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCAT

421 -----+-----+-----+-----+-----+-----+ 480

AATTGAGACCATAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTA

CP-8.10

15

CP-9.10

Y R A S G S D R V I A S A E K F I E G F 177

TCGTTAGAGCTTCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTTATTGAAGGTT

481 -----+-----+-----+-----+-----+-----+ 540

AGCAATCTCGAAGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAAGTTCCAA

20

Q S A K L A D P G A N P H Q A S P V I N 197

TCCAATCTGCTAAGTTGGCTGACCCAGGTGCTAACCCACACCAAGCTTCTCCAGTTATTA

541 -----+-----+-----+-----+-----+-----+ 600

AGGTTAGACGATTCAACCGACTGGGTCCACGATTGGGTGTGGTTCGAAGAGGTCAATAAT

25

CP-10.10

CP-11.10

V I I P E G A G Y N N T L D H G L C T A 217

ACGTTATTATTCCAGAAGGTGCTGGTTACAACAACACTTTGGACCACGGTTTGTGTACTG

601 -----+-----+-----+-----+-----+-----+ 660

30

TGCAATAATAAGGTCTTCCACGACCAATGTTGTTGTGAAACCTGGTGCCAAACACATGAC

F E E S E L G D D V E A N F T A Y F A P 237

CTTTTCGAAGAATCTGAATTGGGTGACGACGTTGAAGCTAACTTCACTGCTGTTTTCGCTC

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661 -----+-----+-----+-----+-----+-----+ 720
GAAAGCTTCTTAGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGACAAAAGCGAG

CP-12.10

5 P I R A R L E A H L P G V N L T D E D V 257
CACCTATTAGAGCTAGATTGGAAGCTCACTTGCCAGGTGTTAACTTGACTGACGAAGACG

721 -----+-----+-----+-----+-----+-----+ 780
GTGGATAATCTCGATCTAACCTTCGAGTGAACGGTCCACAATTGAACTGACTGCTTCTGC

10 CP-13.10

V N L M D M C P F D T V A R T S D A T Q 277
TTGTAACTTGATGGACATGTGTCCATTGACACTGTTGCTAGAACTTCTGACGCTACTC

781 -----+-----+-----+-----+-----+-----+ 840
AACAATTGAACTACCTGTACACAGGTAAGCTGTGACAACGATCTTGAAGACTGCGATGAG

15 L S P F C D L F T H D E W I Q Y D Y L Q 297
AATTGTCTCCATTCTGTGACTTGTTCACTCACGACGAATGGATTCAATACGACTACTTGC

841 -----+-----+-----+-----+-----+-----+ 900
TTAACAGAGGTAAGACACTGAACAAGTGAGTGCTGCTTACCTAAGTTATGCTGATGAACG

20 CP-14.10

CP-15.10

S L G K Y Y G Y G A G N P L G P A Q G V 317
AATCTTTGGGTAAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTG

901 -----+-----+-----+-----+-----+-----+ 960

25 TTAGAAACCCATTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCAC

G F Y N E L I A R L T H S P V Q D H T S 337
TTGGTTTCGTTAACGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTT

30 1020 961 -----+-----+-----+-----+-----+-----+
AACCAAAGCAATTGCTTAACTAACGATCTAACTGAGTGAGAGGTCAAGTTCTGGTGTGAA

CP-16.10

CP-17.10

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T N H T L D S N P A T F P L N A T L Y A 357
CTACTAACCACACTTTGGACTCTAACCCAGCTACTTTCCCATTGAACGCTACTTTGTACG
1021 -----+-----+-----+-----+-----+
1080
5 GATGATTGGTGTGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGCGATGAAACATGC
D F S H D N T M Y S I F F A L G L Y N G 377
CTGACTTCTCTCAGACAACACTATGGTTTCTATTTTCTTCGCTTTGGGTTTGTACAACG
1081 -----+-----+-----+-----+-----+
10 1140
GACTGAAGAGAGTGCTGTTGTGATACCAAAGATAAAAGAAGCGAAACCCAAACATGTTGC
CP-18.10
CP-19.10
T K P L S T T S V E S I E E T D G Y A A 397
15 GTACTAAGCCATTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACGCTG
1141 -----+-----+-----+-----+-----+
1200
CATGATTCGGTAACAGATGATGAAGACAACCTTAGATAACTTCTTTGACTGCCAATGCGAC
20 S W T V P F A A R A Y V E M M Q C E A E 417
CTTCTTGACTGTTCCATTGCTGCTAGAGCTTACGTTGAAATGATGCAATGTGAAGCTG
1201 -----+-----+-----+-----+-----+
1260
GAAGAACCTGACAAGGTAAGCCACGATCTCGAATGCAACTTTACTACGTTACACTTCGAC
25 CP-20.10
CP-21.10
K E P L V R V L V N D R V V P L H G C G 437
AAAAGGAACCATTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG
1261 -----+-----+-----+-----+-----+
30 1320
TTTTCCTTGGTAAACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACAC
V D K L G R C K R D D F V E G L S F A R 457
GTGTTGACAAGTTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTGTCTTTTCGCTA

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1321 -----+-----+-----+-----+-----+-----+
 1380
 CACAACCTGTTCAACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGAT
 5 S G G N W E E C F A * Eco RI CP-22.10 467
 GATCTGGTGGTAACTGGGAAGAATGTTTCGCTTAAGAATTCATATA
 1381 -----+-----+-----+-----+-----+----- 1426
 CTAGACCACCATTGACCCTTCTTACAAAGCGAATTCTTAAGTATAT

10

Figure 6

50 1
 15 *P. involutus* (phyA1) ----- ~FPipeseqR nWSPYSPYFP LAEyKA....
 pPaGCQInqV
P. involutus (phyA2) ----- ~FsipeseqR nWSPYSPYFP LAEyKA....
 pPaGCeInqV
 20 *T. pubescens* ----- ~LDvtRDVqQ sWSmYSPYFP aAtyvA....
 pPaSCQInqV
A. pediades ----- ~pffpPQIQD sWAaYTPYYP VqAyTP....
 pPKDCKITqV
P. lycii ----- ~LPipAQnTs nWGPydPFFP VEpyAA....
 pPEGctVTqV
 25 *A. terreus* 9a1 KhSDCNSVDh GYQCfPELSH kWGLYAPYFS LqDESFPFpD
 VPEDCHITFV
A. terreus cbs NhSDCtSVDr GYQCfPELSH kWGLYAPYFS LqDESFPFpD
 VPDDCHITFV
 30 *A. niger* var. *awamori* NqSTCDTVDq GYQCfSEtSH LWGQYAPFFS LANESAISPD
 VPAGCRVTFa
A. niger T213 NqSSCDTVDq GYQCfSEtSH LWGQYAPFFS LANESvISPD
 VPAGCRVTFa
A. niger NRRL3135 NqSSCDTVDq GYQCfSEtSH LWGQYAPFFS LANESvISPE
 VPAGCRVTFa
 35 *A. fumigatus* ATCC13073 GSKSCDTVDl GYQCSPAtSH LWGQYSPFFS LEDElSVSSK
 LPKDCRITLV

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| | | |
|----|---|---|
| | <i>A. fumigatus</i> ATCC32722 | GSkSCDTVD1 GYQCSPAtSH LWGQYSPFFS LEDElSVSSK LPKDCRITLV |
| | <i>A. fumigatus</i> ATCC58128 | GSkSCDTVD1 GYQCSPAtSH LWGQYSPFFS LEDElSVSSK LPKDCRITLV |
| 5 | <i>A. fumigatus</i> ATCC26906 | GSkSCDTVD1 GYQCSPAtSH LWGQYSPFFS LEDElSVSSK LPKDCRITLV |
| | <i>A. fumigatus</i> ATCC32239 | GSkACDTVE1 GYQCSPGtSH LWGQYSPFFS LEDElSVSSD LPKDCRVTFV |
| 10 | <i>E. nidulans</i> VPhGCeVTFV | QNHSCNTaDg GYQCfPNVSH VWGQYSPYFS IEQESAISeD |
| | <i>T. thermophilus</i> VPQNCKITFV | DSHSCNTVEg GYQCrPEISH sWGQYSPFFS LADQSEISPD |
| | <i>T. lanuginosa</i> VPKGCRVeFV | ----- ----nvDIAR hWGQYSPFFS LAEvSEISPA |
| 15 | <i>M. thermophila</i> IPDDCeVTFa | ESRPCDTpDl GFQCgTAISH FWGQYSPYFS VPSElDaS.. |
| | Consensus Seq. 11 VPKGCRVTFV | NSHSCDTVD- GYQC-PEISH LWGQYSPFFS LADESAISPD |
| 20 | | |
| | | 51 |
| | 100 | |
| | <i>P. involutus</i> (phyA1) KSFKYdLGns | NIIqRHGARF PTSGaTtRik AgLtKLQgvq nftDAKFnFI |
| 25 | <i>P. involutus</i> (phyA2) KSftYdLGts | NIIqRHGARF PTSGaAtRik AgLsKLQsvq nftDPKFDFI |
| | <i>T. pubescens</i> tnYtYSLGqD | HIIqRHGARF PTSGaAKRiq TaVAKLKaaS nytDPlLAFV |
| 30 | <i>A. pediades</i> tnYtYTLGhD | NIIqRHGARF PTSGaGtRiq AaVKKLQsak TytDPRLDfL |
| | <i>P. lycii</i> NdFvYkFGvA | NLIqRHGARW PTSGarsRqv AaVAKIQmar PftDPKYEFL |
| | <i>A. terreus</i> 9a1 QSYNYSLDSE | QVLARHGARS PThSKTKaYA AtIAaIQKSA TaFpGKYAFL |
| 35 | <i>A. terreus</i> cbs KSYNYSMGSE | QVLARHGARS PTdSKTKaYA AtIAaIQKNA TaLpGKYAFL |
| | <i>A. niger</i> var. <i>awamori</i> KTYNYSLGAD | QVLSRHGARY PTeSKGKKYS ALIEeIQQNv TtFDGKYAFL |

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| | | |
|----|---|--|
| | <i>A. niger</i> T213 KTYNYSLGAD | QVLSRHGARY PTeSKGKKYS ALIEeIQQNv TtFDGKYAFL |
| | <i>A. niger</i> NRRL3135 KTYNYSLGAD | QVLSRHGARY PTdSKGKKYS ALIEeIQQNA TtFDGKYAFL |
| 5 | <i>A. fumigatus</i> ATCC13073 KTYNYTLGAD | QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL |
| | <i>A. fumigatus</i> ATCC32722 KTYNYTLGAD | QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL |
| 10 | <i>A. fumigatus</i> ATCC58128 KTYNYTLGAD | QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL |
| | <i>A. fumigatus</i> ATCC26906 KTYNYTLGAD | QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL |
| | <i>A. fumigatus</i> ATCC32239 ETYNYTLGAD | QVLSRHGARY PTASKSKKYk kLVtaIQKNA TeFKGKFAFL |
| 15 | <i>E. nidulans</i> ESYNYTLGAD | QVLSRHGARY PTeSKSKaYS GLIEaIQKNA TsFwQQYAFL |
| | <i>T. thermophilus</i> KdYrYqLGAN | QLLSRHGARY PTSSKTELYS qLIIsRIQKtA TaYKGyYAFL |
| 20 | <i>T. lanuginosa</i> RdYaYhLGAD | QVLSRHGARY PTAhKSEvYA ELLQRIQDtA TeFKGDFAFL |
| | <i>M. thermophila</i> RTYDYTLGAD | QVLSRHGARA PTlkRAasYv DLIDRIHhGA isYgPgYEFL |
| 25 | Consensus Seq. 11 KTYNYTLGAD | QVLSRHGARY PTSSKSKKYS ALIERIQKNA T-FKGKYAFL |

101

150

| | | |
|----|---|---|
| 30 | <i>P. involutus</i> (phyA1) VVDSAtNWtA | DLvPFGAaQs fDAGqEaFaR YskLvSKNnL PFIRAdGSDR |
| | <i>P. involutus</i> (phyA2) VVDtAtNWtA | DLvPFGAaQs fDAGLEvFaR YskLvSsDnL PFIRSDGSDR |
| | <i>T. pubescens</i> VVATANNWtA | sLveLGAtQs sEAGqEaFtR YsSLvSaDeL PFVRASGSDR |
| 35 | <i>A. pediades</i> VVDSAtNWtE | DLvPFGAlQs sQAGeEtFQR YsfLvSKEnL PFVRASSNR |
| | <i>P. lycii</i> VVDSStNWtA | DLlPFGANQs hQTGtDMYtR YsTLfEgGdV PFVRAAGdQR |

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| | | |
|----|---|--|
| | <i>A. terreus</i> 9a1 VhESA EK FVE | ELTPFG rNQL rDlGaQFYeR YNAL.TRHIn PFVRATDAsR |
| | <i>A. terreus</i> cbs VhESA EK FVE | NLTPFG rNQL qDlGaQFYRR YDTL.TRHIn PFVRAADSsR |
| 5 | <i>A. niger</i> var. <i>awamori</i> VIASGEKFIE | DLTPFG EQEL VNSGIKFYQR YESL.TRNII PFIRSSGSsR |
| | <i>A. niger</i> T213 VIASGEKFIE | DLTPFG EQEL VNSGIKFYQR YESL.TRNII PFIRSSGSsR |
| 10 | <i>A. niger</i> NRRL3135 VIASGKKFIE | DLTPFG EQEL VNSGIKFYQR YESL.TRNIV PFIRSSGSsR |
| | <i>A. fumigatus</i> ATCC13073 VIASGEKFIE | DLTPFG EQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR |
| | <i>A. fumigatus</i> ATCC32722 VIASGEKFIE | DLTPFG EQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR |
| 15 | <i>A. fumigatus</i> ATCC58128 VIASGEKFIE | DLTPFG EQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR |
| | <i>A. fumigatus</i> ATCC26906 VIASGEKFIE | DLTAFGEQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR |
| 20 | <i>A. fumigatus</i> ATCC32239 VIASGEKFIE | DLTPFG EQQM VNSGIKFYQK YKAL.AgSVV PFIRSSGSsR |
| | <i>E. nidulans</i> VVASAEKFIN | DLTiFGENQM VD SGaKFYRR YKnL.ARKNt PFIRASGSDR |
| | <i>T. thermophilus</i> VIASGrIFIE | DLTPFGENQM IQlGIKFYnH YKSL.ARN aV PFVRCSGSDR |
| 25 | <i>T. lanuginosa</i> VIASAEfFnR | NLTRFGEEQM MESGrQFYHR YREq.AREIV PFVRAAGSAR |
| | <i>M. thermophila</i> VhSAENFtQ | ELTRtGQQQM VNSGIKFYRR YRAL.ARKsI PFVRTAGqDR |
| 30 | Consensus Seq. 11 VIASAEKFIE | DLTPFGENQM VNSGIKFYRR YKAL-ARNIV PFVRASGSDR |
| | | 151 |
| | 200 | |
| 35 | <i>P. involutus</i> (phyA1) PAaGD..... | GFaSA..... ..shNtvqPk LNLILPQ..T gNDTLEDNMC |
| | <i>P. involutus</i> (phyA2) PAaGE..... | GFaSA..... ..srNaiqPk LDLILPQ..T gNDTLEDNMC |

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| | | |
|----|---|---|
| | <i>T. pubescens</i> PAaGD..... | GFaIA..... ..ssNsITPV LSVIISE..A gNDTLDDNMC |
| | <i>A. pediades</i> PnaGs..... | GFsAA..... ..shHvINPI LfVILSE..S LNDTLDDAMC |
| 5 | <i>P. lycii</i> PnevD..... | GFgdA..... ..sgEtvIPt LQVVLQE..E gNcTLcNNMC |
| | <i>A. terreus</i> 9a1 TAFES...ST | GFQTARqDDh hAnpHQPSPr VDVaIPEGSA YNNTLEHSLC |
| 10 | <i>A. terreus</i> cbs TAFEA...ST | GFQNARqGDP hAnpHQPSPr VDVVIPEGTA YNNTLEHSIC |
| | <i>A. niger</i> var. <i>awamori</i> TvFED...Se | GFQSTKLkDP rAqpgQSSPk IDVWISEASS sNNTLDpGtC |
| | <i>A. niger</i> T213 TvFED...Se | GFQSTKLkDP rAqpgQSSPk IDVWISEASS sNNTLDpGtC |
| 15 | <i>A. niger</i> NRRL3135 TvFED...Se | GFQSTKLkDP rAqpgQSSPk IDVWISEASS sNNTLDpGtC |
| | <i>A. fumigatus</i> ATCC13073 TkFEA...Sq | GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC |
| 20 | <i>A. fumigatus</i> ATCC32722 TkFEA...Sq | GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC |
| | <i>A. fumigatus</i> ATCC58128 TkFEA...Sq | GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC |
| | <i>A. fumigatus</i> ATCC26906 TkFEA...Sq | GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC |
| 25 | <i>A. fumigatus</i> ATCC32239 TnFEA...Se | GFQqANVADP gAt.NRAAPV ISVIIPESeT YNNTLDHSVC |
| | <i>E. nidulans</i> vSFEN...de | GFRkaQLhDh g.s.gQATPV VNVIIPeIdG FNNTLDHStC |
| 30 | <i>T. thermophilus</i> PvFED...SS | GFQSAKVlDP hSdKHDPpT INVIIeEGPS YNNTLDtGsC |
| | <i>T. lanuginosa</i> PAaEE...AP | GFQdAKdrDP rSnkDQAePV INVIISEETG sNNTLDgltC |
| | <i>M. thermophila</i> TAFEEgpyST | GFHSALLADR gStvRPTlPy dmVVIPETAG aNNTLHNDLC |
| 35 | | |
| | Consensus Seq. 11 TAFED---ST | GFQSAKLADP -A--HQASPV INVIIPEGSG YNNTLDHGLC |

| | | |
|----|--|---|
| | | 201 |
| | 250 | |
| | <i>P. involutus</i> (phyA1) LCAFlTVSK. | .SDpqvnaWl AVafPSItAR LNAaaPSVNL TDtDafNLVs |
| 5 | <i>P. involutus</i> (phyA2) LCPFmTVSK. | .SDpqvDaWl AsafPSVtAQ LNAaaPGaNL TDADafNLVs |
| | <i>T. pubescens</i> LCPFETVAt. | .SDpqvnQWl AqFAPPMtAR LNAgaPGaNL TDtDtyNLLt |
| 10 | <i>A. pediades</i> LCAFETIvK. | .SDpqtGiWT SIYGTPIanR LNqqaPGaNI TAADVsnLIp |
| | <i>P. lycii</i> MCPFDTLSS. | .GDESt.tWl GVFApNItAR LNAaaPSaNL SDsDaLtLMD |
| | <i>A. terreus</i> 9a1 MCPFETVS1T | VGDDAvANFT AVFAPAIaqR LEAdLPGVQL StDDVVNLMA |
| 15 | <i>A. terreus</i> cbs MCPFETVS1T | VGDAADNFT AVFAPAIakR LEAdLPGVQL SADDVVNLMA |
| | <i>A. niger</i> var. <i>awamori</i> MCSFDTISs | LADtveANFT AtFAPSIRqR LEndLSGvtL TDtEVtyLMD |
| 20 | <i>A. niger</i> T213 MCSFDTISs | LADtveANFT AtFAPSIRqR LEndLSGvtL TDtEVtyLMD |
| | <i>A. niger</i> NRRL3135 MCSFDTISs | LADtveANFT AtFvPSIRqR LEndLSGvtL TDtEVtyLMD |
| | <i>A. fumigatus</i> ATCC13073 MCSFDTVART | LGDEvAANFT ALFAPdIRAR aEkhlPGvtL TDEDVVSLMD |
| 25 | <i>A. fumigatus</i> ATCC32722 MCSFDTVART | LGDEvAANFT ALFAPdIRAR aEkhlPGvtL TDEDVVSLMD |
| | <i>A. fumigatus</i> ATCC58128 MCSFDTVART | LGDEvAANFT ALFAPdIRAR aEkhlPGvtL TDEDVVSLMD |
| 30 | <i>A. fumigatus</i> ATCC26906 MCSFDTVART | LGDEvAANFT ALFAPdIRAR aKkhLPGvtL TDEDVVSLMD |
| | <i>A. fumigatus</i> ATCC32239 MCSFDTVART | LGDEvAANFT ALFAPAIRAR IEkhLPGVQL TDDDVVSLMD |
| | <i>E. nidulans</i> MCSFDTMART | rADEiEANFT AIMGPPIRkR LEndLPGIKL TNENViyLMD |
| 35 | <i>T. thermophilus</i> LCPFETLArN | gGHDAQEKFA kqFAPAILEK IKDhLPGVDL AvsDVpyLMD |
| | <i>T. lanuginosa</i> LCPFDTVGSd | .DptqpAEFl qVFGPRVlkK ItkhMPGVNL TLEDVplFMD |

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| | | |
|----|---|--|
| | <i>M. thermophila</i> LCPFETVAsS | IGDDAQDtYl StFAGPiTAR VNAnLPGaNL TDADtVaLMD |
| 5 | Consensus Seq. 11 MCPFDTVART | LGDDAEANFT AVFAPPiRAR LEA-LPGVNL TDDEDVvNLMD |
| | | 251 |
| | 300 | |
| 10 | <i>P. involutus</i> (phyA1) dKFYGTGyGQ | ekkSdF CtLFegiPGs FeaFAYggdL |
| | <i>P. involutus</i> (phyA2) dKFYGTGyGQ | eqkSdF CtLFegiPGs FeaFAYagdL |
| | <i>T. pubescens</i> dKFYGTGyGQ | errSeF CDiYeelqAE .daFAYnadL |
| 15 | <i>A. pediades</i> dKFYGTGyGQ | etpSPF CNLF..TPEE FaQFEYfgdL |
| | <i>P. lycii</i> dKYYGTGPGN | gnaSPF CDLF..TAAE YvsYEYYydL |
| 20 | <i>A. terreus</i> 9a1 dKYYGYGGGN | dD..Aht... ..LSPF CDLF..TatE WtQYNYLlSL |
| | <i>A. terreus</i> cbs dKYYGYGGGN | dD..Aht... ..LSPF CDLF..TAAE WtQYNYLlSL |
| | <i>A. niger</i> var. <i>awamori</i> kKYYGHGAGN | Tv..DTK... ..LSPF CDLF..ThDE WiHYDYLQSL |
| 25 | <i>A. niger</i> T213 kKYYGHGAGN | Tv..DTK... ..LSPF CDLF..ThDE WiHYDYLRLS |
| | <i>A. niger</i> NRRL3135 kKYYGHGAGN | Tv..DTK... ..LSPF CDLF..ThDE WiNYDYLQSL |
| 30 | <i>A. fumigatus</i> ATCC13073 gKYYGYGAGN | SD..ASQ... ..LSPF CQLF..ThNE WkKYNYLQSL |
| | <i>A. fumigatus</i> ATCC32722 gKYYGYGAGN | SD..ASQ... ..LSPF CQLF..ThNE WkKYNYLQSL |
| | <i>A. fumigatus</i> ATCC58128 gKYYGYGAGN | SD..ASQ... ..LSPF CQLF..ThNE WkKYNYLQSL |
| 35 | <i>A. fumigatus</i> ATCC26906 gKYYGYGAGN | SD..ASQ... ..LSPF CQLF..ThNE WkKYNYLQSL |
| | <i>A. fumigatus</i> ATCC32239 gKYYGYGAGN | AD..ASE... ..LSPF CAIF..ThNE WkKYDYLQSL |

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| | | |
|----|---|--|
| | <i>E. nidulans</i> sKYYGYGAGS | AH..GTE... ..LSPF CAIF..TEKE WlQYDYLQSL |
| | <i>T. thermophilus</i> gKYYGnGGGN | ht..DT.... ..LSPF CALs..TqEE WqayDYyQSL |
| 5 | <i>T. lanuginosa</i> dKYYSHGGGS | PvlfPrQ... ..LSPF CHLF..TADD WmaYDYyTL |
| | <i>M. thermophila</i> gKWYGYGPGN | SsdpATadag ggngprLSPF CrLF..SEsE WrayDYLQSV |
| 10 | Consensus Seq. 11 KYYGYGAGN | SD--ATQ--- -----LSPF CDLF--TADK W-QYDYLQSL - |
| | | 301 |
| | 350 | |
| 15 | <i>P. involutus</i> (phyA1) FPLNkTFYAD | eLGPvQGvGY vNELIARLTN S.AVRDNTqT NRTLdASPvT |
| | <i>P. involutus</i> (phyA2) FPLNkTMYAD | ALGPvQGvGY iNELLARLTN S.AVNDNTqT NRTLdAaPDT |
| 20 | <i>T. pubescens</i> FPLNrTLyAD | PLGPvQGvGY iNELIARLTa q.nVsDHTqT NsTLdSSPET |
| | <i>A. pediades</i> FPLDrSIYAD | PLGPvQGvGY iNELLARLTe m.PVRDNTqT NRTLdSSPlT |
| | <i>P. lycii</i> FPLNrTFYAD | ALGPvQGvGY vNELLARLTg q.AVRDETqT NRTLdSDPAT |
| 25 | <i>A. terreus</i> 9a1 FPLNATLYAD | PLGPvQGvGW aNELMARLTR A.PVHDHTCv NNTLdASPAT |
| | <i>A. terreus</i> cbs FPLNATLYAD | PLGPvQGvGW aNELIARLTR S.PVHDHTCv NNTLdANPAT |
| 30 | <i>A. niger</i> var. <i>awamori</i> FPLNSTLYAD | PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLdSNPAT |
| | <i>A. niger</i> T213 FPLNSTLYAD | PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLdSNPAT |
| | <i>A. niger</i> NRRL3135 FPLNSTLYAD | PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLdSSPAT |
| 35 | <i>A. fumigatus</i> ATCC13073 FPLNATMYvD | PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT |
| | <i>A. fumigatus</i> ATCC32722 FPLNATMYvD | PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT |

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| | | |
|----|---|---|
| | <i>A. fumigatus</i> ATCC58128 | PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT FPLNATMYvD |
| | <i>A. fumigatus</i> ATCC26906 | PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT FPLNATMYvD |
| 5 | <i>A. fumigatus</i> ATCC32239 | PLGPAQGIGF tNELIARLTN S.PVQDHTST NsTLDSNPAT FPLNATIYvD |
| | <i>E. nidulans</i> FPLDrkLYAD | PLGPAQGIGF tNELIARLTQ S.PVQDNTST NHTLDSNPAT |
| 10 | <i>T. thermophilus</i> FPLNATLYAD | PLGPAQGVGF vNELIARMTH S.PVQDYTTv NHTLDSNPAT |
| | <i>T. lanuginosa</i> FPLDAvLYAD | AFGPSRGVGF vNELIARMTg NlPVKDHTTv NHTLDdNPET |
| | <i>M. thermophila</i> FPLGrPLYAD | PLGPTQGVGF vNELLARLA. GvPVRDgTST NRTLGDGPrt |
| 15 | Consensus Seq. 11 FPLNATLYAD | PLGPAQGVGF -NELIARLTH S-PVQDHTST NHTLDSNPAT |
| | | 351 |
| 20 | 400 | |
| | <i>P. involutus</i> (phyA1) TSSlVPFSGR | FSHDNlMVAV FsAMGLFrqP aPLSTsvpNP wrt.....Wr |
| | <i>P. involutus</i> (phyA2) TSSvVPFSAR | FSHDNlMVAV FsAMGLFrqS aPLSTSTpDP nrt.....Wl |
| 25 | <i>T. pubescens</i> vkkivPFASR | FSHDNqMVAI FsAMGLFNqS aPLdPTTpDP art.....Fl |
| | <i>A. pediades</i> TSRltPFASR | LSHDNqMIAI FsAMGLFNqS sPLdPSfpNP krt.....Wv |
| 30 | <i>P. lycii</i> DSklVPFSGH | FSHDNTMVPI FaALGLFNAT a.LdPlkpDe nrl.....Wv |
| | <i>A. terreus</i> 9a1 AAWTVPFAAR | FSHDSnLVSI FWALGLYNGT aPLSqTSVES Vs..QTDGYA |
| | <i>A. terreus</i> cbs AAWTVPFAAR | FSHDSnLVSI FWALGLYNGT KPLSqTTVED It..rTDGYA |
| 35 | <i>A. niger</i> var. <i>awamori</i> SAWTVPFASR | FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS |
| | <i>A. niger</i> T213 SAWTVPFASR | FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS |

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| | | |
|----|---|---|
| | <i>A. niger</i> NRRL3135 SAWTVPFASR | FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS |
| | <i>A. fumigatus</i> ATCC13073 ASWvVPFGAR | FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..EldGYS |
| 5 | <i>A. fumigatus</i> ATCC32722 ASWvVPFGAR | FSHDNSMVSI FFALGLYNGT gPLSrTSVES ak..EldGYS |
| | <i>A. fumigatus</i> ATCC58128 ASWvVPFGAR | FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..EldGYS |
| 10 | <i>A. fumigatus</i> ATCC26906 ASWvVPFGAR | FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..EldGYS |
| | <i>A. fumigatus</i> ATCC32239 ASWAVPFGAR | FSHDNGMIPI FFAMGLYNGT EPLSqTSeES tk..ESNGYS |
| | <i>E. nidulans</i> ASWTVPFGAR | FSHDNSMISI FFAMGLYNGT QPLSmdSVES Iq..EmDGYA |
| 15 | <i>T. thermophilus</i> AAWTVPFGR | FSHDNTMtSI FaALGLYNGT akLSTTeIKS Ie..ETDGYS |
| | <i>T. lanuginosa</i> ASWTVPFAAR | FSHDNTMtGI FsAMGLYNGT KPLSTSkIQP ptgaAADGYA |
| 20 | <i>M. thermophila</i> ASWAVPFAAR | FSHDNdMMGV LgALGaYDgv pPLdkTArrd ..peElGGYA |
| | Consensus Seq. 11 ASWTVPFAAR | FSHDNTMVSI FFALGLYNGT KPLSTTSVES I---ETDGYA |
| 25 | 450 | 401 |
| | <i>P. involutus</i> (phyA1) PLEfCGgDRn | mvVErLsC.. fGt..... Tk VRVLVQDQVq |
| 30 | <i>P. involutus</i> (phyA2) PLEfCGgDQd | maVErLsC.. AGt..... Tk VRVLVQDQVq |
| | <i>T. pubescens</i> PLafCGaDts | mvVErLDC.. GGa..... Qs VRLLVNDaVq |
| | <i>A. pediades</i> PLkfCGgDmd | mvTErLLCQr DGtGsGGpsr imrNgnvQTF VRILVNDaLq |
| 35 | <i>P. lycii</i> PLEfCGg.vd | mtVEkLaC..sgKea VRVLVNDaVq |
| | <i>A. terreus</i> 9a1 PLHGCPtDKL | AYVEMMQCrAEK...EPL VRVLVNDRVM |

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| | | |
|----|---|--|
| | <i>A. terreus</i> cbs PLHGCAVDNL | AYIEMMQCrAEK...QPL VRVLVNDVRVM |
| | <i>A. niger</i> var. <i>awamori</i> PLHGCPIDaL | 1YVEMMQCQAEQ...EPL VRVLVNDRVV |
| 5 | <i>A. niger</i> T213 PLHGCPIDaL | 1YVEMMQCQAEQ...EPL VRVLVNDRVV |
| | <i>A. niger</i> NRRL3135 PLHGCPVDaL | 1YVEMMQCQAEQ...EPL VRVLVNDRVV |
| 10 | <i>A. fumigatus</i> ATCC13073 PLHGCDVDKL | AYfEtMQCKSEK...EPL VRaLINDRVV |
| | <i>A. fumigatus</i> ATCC32722 PLHGCDVDKL | AYfEtMQCKSEK...EPL VRaLINDRVV |
| | <i>A. fumigatus</i> ATCC58128 PLHGCDVDKL | AYfEtMQCKSEK...ESL VRaLINDRVV |
| 15 | <i>A. fumigatus</i> ATCC26906 PLHGCDVDKL | AYfEtMQCKSEK...EPL VRaLINDRVV |
| | <i>A. fumigatus</i> ATCC32239 PLHGCAVDKL | AYfEtMQCKSEK...EPL VRaLINDRVV |
| 20 | <i>E. nidulans</i> PLHGCAVDKF | AYfELMQCE.KK...EPL VRVLVNDRVV |
| | <i>T. thermophilus</i> PLHGCEVDsL | AYIEMMQCDDsD...EPV VRVLVNDRVV |
| | <i>T. lanuginosa</i> PLHGCrVDRW | AYVELLRcET ETsSeeEEeEG ..ED...EPF VRVLVNDRVV |
| 25 | <i>M. thermophila</i> TLkGCGaDEr | iYVEkMRCsG GGgGgGGgEG ..rQekdEeM VRVLVNDVRM |
| 30 | Consensus Seq. 11 PLHGCGVDKL | AYVEMMQCEA GG-G-GG-EG --EK---EPL VRVLVNDRVV |
| | | 451 482 |
| | <i>P. involutus</i> (phyA1) | G1CtLAKFVE SqTFARSDga GDFEKCFats a- |
| | <i>P. involutus</i> (phyA2) | G1CaLDKFVE SqAYARSGga GDFEKCLAtt v- |
| | <i>T. pubescens</i> | GvCtLDAFVE SqAYARNDge GDFEKCFat- -- |
| 35 | <i>A. pediades</i> | S1CtLEAFVE SqkYAReDgq GDFEKCFD-- -- |
| | <i>P. lycii</i> | GvCELSAFVE SqTYAReNgq GDFAKCgfvp se |

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| | | | |
|----|-------------------------------------|-------------------------------------|----|
| | <i>A. terreus</i> 9a1 | GRCKrDAFVA GLSFAQAG.. GNWADCF---- | -- |
| | <i>A. terreus</i> cbs | GRCKrDDFVE GLSFARAG.. GNWAECE---- | -- |
| | <i>A. niger</i> var. <i>awamori</i> | GRCtrDsFVr GLSFARSG.. GDWAECSA---- | -- |
| | <i>A. niger</i> T213 | GRCtrDsFVr GLSFARSG.. GDWAECEFA---- | -- |
| 5 | <i>A. niger</i> NRRL3135 | GRCtrDsFVr GLSFARSG.. GDWAECEFA---- | -- |
| | <i>A. fumigatus</i> ATCC13073 | GRCKLNDFVK GLSWARSG.. GNWGECEFS---- | -- |
| | <i>A. fumigatus</i> ATCC32722 | GRCKLNDFVK GLSWARSG.. GNWGECEFS---- | -- |
| | <i>A. fumigatus</i> ATCC58128 | GRCKLNDFVK GLSWARSG.. GNWGECEFS---- | -- |
| | <i>A. fumigatus</i> ATCC26906 | GRCKLNDFVK GLSWARSG.. GNWGECEFS---- | -- |
| 10 | <i>A. fumigatus</i> ATCC32239 | GRCKLKDFVK GLSWARSG.. GNSEQSFS---- | -- |
| | <i>E. nidulans</i> | GRCTLDDWVE GLNFARSG.. GNWktCFTl---- | -- |
| | <i>T. thermophilus</i> | GRCKrDDFVr GLSFARqG.. GNWEGCYAas e- | |
| | <i>T. lanuginosa</i> | GRCRrDEWIK GLTFARqG.. GHWDrCF---- | -- |
| | <i>M. thermophila</i> | GmCtLErFIE SMAFARGN.. GKWDlCFA---- | -- |
| 15 | | | |
| | Consensus Seq. 11 | GRCKLDDFVE GLSFARSG-- GNWAECEFA---- | -- |

20

25

Figure 7

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20 M G V F V V L L S I A T L F G S T S G T
ATGGGCGTGTTTCGTCTGTCTACTGTCCATTGCCACCTTGTTCGGTTCACATCCGGTACC

5 1 ---+-----+-----+-----+-----+-----+-----
60 TACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAAACAAGCCAAGGTGTAGGCCATGG

10 40 A L G P R G N S H S C D T V D G G Y Q C
GCCTTGGGTCCTCGTGGTAATTCTCACTCTTGTGACACTGTTGACGGTGGTTACCAATGT

120 61 ---+-----+-----+-----+-----+-----+-----
CGGAACCCAGGAGCACCATTAAAGAGTGAGAACACTGTGACAACCTGCCACCAATGGTTACA

15 60 F P E I S H L W G T Y S P Y F S L A D E
TTCCCAGAAATTTCTCACTTGTGGGGTACCTACTCTCCATACTTCTCTTTGGCAGACGAA

20 180 121 ---+-----+-----+-----+-----+-----+-----
AAGGGTCTTTAAAGAGTGAACACCCCATGGATGAGAGGTATGAAGAGAAACCGTCTGCTT

80 S A I S P D V P D D C R V T F V Q V L S
TCTGCTATTTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTTCGTTCAAGTTTGTCT

25 240 187 ---+-----+-----+-----+-----+-----+-----
AGACGATAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAGTTCAAAACAGA

30 100 R H G A R Y P T S S A S K A Y S A L I E
AGACACGGTGCTAGATACCCAACCTTCTTCTGCGTCTAAGGCTTACTCTGCTTTGATTGAA

300 241 ---+-----+-----+-----+-----+-----+-----
TCTGTGCCACGATCTATGGGTTGAAGAAGACGCAGATTCCGAATGAGACGAACTAACTT

35 120 A I Q K N A T A F K G K Y A F L K T Y N

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GCTATTCAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTTCTTGAAGACTTACAAC
301 ---+-----+-----+-----+-----+-----+-----
360
CGATAAGTTTTCTTGCGATGACGAAAGTTCCCATTCATGCGAAAGAACTTCTGAATGTTG
5
Y T L G A D D L T P F G E N Q M V N S G
140
TACACTTTGGGTGCTGACGACTTGACTCCATTCCGGTGAAAACCAAATGGTTAACTCTGGT
361 ---+-----+-----+-----+-----+-----+-----
10 420
ATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTTTGGTTTACCAATTGAGACCA
I K F Y R R Y K A L A R K I V P F I R A
160
ATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCATTCATTAGAGCT
15
421 ---+-----+-----+-----+-----+-----+-----
480
TAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTAAGTAATCTCGA
S G S D R V I A S A E K F I E G F Q S A
20
180
TCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTTTCCAATCTGCT
481 ---+-----+-----+-----+-----+-----+-----
540
AGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAACTTCCAAAGGTTAGACGA
25
K L A D P G S Q P H Q A S P V I N V I I
200
AAGTTGGCTGACCCAGGTTCTCAACCACACCAAGCTTCTCCAGTTATTAACGTGATCATT
30
541 ---+-----+-----+-----+-----+-----+-----
600
TTCAACCGACTGGGTCCAAGAGTTGGTGTGGTTCGAAGAGGTCAATAATTGCACTAGTAA
P E G S G Y N N T L D H G T C T A F E D
35 220
CCAGAAGGATCCGGTTACAACAACACTTTGGACCACGGTACTTGTACTGCTTTTGAAGAC

601 ---+-----+-----+-----+-----+-----+-----
660
GGTCTTCCTAGGCCAATGTTGTTGTGAAACCTGGTGCCATGAACATGACGAAAGCTTCTG

5 240 S E L G D D V E A N F T A L F A P A I R
TCTGAATTAGGTGACGACGTTGAAGCTAACTTCACTGCTTTGTTGCTCCAGCTATTAGA

661 ---+-----+-----+-----+-----+-----+-----
720
10 AGACTTAATCCACTGCTGCAACTTCGATTGAAGTGACGAAACAAGCGAGGTCGATAATCT

A R L E A D L P G V T L T D E D V V Y L
260
GCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTTGACTGACGAAGACGTTGTTTACTTG

15 721 ---+-----+-----+-----+-----+-----+-----
780
CGATCTAACCTTCGACTGAACGGTCCACAATGAAACTGACTGCTTCTGCAACAAATGAAC

M D M C P F D T V A R T S D A T E L S P
20 280
ATGGACATGTGTCCATTCGACACTGTGCTAGAACTTCTGACGCTACTGAATTGTCTCCA

781 ---+-----+-----+-----+-----+-----+-----
840
TACCTGTACACAGGTAAGCTGTGACAGCGATCTTGAAGACTGCGATGACTTAACAGAGGT

25 300 F C A L F T H D E W I Q Y D Y L Q S L G
TTCTGTGCTTTGTTCACTCACGACGAATGGATCCAATACGACTACTTGCAAAGCTTGGGT

841 ---+-----+-----+-----+-----+-----+-----
900
30 AAGACACGAAACAAGTGAGTGCTGCTTACCTAGGTTATGCTGATGAACGTTTCGAACCCA

K Y Y G Y G A G N P L G P A Q G V G F A
320
35 AAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTGTTGGTTTCGCT

901 ---+-----+-----+-----+-----+-----+-----
960

Modtag
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TTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCACAACCAAAGCGA
N E L I A R L T H S P V Q D H T S T N H
340
5 AACGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTTCTACTAACCAC
961 ---+-----+-----+-----+-----+-----+-----+-----
1020
TTGCTTAACTAACGATCTAACTGAGTGAGAGGTCAAGTTCTGGTGTGAAGATGATTGGTG
10 T L D S N P A T F P L N A T L Y A D F S
360
ACTTTGGACTCTAACCCAGCTACTTTCCCATTGAACGCTACTTTGTACGCTGACTTCTCT
1021 ---+-----+-----+-----+-----+-----+-----+-----
1080
15 TGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGCGATGAAACATGCGACTGAAGAGA
H D N I M I S I F F A L G L Y N G T K P
380
CACGACAACACTATGATATCTATTTTCTTCGCTTTGGGTTTGTACAACGGTACCAAGCCA
20 1081 ---+-----+-----+-----+-----+-----+-----+-----
1140
GTGCTGTTGTGATACTATAGATAAAAGAAGCGAAACCCAAACATGTTGCCATGGTTCGGT
L S T T S V E S I E E T D G Y S A S W T
25 400
TTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTGCTTCTTGGACT
1141 ---+-----+-----+-----+-----+-----+-----+-----
1200
AACAGATGATGAAGACAACCTTAGATAACTTCTTTGACTGCCAATGAGACGAAGAACCTGA
30 V P F A A R A Y V E M M Q C Q A E K E P
420
GTTCCATTGCTGCTAGAGCTTACGTTGAAATGATGCAATGTCAAGCTGAAAAGGAACCA
1201 ---+-----+-----+-----+-----+-----+-----+-----
35 1260
CAAGGTAAGCGACGATCTCGAATGCAACTTTACTACGTTACAGTTCGACTTTTCCTTGGT

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440 L V R V L V N D R V V P L H G C A V D K
TTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTGCTGTTGACAAG
5 1261 ---+-----+-----+-----+-----+-----+-----
1320
AACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACACGACAACCTGTTTC
10 460 L G R C K R D D F V E G L S F A R S G G
TTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTCGCTAGATCTGGTGGT
1321 ---+-----+-----+-----+-----+-----+-----
1380
AACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGATCTAGACCACCA
15 N W A E C F A * 467
AACTGGGCTGAATGTTTCGCTTAA
1381 ---+-----+-----+ 1410
TTGACCCGACTTACAAAGCGAATT
20

25

30

5

Figure 8

| | | |
|----|-----|---|
| 10 | | M G V F V V L L S I A T L F G S T S G T |
| 20 | | ATGGGCGTGTTTCGTGCTGCTACTGTCCATTGCCACCTTGTTCGGTTCCACATCCGGTACC |
| 15 | 60 | 1 -----+-----+-----+-----+-----+-----+ |
| | | TACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAACAAGCCAAGGTGTAGGCCATGG |
| 40 | | A L G P R G N S H S C D T V D G G Y Q C |
| 20 | | GCCTTGGGTCCTCGTGGTAACTCTCACTCTTGTGACACTGTTGACGGTGGTTACCAATGT |
| | 120 | 61 -----+-----+-----+-----+-----+-----+ |
| | | CGGAACCCAGGAGCACCATTGAGAGTGAGAACACTGTGACAACTGCCACCAATGGTTACA |
| 25 | A | F P E I S H L W G T Y S P F F S L A D E |
| | 60 | TTCCCAGAAATTTCTCACTTGTGGGGTACATACTCTCCATTCTTCTCTTTGGCTGACGAA |
| | 180 | 121 -----+-----+-----+-----+-----+-----+ |
| 30 | | AAGGGTCTTTAAAGAGTGAACACCCCATGTATGAGAGGTAAGAAGAGAAACCGACTGCTT |

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80 S A I S P D V P K G C R V T F V Q V L S

TCTGCTATTCTCCAGACGTTCCAAAGGGTTGTAGAGTTACTTTCGTTCAAGTTTTGTCT

5 181 -----+-----+-----+-----+-----+

240 AGACGATAAAGAGGTCTGCAAGGTTCCCAACATCTCAATGAAAGCAAGTTCAAAACAGA

10 100 R H G A R Y P T S S A S K A Y S A L I E

AGACACGGTGCTAGATACCCAACTTCTTCTGCGTCTAAGGCGTACTCTGCTTTGATTGAA

241 -----+-----+-----+-----+-----+

300 TCTGTGCCACGATCTATGGGTTGAAGAAGACGCAGATTCCGCATGAGACGAACTAACTT

15 120 A I Q K N A T A F K G K Y A F L K T Y N

GCTATTCAAAGAAGCGTACTGCTTTCAAGGGTAAGTACGCTTTCTTGAAGACTTACAAC

301 -----+-----+-----+-----+-----+

20 360 CGATAAGTTTTCTTGCGATGACGAAAGTTCCCATTCATGCGAAAGAACTTCTGAATGTTG

A 140 Y T L G A D D L T P F G E Q Q M V N S G

25 TACACTTTGGGTGCTGACGACTTGACTCCATTCCGGTGAACAACAAATGGTTAACTCTGGT

361 -----+-----+-----+-----+-----+

420 ATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTGTTGTTTACCAATTGAGACCA

30 160 I K F Y R R Y K A L A R K I V P F I R A

ATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCATTCATTAGAGCT

421 -----+-----+-----+-----+-----+

480 TAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTAAGTAATCTCGA

35 180 S G S D R V I A S A E K F I E G F Q S A

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TCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTTTCCAATCTGCT
481 -----+-----+-----+-----+-----+-----+
540
AGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAACCTCCAAAGGTTAGACGA
5
K L A D P G A N P H Q A S P V I N V I I
200
AAGTTGGCTGACCCAGGTGCTAACCCACACCAAGCTTCTCCAGTTATTAACGTTATTATT
541 -----+-----+-----+-----+-----+-----+
10 600
TTCAACCGACTGGGTCCACGATTGGGTGTGGTTCGAAGAGGTCAATAATTGCAATAATAA
P E G A G Y N N T L D H G L C T A F E E
220
15 CCAGAAGGTGCTGGTTACAACAACACTTTGGACCACGGTTTGTGTACTGCTTTCGAAGAA
601 -----+-----+-----+-----+-----+-----+
660
GGTCTTCCACGACCAATGTTGTTGTGAAACCTGGTGCCAAACACATGACGAAAGCTTCTT
20
S E L G D D V E A N F T A V F A P P I R
240
TCTGAATTGGGTGACGACGTTGAAGCTAACTTCACTGCTGTTTTTCGCTCCACCAATTAGA
661 -----+-----+-----+-----+-----+-----+
720
25 AGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGACAAAAGCGAGGTGGTTAATCT
A R L E A H L P G V N L T D E D V V N L
260
GCTAGATTGGAAGCTCACTTGCCAGGTGTTAACTTGACTGACGAAGACGTTGTAACTTG
30 721 -----+-----+-----+-----+-----+-----+
780
CGATCTAACCTTCGAGTGAACGGTCCACAATTGAACTGACTGCTTCTGCAACAATTGAAC
M D M C P F D T V A R T S D A T Q L S P
35 280
ATGGACATGTGTCCATTGACACTGTTGCTAGAACTTCTGACGCTACTCAATTGTCTCCA

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840 781 -----+-----+-----+-----+-----+-----+
TACCTGTACACAGGTAAGCTGTGACAACGATCTTGAAGACTGCGATGAGTTAACAGAGGT
5 300 F C D L F T H D E W I Q Y D Y L Q S L G
TTCTGTGACTTGTTCACTCACGACGAATGGATTCAATACGACTACTTGCAATCTTTGGGT
900 841 -----+-----+-----+-----+-----+-----+
10 AAGACACTGAACAAGTGAGTGCTGCTTACCTAAGTTATGCTGATGAACGTTAGAAACCCA
320 K Y Y G Y G A G N P L G P A Q G V G F V
AAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTGTTGGTTTCGTT
15 901 -----+-----+-----+-----+-----+-----+
960 TTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCACAACCAAAGCAA
20 340 N E L I A R L T H S P V Q D H T S T N H
AACGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTTCTACTAACCAC
1020 961 -----+-----+-----+-----+-----+-----+
TTGCTTAACTAACGATCTAACTGAGTGAGAGGTCAAGTTCTGGTGTGAAGATGATTGGTG
25 360 T L D S N P A T F P L N A T L Y A D F S
ACTTTGGACTCTAACCAGCTACTTTCCCATTTGAACGCTACTTTGTACGCTGACTTCTCT
1021 -----+-----+-----+-----+-----+-----+
30 1080 TGAAACCTGAGATTGGGTCGATGAAAGGGTAACCTGCGATGAAACATGCGACTGAAGAGA
380 H D N T M V S I F F A L G L Y N G T K P
35 CACGACAACACTATGGTTTCTATTTTCTTCGCTTTGGGTTTGTACAACGGTACTAAGCCA
1081 -----+-----+-----+-----+-----+-----+
1140

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GTGCTGTTGTGATACCAAAGATAAAAGAAGCGAAACCCAAACATGTTGCCATGATTCCGGT

400 L S T T S V E S I E E T D G Y S A S W T

5 TTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTGCTTCTTGGACT

1141 -----+-----+-----+-----+-----+-----+-----+
1200

AACAGATGATGAAGACAACCTTAGATAAATTCTTTGACTGCCAATGAGACGAAGAACCTGA

10 V P F A A R A Y V E M M Q C E A E K E P

420

GTTCCATTGCTGCTAGAGCTTACGTTGAAATGATGCAATGTGAAGCTGAAAAGGAACCA

1201 -----+-----+-----+-----+-----+-----+-----+
1260

15 CAAGGTAAGCGACGATCTCGAATGCAACTTTACTACGTTACACTTCGACTTTTCCTTGGT

L V R V L V N D R V V P L H G C G V D K

440

TTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTGGTGTGACAAG

20 1261 -----+-----+-----+-----+-----+-----+-----+
1320

AACCAATCTCAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACACCACAACCTGTTC

L G R C K R D D F V E G L S F A R S G G

25 460

TTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTCGCTAGATCTGGTGGT

1321 -----+-----+-----+-----+-----+-----+-----+
1380

AACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGATCTAGACCACCA

30

N W E E C F A * 467

AACTGGGAAGAATGTTTCGCTTAA

1381 -----+-----+----- 1404

TTGACCCTTCTTACAAAGCGAATT

35

5

10

15

Figure 9

20

M G V F V V L L S I A T L F G S T S G T 20
ATGGGGGTTTTTCGTCGTTCTATTATCTATCGCGACTCTGTTCGGCAGCACATCGGGCACT
1 -----+-----+-----+-----+-----+ 60
TACCCCCAAAAGCAGCAAGATAATAGATAGCGCTGAGACAAGCCGTCGTGTAGCCCGTGA

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A L G P R G N H S K S C D T V D L G Y Q 40
GCGCTGGGCCCCCGTGGAAATCACTCCAAGTCCTGCGATACGGTAGACCTAGGGTACCAG
61 -----+-----+-----+-----+-----+-----+ 120
5 CGCGACCCGGGGGCACCTTTAGTGAGGTTTCAGGACGCTATGCCATCTGGATCCCATGGTC

C S P A T S H L W G T Y S P Y F S L E D 60
TGCTCCCCTGCGACTTCTCATCTATGGGGCACGTA CTGCCATaCTTTTCGCTCGAGGAC
121 -----+-----+-----+-----+-----+-----+ 180
10 ACGAGGGGACGCTGAAGAGTAGATACCCCGtgCATGAGCGGTAtGAAAAGCGAGCTCCTG

E L S V S S K L P K D C R I T L V Q V L 80
GAGCTGTCCGTGTCGAGTAAGCTTCCCAAGGATTGCCGGATCACCTTGGTACAGGTGCTA
181 -----+-----+-----+-----+-----+-----+ 240
15 CTCGACAGGCACAGCTCATTGGAAGGGTTCCTAACGGCCTAGTGGAACCATGTCCACGAT

S R H G A R Y P T S S K S K K Y K K L I 100
TCGCGCCATGGAGCGCGGTACCCAACCAGCTCCAAGAGCAAAAAGTATAAGAAGCTTaTt
241 -----+-----+-----+-----+-----+-----+ 300
20 AGCGCGGTACCTCGCGCCATGGGTTGGTTCGAGGTTCTCGTTTTTCATATCTTTCGAAtAa

T A I Q A N A T D F K G K Y A F L K T Y 120
ACGGCGATCCAGGCCAATGCCACCGACTTCAAGGGCAAGTAcGCCTTTTTGAAGACGTAC
301 -----+-----+-----+-----+-----+-----+ 360
25 TGCCGCTAGGTCCGTTACGGTGGCTGAAGTTCCCGTTCAtgCGGAAAAAATTCTGCATG

N Y T L G A D D L T P F G E Q Q L V N S 140
AACTATACTCTGGGTGCGGATGACCTCACTCCCTTTGGGGAGCAGCAGCTGGTGAACTCG
361 -----+-----+-----+-----+-----+-----+ 420
30 TTGATATGAGACCCACGCCTACTGGAGTGAGGGAAACCCCTCGTCGTCGACCACTTGAGC

G I K F Y Q R Y K A L A R S V V P F I R 160
GGCATCAAGTTCTACCAGAGGTACAAGGCTCTGGCGCGCAGTGTGGTGCCGTTTATTTCGC

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421 -----+-----+-----+-----+-----+ 480
CCGTAGTTCAAGATGGTCTCCATGTTCCGAGACCGCGCGTCACACCACGGCAAATAAGCG
A S G S D R V I A S G E K F I E G F Q Q 180
5 GCCTCAGGCTCGGACCGGGTTATTGCTTCGGGAGAGAAGTTCATCGAGGGGTTCCAGCAG
481 -----+-----+-----+-----+-----+ 540
CGGAGTCCGAGCCTGGCCCAATAACGAAGCCCTCTCTTCAAGTAGCTCCCCAAGGTCGTC
A K L A D P G A T N R A A P A I S V I I 200
10 GCGAAGCTGGCTGATCCTGGCGCGACGAACCGCGCCGCTCCGGCGATTAGTGTGATTATT
541 -----+-----+-----+-----+-----+ 600
CGCTTCGACCGACTAGGACCGCGCTGCTTGGCGCGGCGAGGCCGCTAATCACACTAATAA
P E S E T F N N T L D H G V C T K F E A 220
15 CCGGAGAGCGAGACGTTCAACAATACGCTGGACCACGGTGTGTGCACGAAGTTTGAGGCG
601 -----+-----+-----+-----+-----+ 660
GGCCTCTCGCTCTGCAAGTTGTTATGCGACCTGGTGCCACACACGTGCTTCAAACCTCCGC
S Q L G D E V A A N F T A L F A P D I R 240
20 AGTCAGCTGGGAGATGAGGTTGCGGCCAATTTCACTGCGCTCTTTGCACCCGACATCCGA
661 -----+-----+-----+-----+-----+ 720
TCAGTCGACCCTCTACTCCAACGCCGTTAAAGTGACGCGAGAAACGTGGGCTGTAGGCT
A R L E K H L P G V T L T D E D V V S L 260
25 GCTCGCctCGAGAAGCATCTTCCTGGCGTGACGCTGACAGACGAGGACGTTGTCAGTCTA
721 -----+-----+-----+-----+-----+ 780
CGAGCGgaGCTCTTCGTAGAAGGACCGCACTGCGACTGTCTGCTCCTGCAACAGTCAGAT
M D M C P F D T V A R T S D A S Q L S P 280
30 ATGGACATGTGTcCGTTTGATACGGTAGCGCGCACCAGCGACGCAAGTCAGCTGTCACCG
781 -----+-----+-----+-----+-----+ 840

TACCTGTACACAgGCAAACTATGCCATCGCGCGTGGTCGCTGCGTTTCAGTCGACAGTGGC

F C Q L F T H N E W K K Y D Y L Q S L G 300

TTCTGTCAACTCTTCACTCACAATGAGTGGAGAAGTACgACTACCTTCAGTCCTTGGGC

5 841 -----+-----+-----+-----+-----+-----+ 900

AAGACAGTTGAGAAGTGAGTGTACTCACCTTCTTCATGcTGATGGAAGTCAGGAACCCG

K Y Y G Y G A G N P L G P A Q G I G F T 320

AAGTACTACGGCTACGGCGCAGGCAACCCTCTGGGACCGGCTCAGGGGATAGGGTTCCAC

10 901 -----+-----+-----+-----+-----+-----+ 960

TTCATGATGCCGATGCCGCGTCCGTTGGGAGACCCTGGCCGAGTCCCCTATCCCAAGTGG

N E L I A R L T R S P V Q D H T S T N S

340

15 AACGAGCTGATTGCCCCGTTGACgCGTTCCGCCAGTGCAGGACCACACCAGCACTAACTCG

961 -----+-----+-----+-----+-----+-----+ 1020

TTGCTCGACTAACGGGCCAACTGcGCAAGCGGTACGTCCTGGTGTGGTCGTGATTGAGC

T L V S N P A T F P L N A T M Y V D F S

20 360

ACTCTAGTCTCCAACCCGGCCACCTTCCC GTTGAACGCTACCATGTACGTCGACTTTTCA

1021 -----+-----+-----+-----+-----+-----+ 1080

25 TGAGATCAGAGGTTGGGCCGGTGGAAAGGGCAACTTGCGATGGTACATGCAGCTGAAAAGT

H D N S M V S I F F A L G L Y N G T E P

380

CACGACAACAGCATGGTTTCCATCTTCTTGCATTGGGCCTGTACAACGGCACTGAACCC

30 1081 -----+-----+-----+-----+-----+-----+ 1140

GTGCTGTTGTCGTACCAAAGGTAGAAGAAACGTAACCCGGACATGTTGCCGTGACTTGGG

L S R T S V E S A K E L D G Y S A S W V

35 400

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TTGTCCCGGACCTCGGTGGAAAGCGCCAAGGAATTGGATGGGTATTCTGCATCCTGGGTG
1141 -----+-----+-----+-----+-----+
1200
AACAGGGCCTGGAGCCACCTTTCGCGGTTCTTAACCTACCCATAAGACGTAGGACCCAC
5
V P F G A R A Y F E T M Q C K S E K E P
420
GTGCCTTTCGGCGCGCGAGCCTACTTTCGAGACGATGCAATGCAAGTCGGAAAAGGAGCCT
1201 -----+-----+-----+-----+-----+
10 1260
CACGGAAAGCCGCGCGCTCGGATGAAGCTCTGCTACGTTACGTTACGCCCTTTTCCTCGGA
L V R A L I N D R V V P L H G C D V D K
440
CTTGTTTCGCGCTTTGATTAATGACCGGGTTGTGCCACTGCATGGCTGCGATGTGGACAAG
15
1261 -----+-----+-----+-----+-----+
1320
GAACAAGCGCGAAACTAATTACTGGCCCAACACGGTGACGTACCGACGCTACACCTGTTC
20
L G R C K L N D F V K G L S W A R S G G
460
CTGGGGCGATGCAAGCTGAATGACTTTGTCAAGGGATTGAGTTGGGCCAGATCTGGGGGC
1321 -----+-----+-----+-----+-----+
1380
25
GACCCCGCTACGTTGACTTACTGAAACAGTTCCTTAACCTCAACCCGGTCTAGACCCCCG
N W G E C F S * 467
AACTGGGGAGAGTGCTTTAGTTGA
1381 -----+-----+----- 1404
30
TTGACCCCTCTCACGAAATCAACT

```

Figure 10

CP-1

5 Eco RI M G V F V V L L S I A T L F G S T
TATATGAATTCATGGGCGTGTTCGTCTGCTACTGTCCATTGCCACCTTGTTCCGGTTCCA
1 -----+-----+-----+-----+-----+-----+ 60
ATATACTTAAGTACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAACAAGCCAAGGT

10 S G T A L G P R G N S H S C D T V D G G
CATCCGGTACCGCCTTGGGTCCTCGTGGTAATTCTCACTCTTGTGACACTGTTGACGGTG
61 -----+-----+-----+-----+-----+-----+ 120
GTAGGCCATGGCGGAACCCAGGAGCACCATTAAAGAGTGAGAACACTGTGACAACCTGCCAC

CP-2
15 CP-3
Y Q C F P E I S H L W G Q Y S P Y F S L
GTTACCAATGTTTCCCAGAAATTTCTCACTTGTGGGGTCAATACTCTCCATACTTCTCTT
121 -----+-----+-----+-----+-----+-----+ 180
CAATGGTTACAAAGGGTCTTTAAAGAGTGAACACCCCAAGTTATGAGAGGTATGAAGAGAA

20 E D E S A I S P D V P D D C R V T F V Q
TGGAAGACGAATCTGCTATTTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTCTGTTT
181 -----+-----+-----+-----+-----+-----+ 240
ACCTTCTGCTTAGACGATAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAG

25 CP-4.7
 CP-5.7
V L S R H G A R Y P T D S K G K K Y S A
AAGTTTGTCTAGACACGGTGCTAGATACCCAAGTgacTCTAAGggtAAGaagTACTCTG
241 -----+-----+-----+-----+-----+-----+ 300
30 TTCAAAACAGATCTGTGCCACGATCTATGGGTTGActgAGATTCCAATTCttcATGAGAC

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L I E A I Q K N A T A F K G K Y A F L K
CTTTGATTGAAGCTATTCAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTTCTTGA
301 -----+-----+-----+-----+-----+ 360
5 GAAACTAACTTCGATAAGTTTTCTTGCGATGACGAAAGTTCCCATTTCATGCGAAAGAAGT
CP-6
CP-7
T Y N Y T L G A D D L T P F G E N Q M V
AGACTTACAACCTACACTTTGGGTGCTGACGACTTGACTCCATTCCGGTGAAAACCAAATGG
10 361 -----+-----+-----+-----+-----+ 420
TCTGAATGTTGATGTGAACCCACGACTGCTGAACTGAGGTAAGCCACTTTTGGTTTACC
N S G I K F Y R R Y K A L A R K I V P F
TTAACTCTGGTATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCAT
15 421 -----+-----+-----+-----+-----+ 480
AATTGAGACCATAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTA
CP-8.7
CP-9
I R A S G S S R V I A S A E K F I E G F
20 TCATTAGAGCTTCTGGTTCTtctAGAGTTATTGCTTCTGCTGAAAAGTTTCATTGAAGGTT
481 -----+-----+-----+-----+-----+ 540
AGTAATCTCGAAGACCAAGAagaTCTCAATAACGAAGACGACTTTTCAAGTAACTTCCAA
Q S A K L A D P G S Q P H Q A S P V I D
25 TCCAATCTGCTAAGTTGGCTGACCCAGGTTCTCAACCACACCAAGCTTCTCCAGTTATTG
541 -----+-----+-----+-----+-----+ 600
AGGTTAGACGATTCAACCGACTGGGTCCAAGAGTTGGTGTGGTTCTGAAGAGGTCAATAAC
CP-10.7
CP-11.7
30 V I I S E A S S Y N N T L D P G T C T A
ACGTTATTATTtctGAcgctTCTtctTACAACAACACTTTGGACccaGGTACTTGTACTG
601 -----+-----+-----+-----+-----+ 660

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TGCAATAATAAagaCTgcgaAGGagaATGTTGTTGTGAAACCTGggtCCATGAACATGAC

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F E D S E L A D T V E A N F T A L F A P

CTTTCGAAGACTCTGAATTGgctGACactGTTGAAGCTAACTTCACTGCTTTGTTTCGCTC

661 -----+-----+-----+-----+-----+ 720

GAAAGCTTCTGAGACTTAACcgaCTGtgaCAACTTCGATTGAAGTGACGAAACAAGCGAG

5

CP-12.7

A I R A R L E A D L P G V T L T D T E V

CAGCTATTAGAGCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTTGACTGACactgaaG

721 -----+-----+-----+-----+-----+ 780

10

GTCGATAATCTCGATCTAACCTTCGACTGAACGGTCCACAATGAACTGACTGtgacttC

CP-13.7

T Y L M D M C S F E T V A R T S D A T E

TTactTACTTGATGGACATGTGTtctTTCGAAACTGTTGCTAGAACTTCTGACGCTACTG

15

781 -----+-----+-----+-----+-----+ 840

AatgaATGAACTACCTGTACACAagaAAGCTTTGACAACGATCTTGAAGACTGCGATGAC

L S P F C A L F T H D E W R H Y D Y L Q

AATTGTCTCCATTCTGTGCTTTGTTCACTCACGACGAATGGAGAcactACGACTACTTGC

20

841 -----+-----+-----+-----+-----+ 900

TTAACAGAGGTAAGACACGAAACAAGTGAGTGCTGCTTACCTCTgtgATGCTGATGAACG

CP-14.7

CP-15.7

S L K K Y Y G H G A G N P L G P T Q G V

25

AATCTTTGaagAAGTACTACGGTcacGGTGCTGGTAACCCATTGGGTCCAactCAAGGTG

901 -----+-----+-----+-----+-----+ 960

TTAGAAACTtctTTCATGATGCCAgtgCCACGACCATTGGGTAACCCAGGTtgaGTTCCAC

G F A N E L I A R L T R S P V Q D H T S

30

TTGGTTTCGCTAACGAATTGATTGCTAGATTGACTAGATCTCCAGTTCAAGACCACACTT

961 -----+-----+-----+-----+-----+ 1020

AACCAAAGCGATTGCTTAACTAACGATCTAACTGATCTAGAGGTCAAGTTCTGGTGTGAA

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CP-16

CP-17.7

T N H T L D S N P A T F P L N A T L Y A
CTACTAACCACACTTTGGACTCTAACCAGCTACTTTCCCATTTGAACGCTACTTTGTACG
5 1021 -----+-----+-----+-----+-----+-----+
1080
GATGATTGGTGTGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGCGATGAAACATGC
D F S H D N G I I S I F F A L G L Y N G
10 CTGACTTCTCTCAGACAACggtattATTTCTATTTTCTTCGCTTTGGGTTTGTACAACG
1081 -----+-----+-----+-----+-----+-----+
1140
GACTGAAGAGAGTGCTGTTGccataaTAAAGATAAAAGAAGCGAAACCCAAACATGTTGC

CP-18.7

CP-19.7

15 T A P L S T T S V E S I E E T D G Y S S
GTACTGCTCCATTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTt
1141 -----+-----+-----+-----+-----+-----+
1200
20 CATGACGAGGTAACAGATGATGAAGACAACCTTAGATAACTTCTTTGACTGCCAATGAGAA
A W T V P F A S R A Y V E M M Q C Q A E
ctgctTGGACTGTTCCATTTCgttctTAGAGCTTACGTTGAAATGATGCAATGTCAAGCTG
1201 -----+-----+-----+-----+-----+-----+
25 1260
gacgaACCTGACAAGGTAAGcgaagaTCTCGAATGCAACTTTACTACGTTACAGTTTCGAC

CP-20

CP-21

K E P L V R V L V N D R V V P L H G C A
30 AAAAGGAACCATTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG
1261 -----+-----+-----+-----+-----+-----+
1320
TTTTCTTGGTAAACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACAC

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V D K L G R C K R D D F V E G L S F A R

CTGTTGACAAGTTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTTCGCTA

1321 -----+-----+-----+-----+-----+-----+
1380

5 GACAACTGTTCAACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGAT

S G G N W A E C F A * Eco RI CP-22

GATCTGGTGGTAACTGGGCTGAATGTTTCGCTTAAGAATTCATATA

1381 -----+-----+-----+-----+-----+----- 1426

10 CTAGACCACCATTGACCCGACTTACAAAGCGAATTCTTAAGTATAT

Figure 11

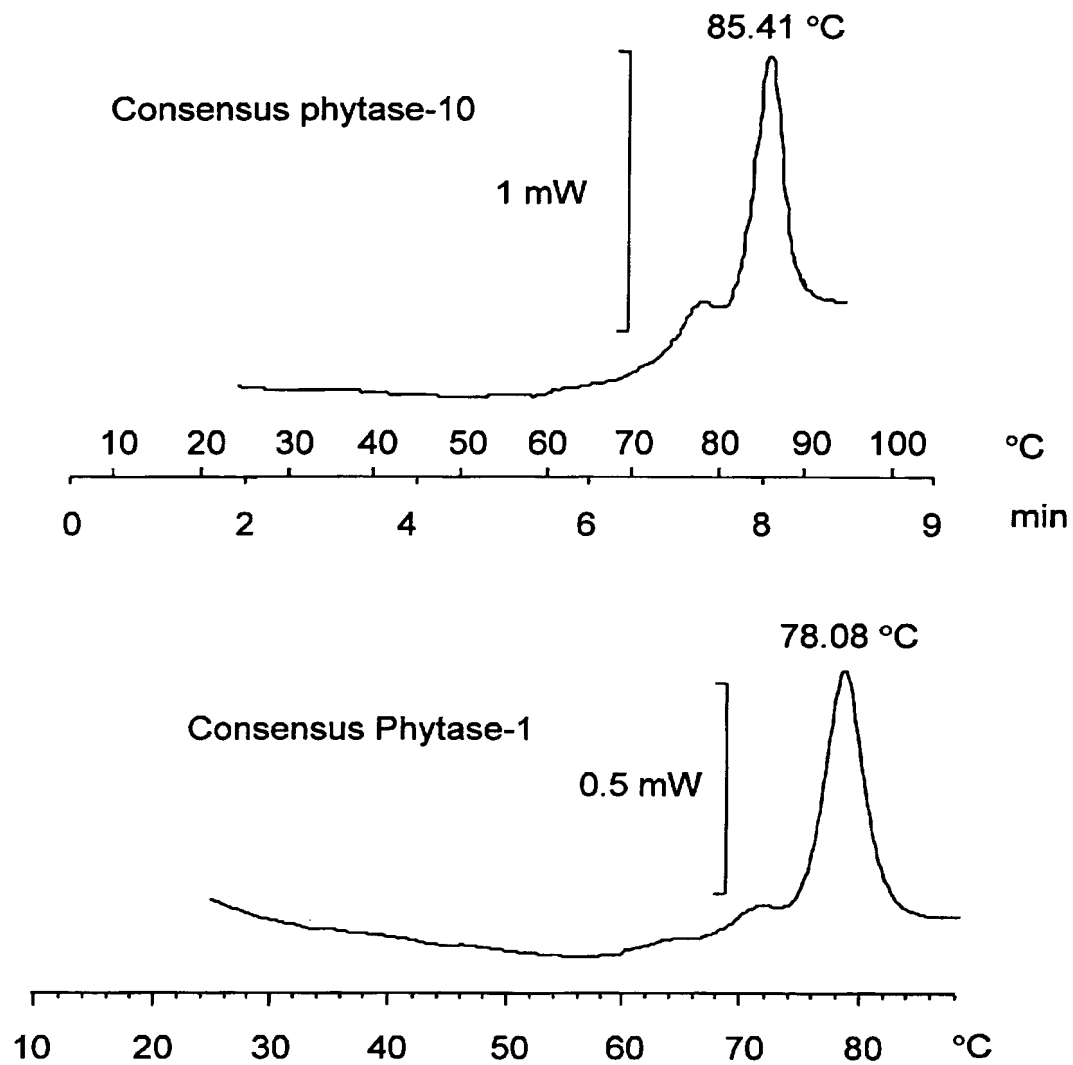


Figure 12

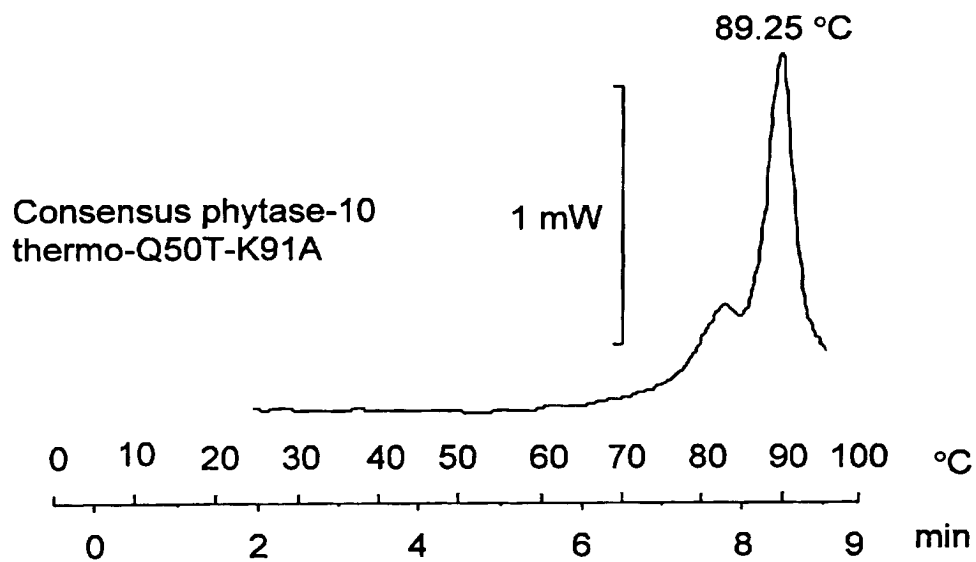
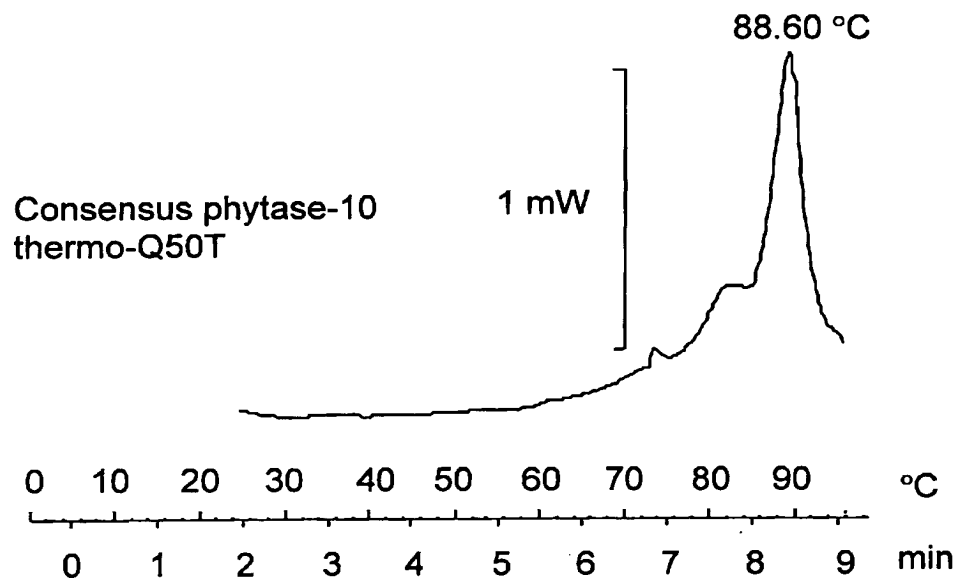


Figure 13

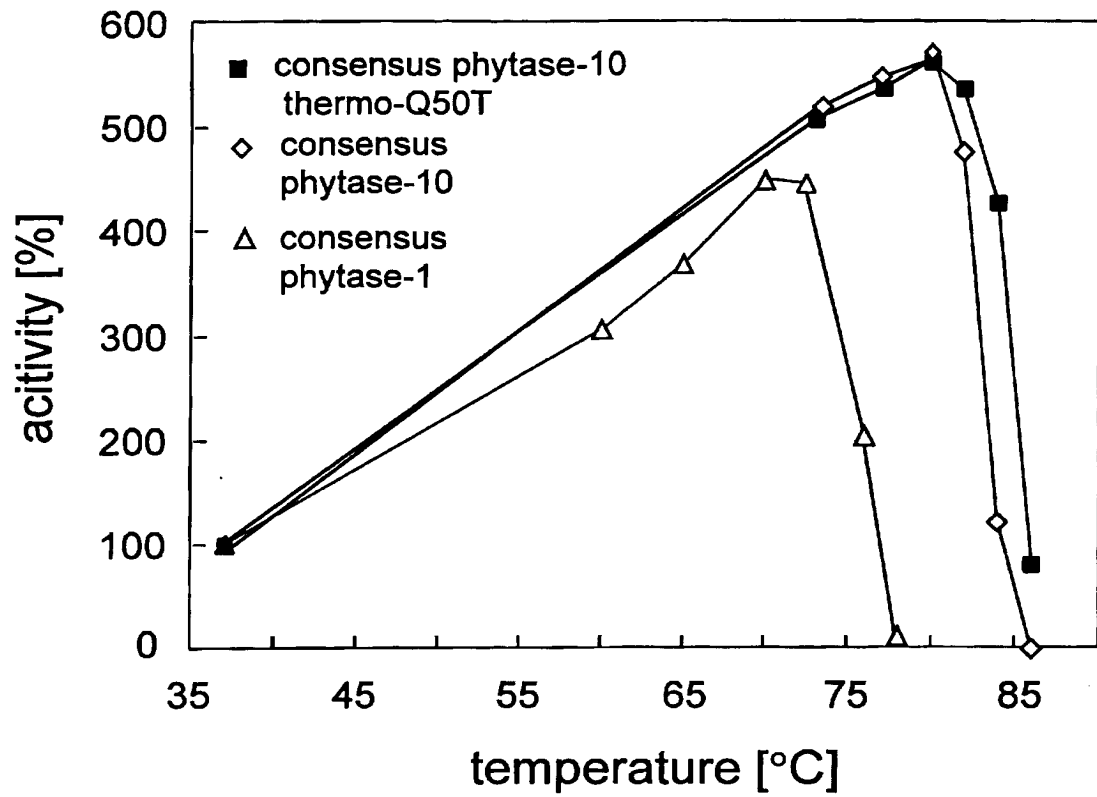


Figure 14

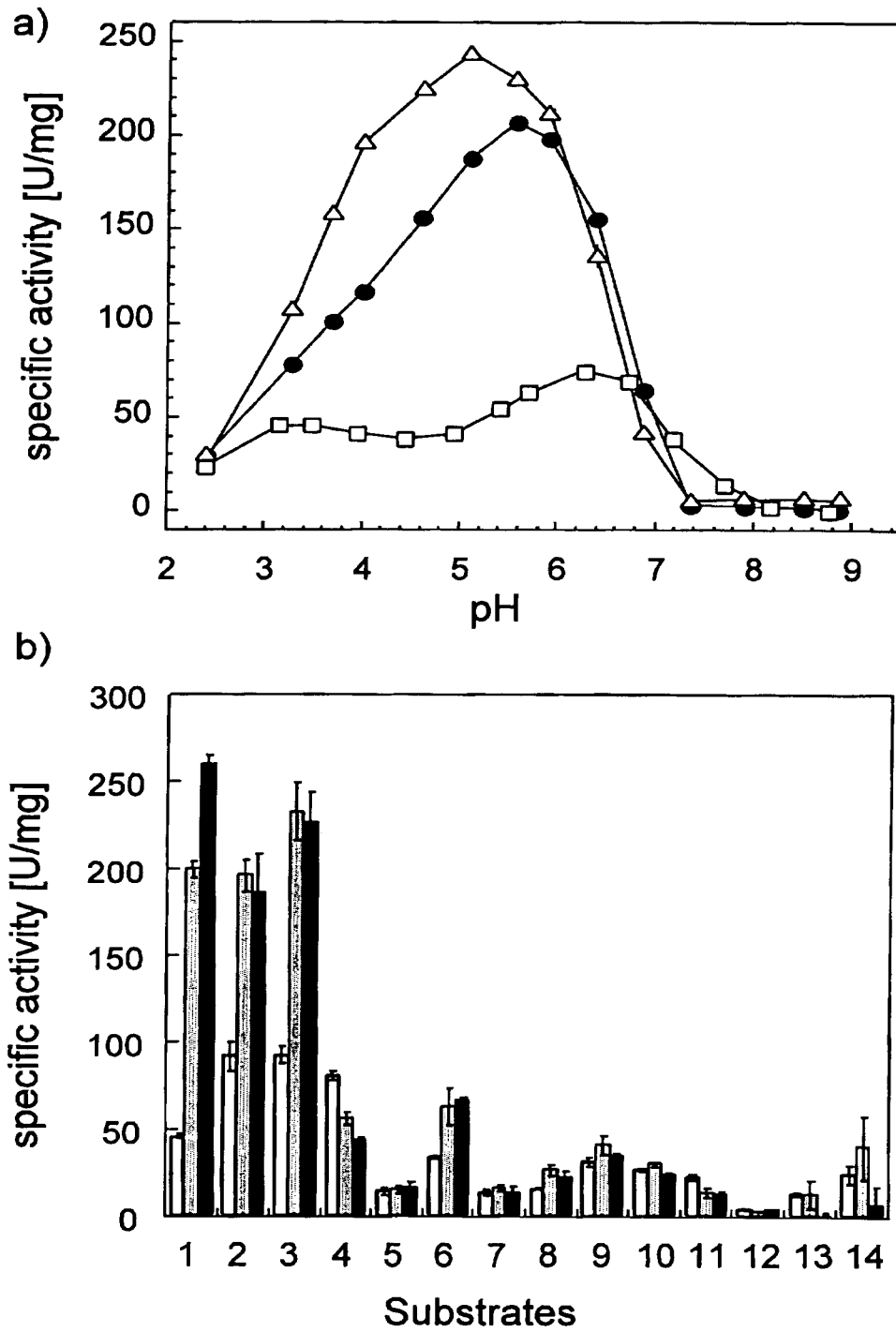


Figure 15

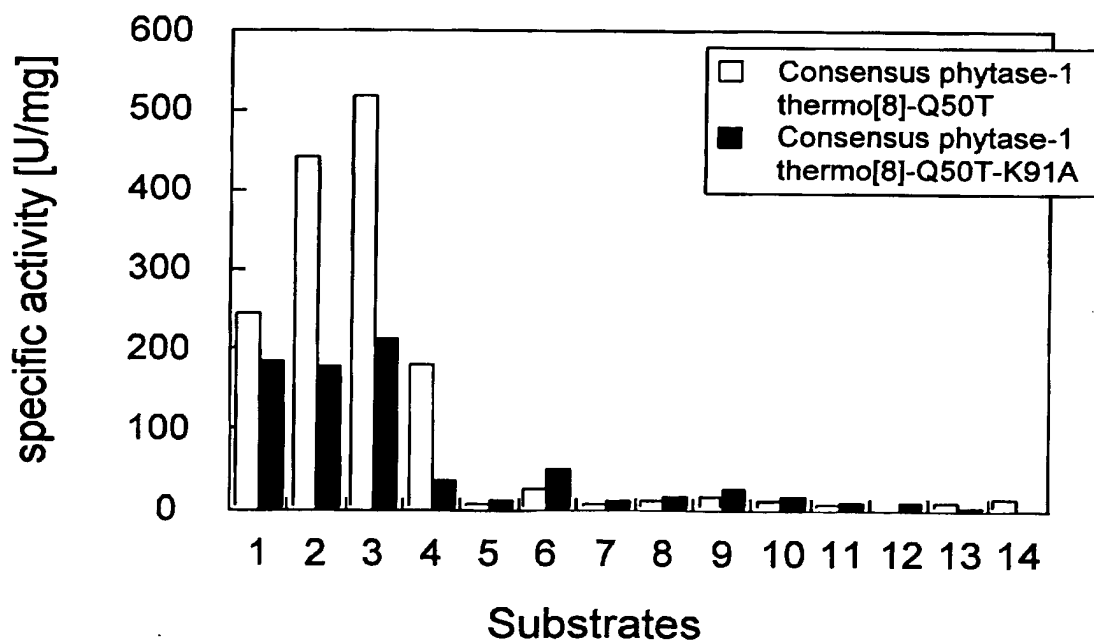
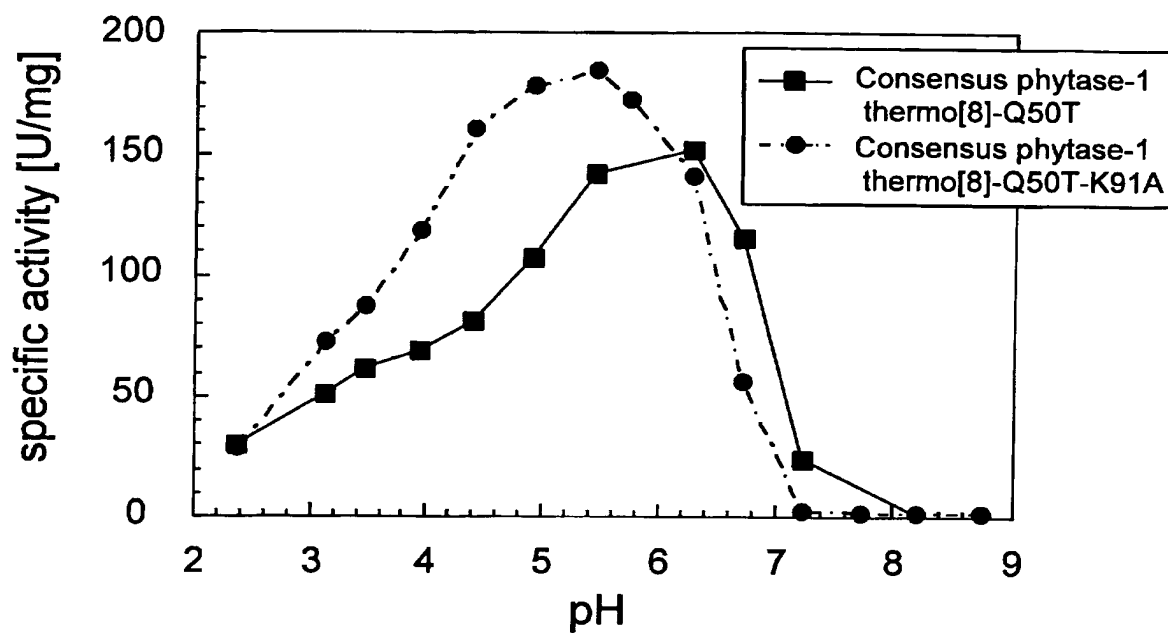


Figure 16

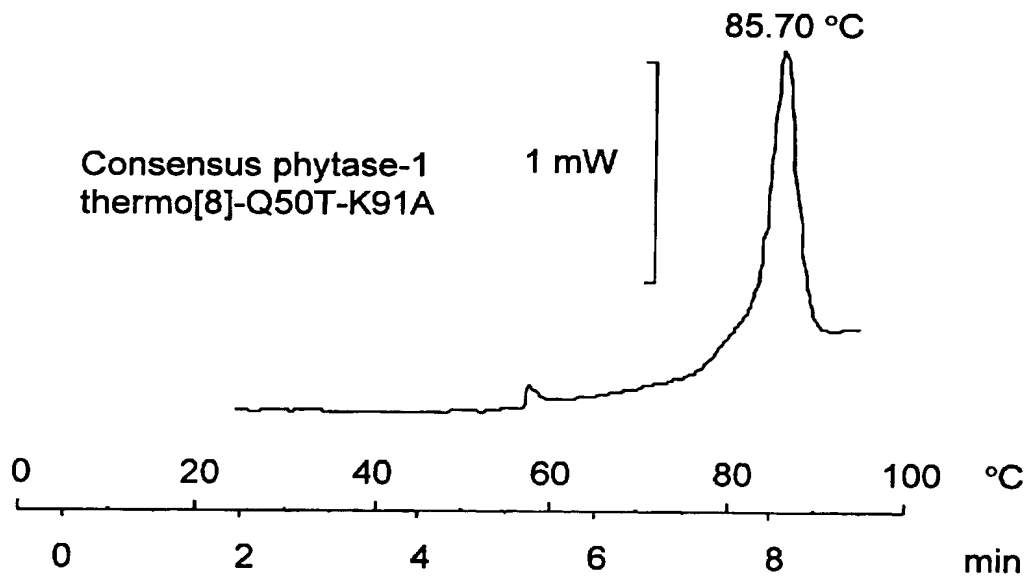
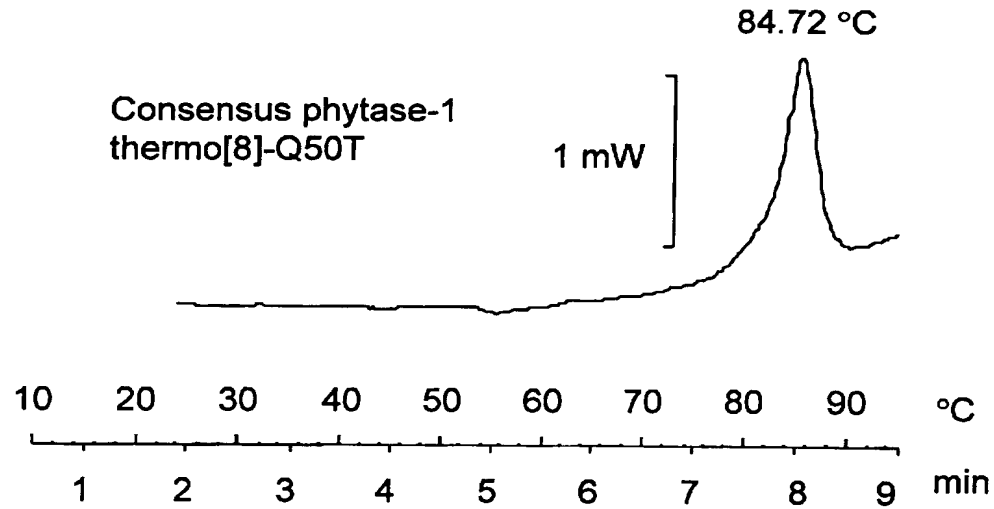
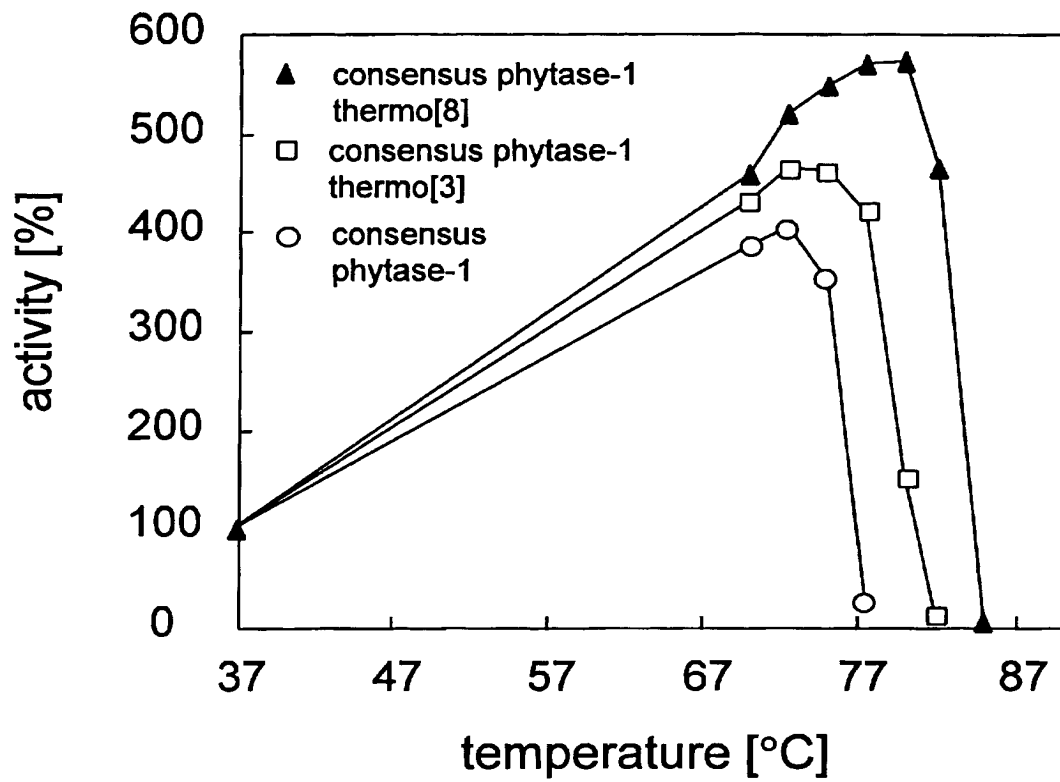
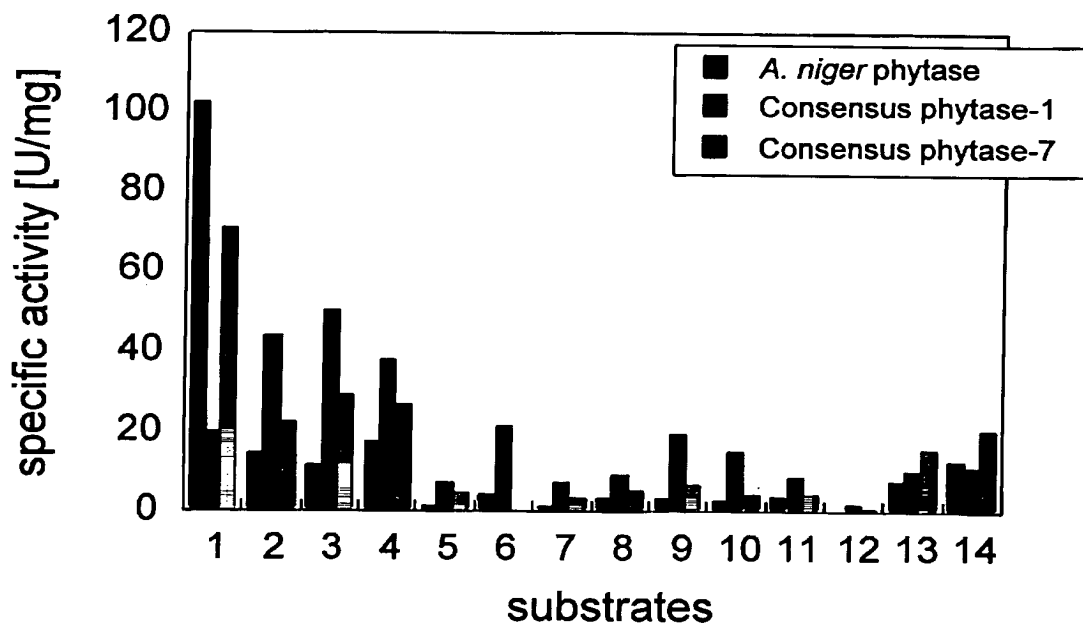
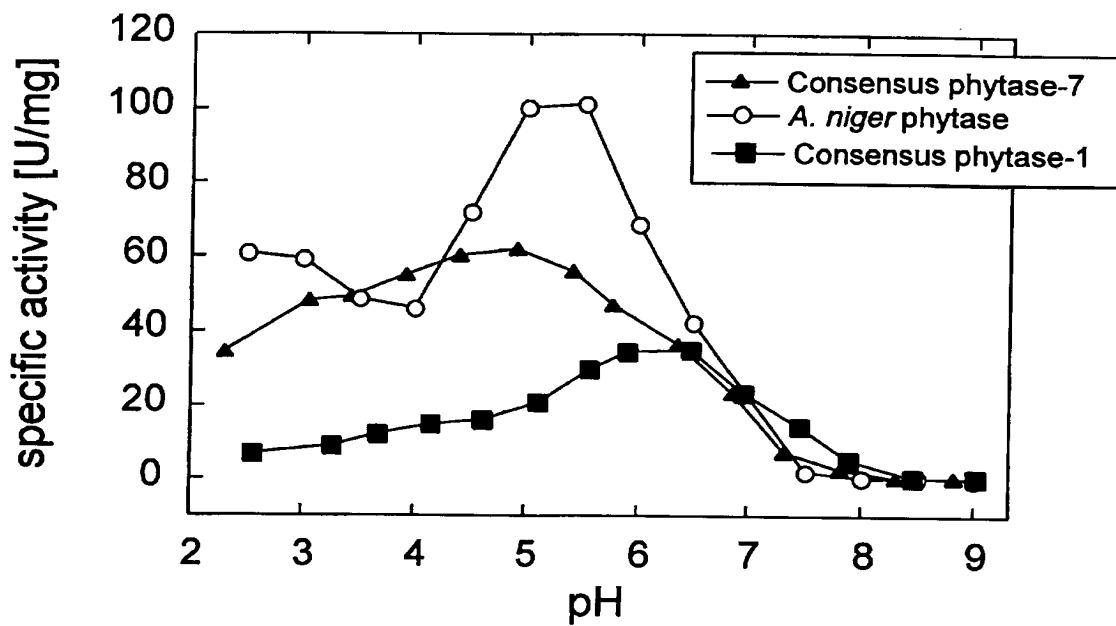


Figure 17



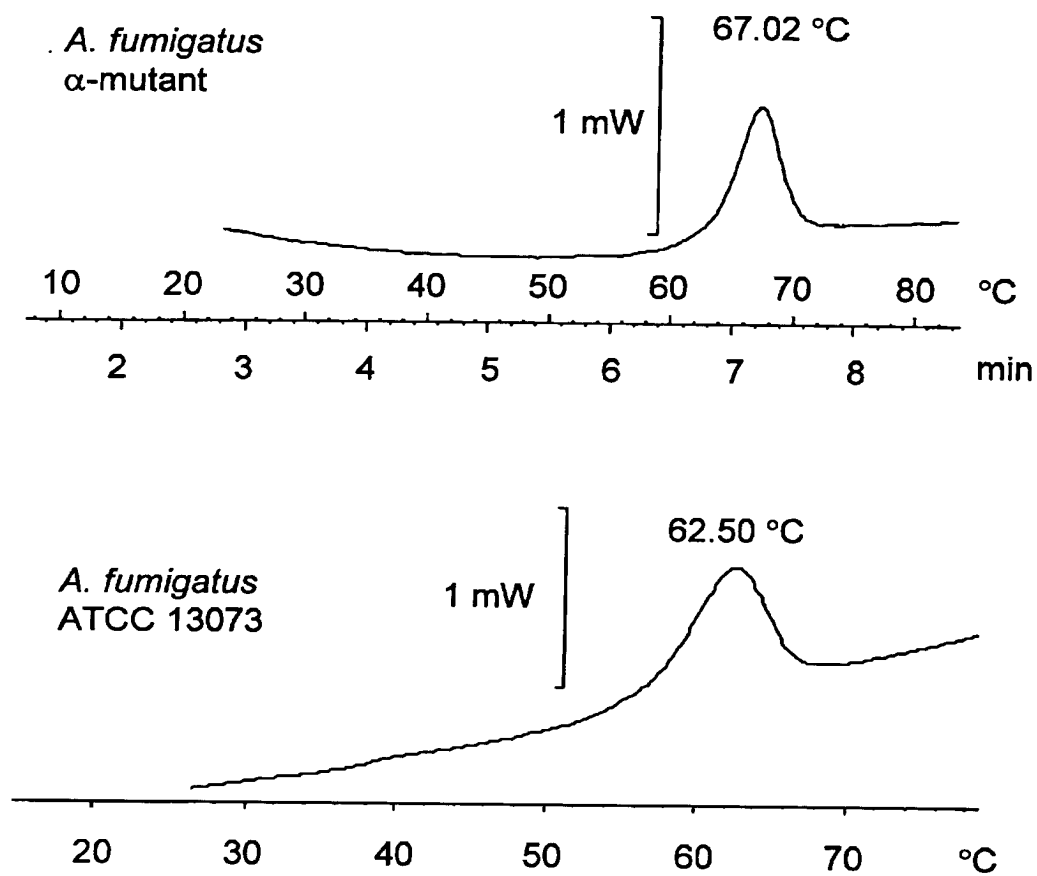
Modtaget PD
22 JAN. 1999

Figure 18



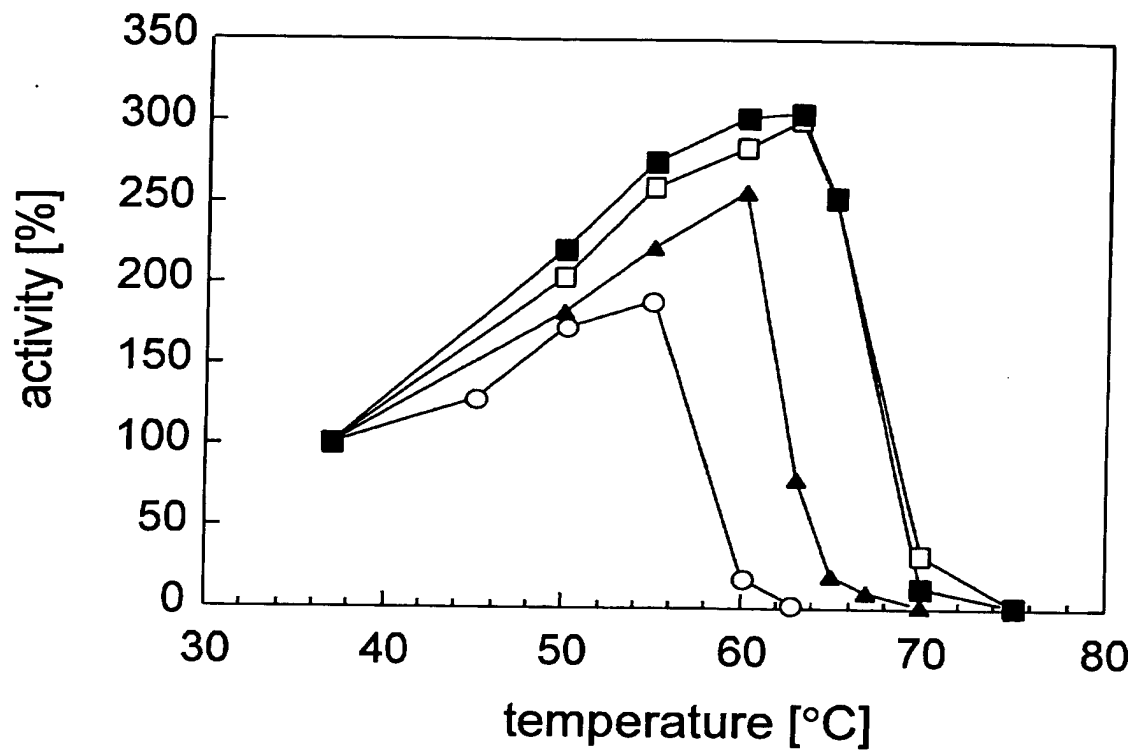
22 JAN. 1999

Figure 19



Modtaget PD
22 JAN. 1959

Figure 20



Modtaget PD
22 JAN. 1993

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Figure 21

1 MGVFVLLSI ATLFGSTSGT ALGPRGNHS CDTVGGYQC FPEISSNWSP
51 YSPYFSLADE SAISPDVPKG CRVTFVQVLQ RHGAREFTSG AATRISALIE
101 AIQKNATAFK GKYAFLKTYN YTLGADDLVP FGANQSSQAG IKFYRRYKAL
5 151 ARKIVPFIRA SGSDRVIDSA TNWIEGFQSA KLADPGANPH QASPVINVII
201 PEGAGYNNTL DHGLCTAFEE SELGDDVEAN FTAVFAPPIR ARLEAHLPGV
251 NLTDEDVVNL MDMCPFDIVA RTSDATELSP FCDLFTHDEW IQDYDLGDL
301 KYYGTGAGNP LGPAQGVGVFV NELIARLTHS PVQDHTSTNH TLDSNPATFP
351 LNATLYADFS HDNTMVAIFF ALGLYNGTKP LSTTSVESIE ETDGYSASWL
10 401 VPFSARMYVE MMQCEAEKEP LVRVLVNDRV VPLHGCGVDK LGRCKRDDFV
451 EGLSFARSGG NWEECFA

MOU--
22 JAN. 1993

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Abstract

This invention relates to a new improved consensus phytase by introduction of
5 additional phytase sequences into the sequence alignment and the method of
the introduction process. Furthermore, the invention relates to the transfer of
stabilizing amino acid exchanges found by the new method into homologous
proteins. Furthermore, the invention relates to the replacement of a whole
active site of a phytase. It also relates to the corresponding DNA sequences
10 and its generation, methods to produce such phytases and the use thereof.
